Diversified natural products in *Rhododendron formosanum* reveal allelochemical and pharmaceutical properties

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China Medical University

Plant Natural Products Play Important Roles in Plant Ecosystem

- **Dominance of vegetation**
- **Climax species**
- **Mechanism of plant succession**
- **Plant productivity**
- **Regulation of biodiversity**

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Tables: 4, Figs : 11

Allelopathic potential of *Rhododendron formosanum* Hemsl in Taiwan

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China Medical University

Rhododendron formosanum

- Endemic specie in Taiwan
- Small trees to 10 m tall
- 600~2400 m asl.
- The soil is acidic (pH 4.0)
- Nitrogen availability is limited

China Medical University

Dasyueshan site (24°14'6.49"N, 120°57'7.29"E at 1911 m asl.; annual rainfall: 2000–3000 mm)

Figure 1. An almost lacking understory plant on the floor of *R. formosanum* (A) as compared to many understory plants on the floor of adjacent woody plant, *C. cuspidata* var. *carlesii* (B), at the Sun Link Sea study site in April, 2008.

Table 1. Comparison of floristic composition and relative coverage of each understory species found on the floors of *C. cuspidata* var. *carlesii* and *R. formosanum* in the Sun Link Sea site

Species	Coverage (%) / Species	
	C. cuspidata var. *carlesii*	*R. formosanum*
Acrophorus stipellatus T. Moore	13.1	-
Arachniodes rhomboides (Wall. *ex* Mett.) Ching	18.1	-
Ardisia crenata Sims	1.1	-
Callicarpa formosana Folfe	6.8	-
Diplazium kawakamii Hayata	1.5	-
Elatostema lineolatum Wight var. *majus* Wedd.	1.2	-
Eurya crenatifolia (Yamam.) Kobuski	1.6	-
Hemiboea bicornuta (Hayata) Ohwi	2.0	-
Hydrangea angustipetala Hayata	0.5	-
Monachosorum henryi Chist	31.8	-
Nanocnide japonica Blume	0.2	-
Pileostegia viburnoides Hook. f. & Thomson	4.2	-
Plagiogyria japonica Nakai	4.7	-
Rubus formosensis Kuntze	0.3	-
Smilax sieboldii Miq.	0.5	-
Stauntonia obovatifoliola Hayata	0.1	-
Strobilanthes formosanus S. Moore	13.2	-
Symplocos caudata Wall. *ex* G. Don	4.5	-
Vandenboschia auriculata (Blume) Copel.	0.5	-
Barthea barthei (Hance) Krass	-	0.4
Cinnamomum subavenium Miq.	-	0.0
Clevera japonica Thunb. *Damnacanthus*	-	1.0
angustifolius Hayata *Dendropanax dentiger*	-	3.5
(Harms *ex* Diels) Merr. *Machilus thunbergii*	-	0.1
Siebold & Zucc.	-	0.3
Myrsine stolonifera (Koidz.) Walker	-	0.6
Neolitsea acuminatissima (Hayata) Kaneh. & Sasaki	-	0.5
Plagiogyria dunnii Copel.	-	3.6
Rhododendron formosanum Hemsl.	-	1.6
Smilax lanceifolia Roxb.	-	1.7
Shortia rotundifolia (Maxim.) Makino	-	0.2
Average total coverage (%)	105.7	13.4
Relative coverage of understory (%)	100.0	12.7
Number of species	19	12

-: Absent

Figure 2. The leaching apparatus system: Upper part is a plastic tray, 55×40×15 cm with numerous needle-sized holes (2 cm between holes) was filled with tap water, which dripped through the holes to make an artificial raindrop, middle part is the same as upper tray, was filled with chopped plant material/organic matter to receive the artificial raindrop from upper tray, lower part is a plastic tray, like that of upper and middle tray, without holes to receive plant leachate from middle tray. The pump re-circulated the water from lower tray to upper tray

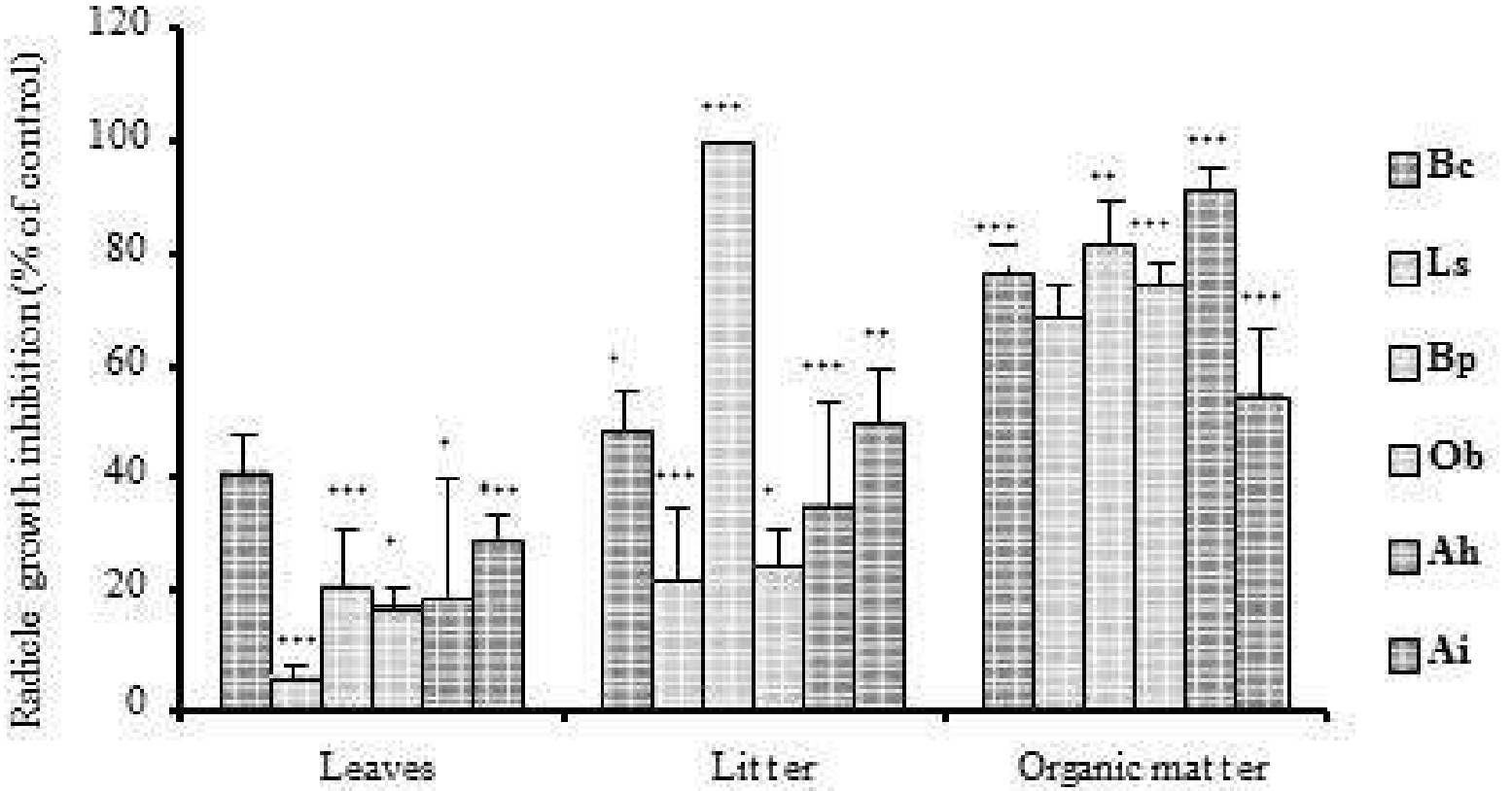

Figure 4. Effects of aqueous leachates of *Rhododendron* leaves, litter, and organic matter on the radicle growth of six bioassay species. Levels of statistical significance are expressed by asterisk: * <0.05, ** <0.01, *** <0.001 . The abbreviations of species names are: *Ageratum houstonianum* (Ah), *Amaranthus inamoenus* (Ai), *Brassica chinensis* (Bc), *Bidens pilosa* (Bp), *Lactuca sativa* (Ls) and *Ocimum basilicum* (Ob)

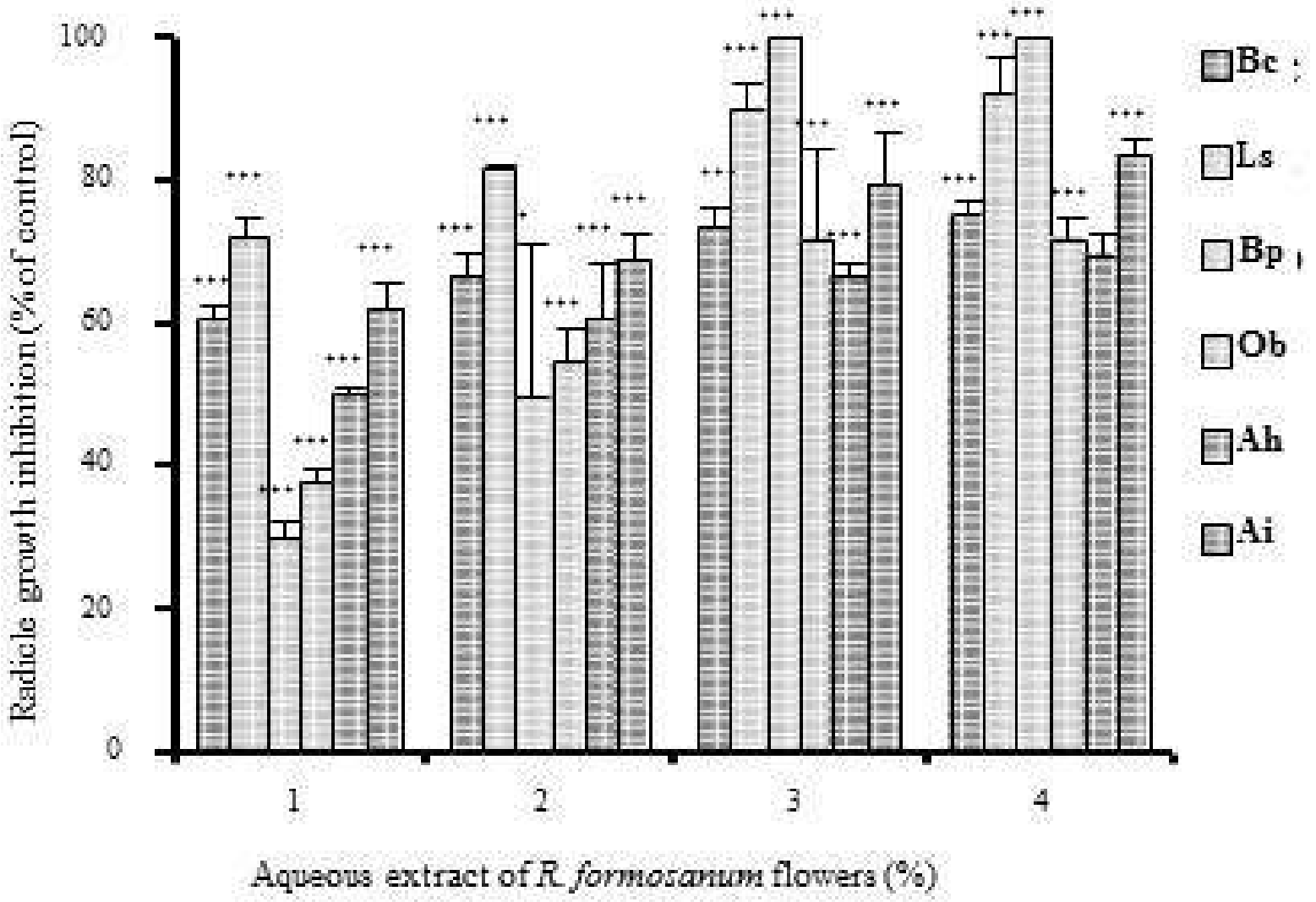

Figure 5. Effects of aqueous extracts concentration of *R. formosanum* flowers on radicle growth of six bioassay species. The abbreviations of legends see Figure 4.

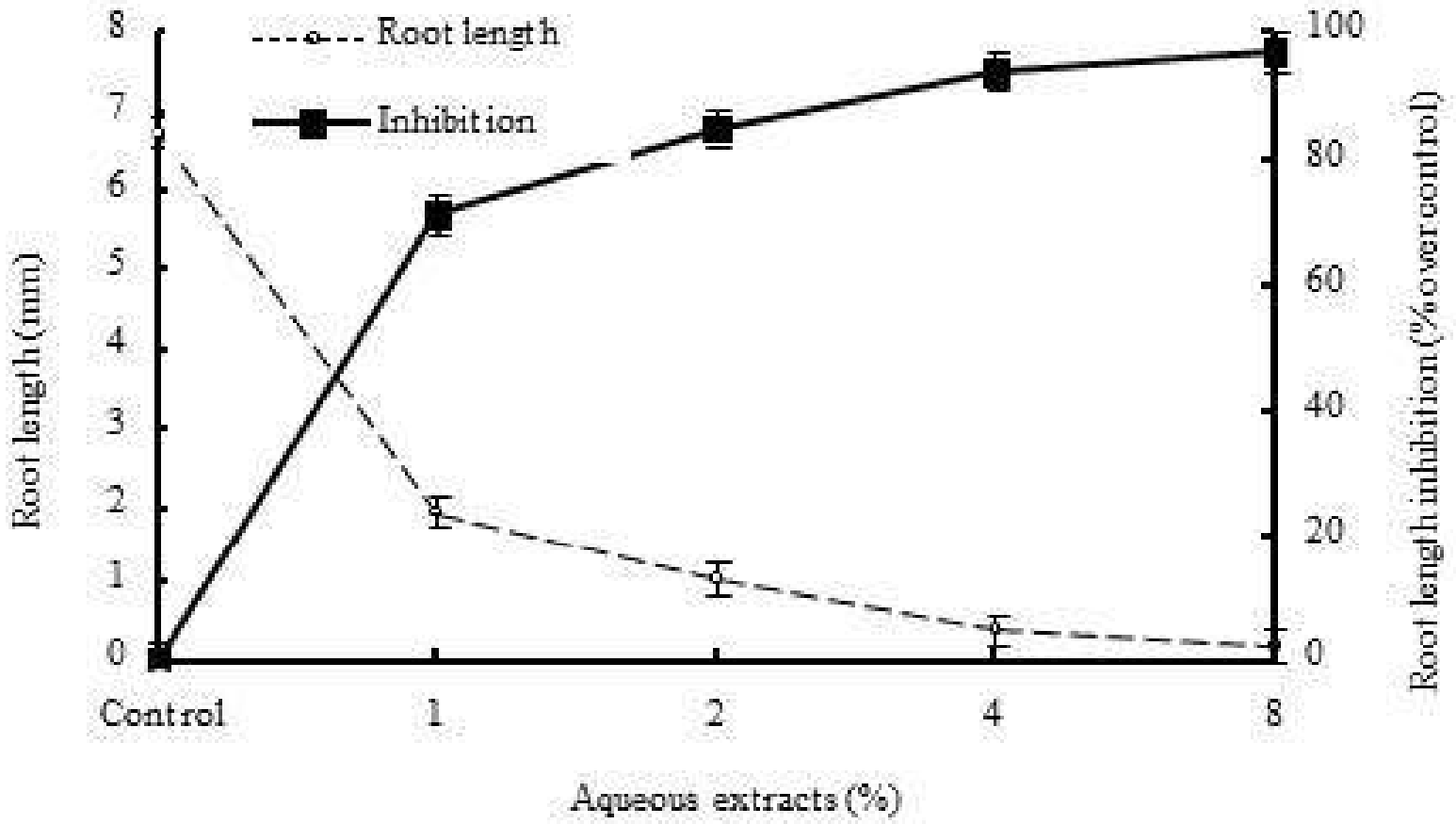

Figure 9. The inhibitory effect of aqueous extract, 0, 1, 2, 4 and 8 % of *R. formosanum* leaves on root length of root initiation of *B. mutica*. The inhibition was significantly different from distilled water control at 5 % level using Student's *t*-test.

Table 4. Phytotoxins isolated from the leaves of *Rhododendron formosanum* in Taiwan*

Compound	Rf values in paper chromatography (2 % HOAc)			HPLC	
	Authentic standard	Isolated from *Rhododendron* leaves	Relatively quantitative comparison**	Compound present	Peak number
p-Hydroxybenzoic acid	0.63	0.66	+	+	(5)***
Chlorogenic acid	0.67			+	(4)
trans p-Coumaric acid	0.43			+	(10)
cis Ferulic acid	0.64	0.68	+++		
Methyl ferulate	0.61	0.61	+	+	(13)
Syringic acid		0.56	++		
Vanillic acid	0.56	0.60	++	+	(6)
Coumarin	0.73			+	(12)
Protocatechuic acid	0.63	0.56	+	+	(2)
(-)-Catechin	0.46	0.47	++	+	(3)

* The identification is primarily based on PC and HPLC.

** The amount of compound is based on the intensity of spot on the PC, showing +++ > ++ > +.

*** Data in parenthesis indicate the number of compound as compared to the authentic peak shown in Figure 11, the peak intensity that is larger than 200 mv in HPLC chromatogram is consider as +.

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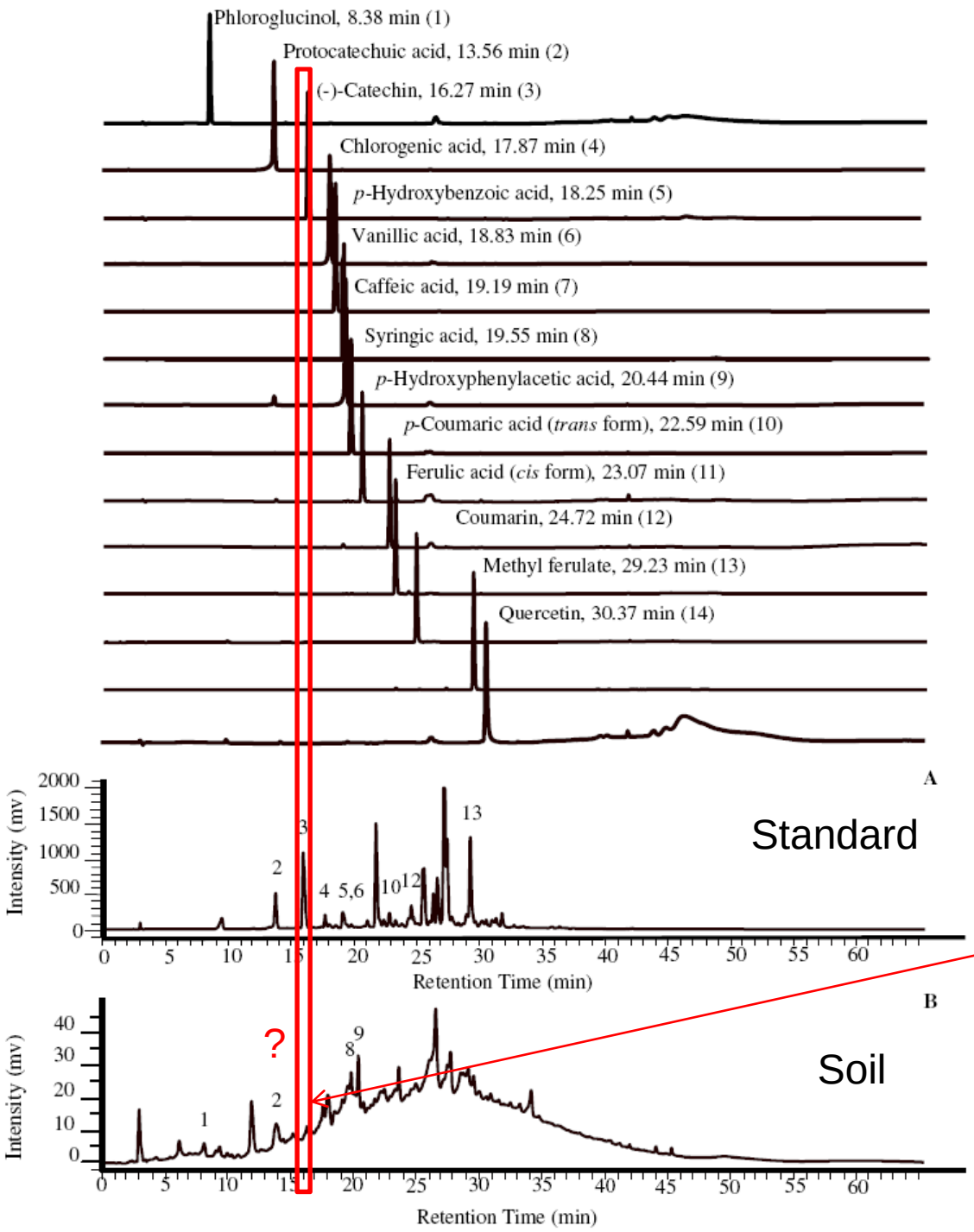

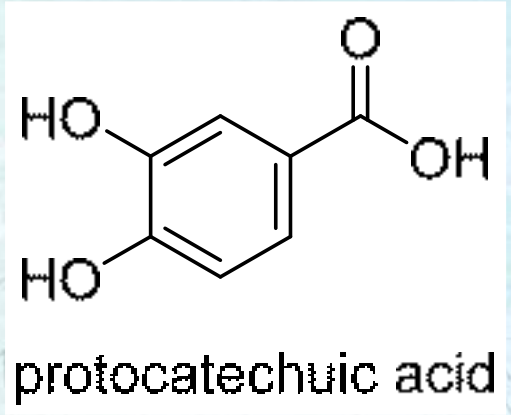

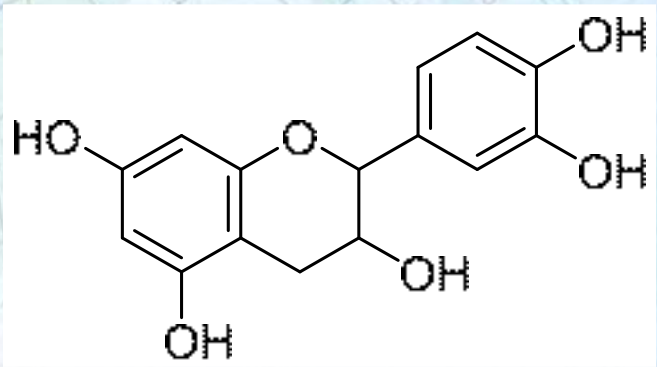

Catechin degradation

Chou et al., 2010

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The Impact of Microbial Biotransformation of Catechin in Enhancing the Allelopathic Effects of *Rhododendron formosanum*

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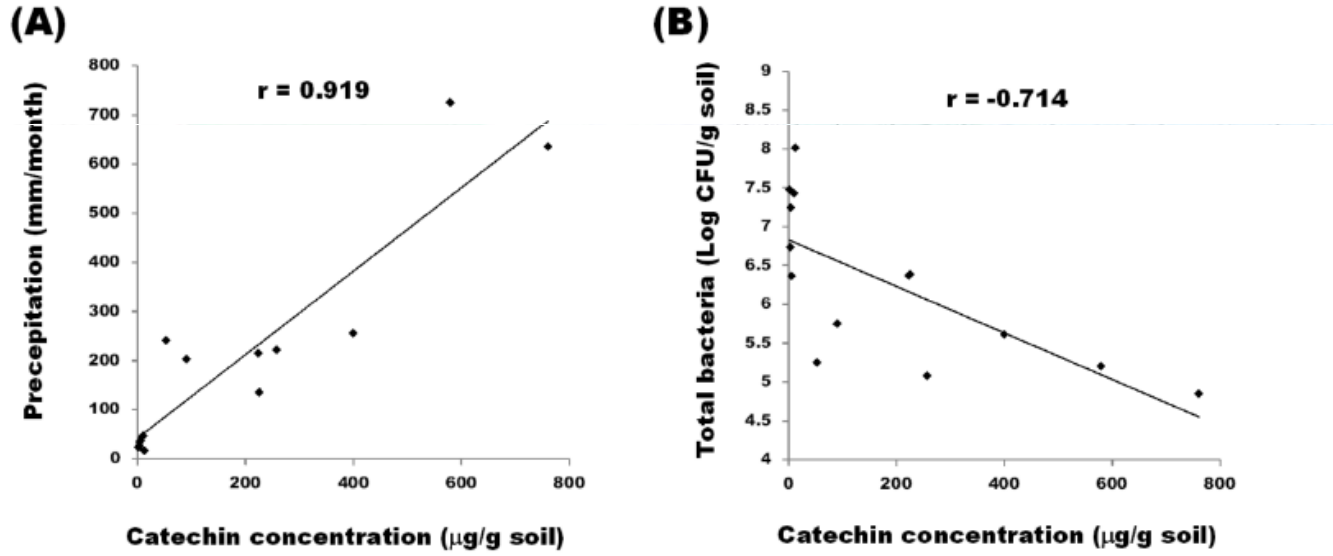
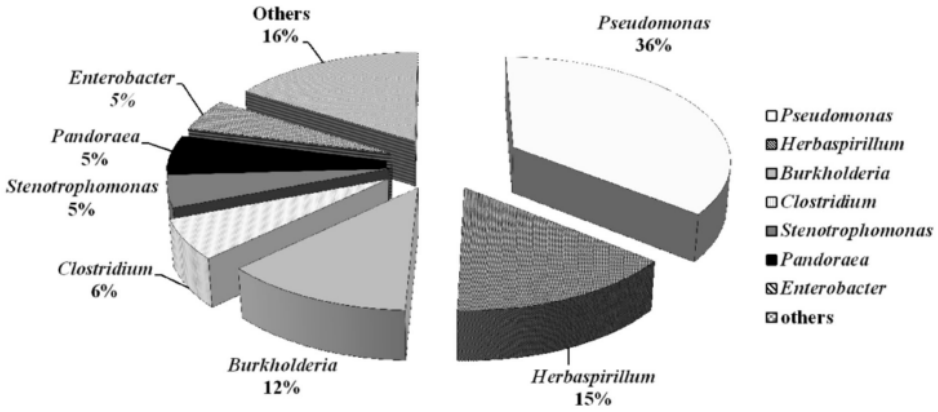


Figure 1. Pairwise correlations between (-)-catechin concentration, monthly precipitation, and bacterial populations. (A) Correlation between the concentration of (-)-catechin in soil of *R. formosanum* and monthly precipitation ($r = 0.919$, $P < 0.0001$, $n = 14$). (B) Correlation between the concentration of (-)-catechin and bacterial populations in soil of *R. formosanum* ($r = -0.714$, $P = 0.0041$, $n = 14$).

doi: 10.1371/journal.pone.0085162.g001

(A)

Bacterial flora in soil of *R. formosanum*



(B)

Catechin-Utilizing Bacteria

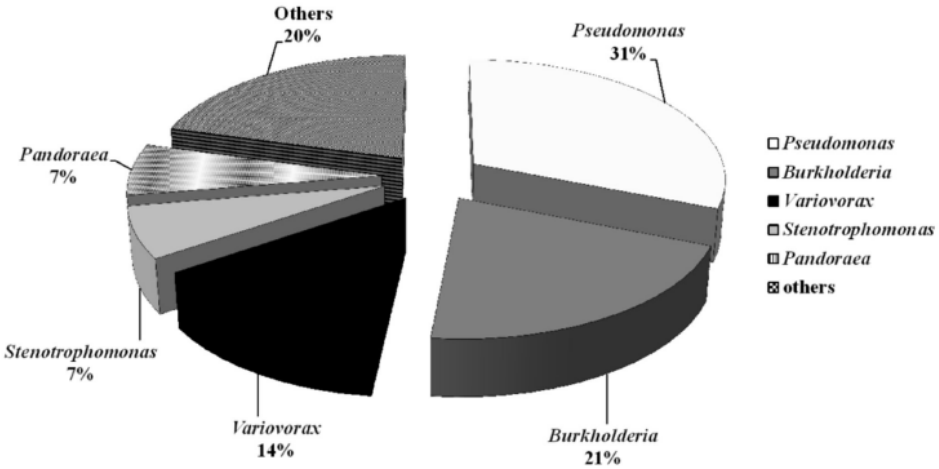


Figure 2. Bacterial flora (A) and catechin utilizing bacteria (B) in the rhizosphere of a *Rhododendron formosanum* plantation. (-)-Catechin was the only carbon source added to the medium that was used for microbial isolation. After 3 months of incubation, *Pseudomonas* spp., *Burkholderia* spp., *Variovorax* spp. *Stenotrophomonas* spp., and *Pandoraea* spp. were isolated and identified as dominant catechin-utilizing bacteria.

doi: 10.1371/journal.pone.0085162.g002

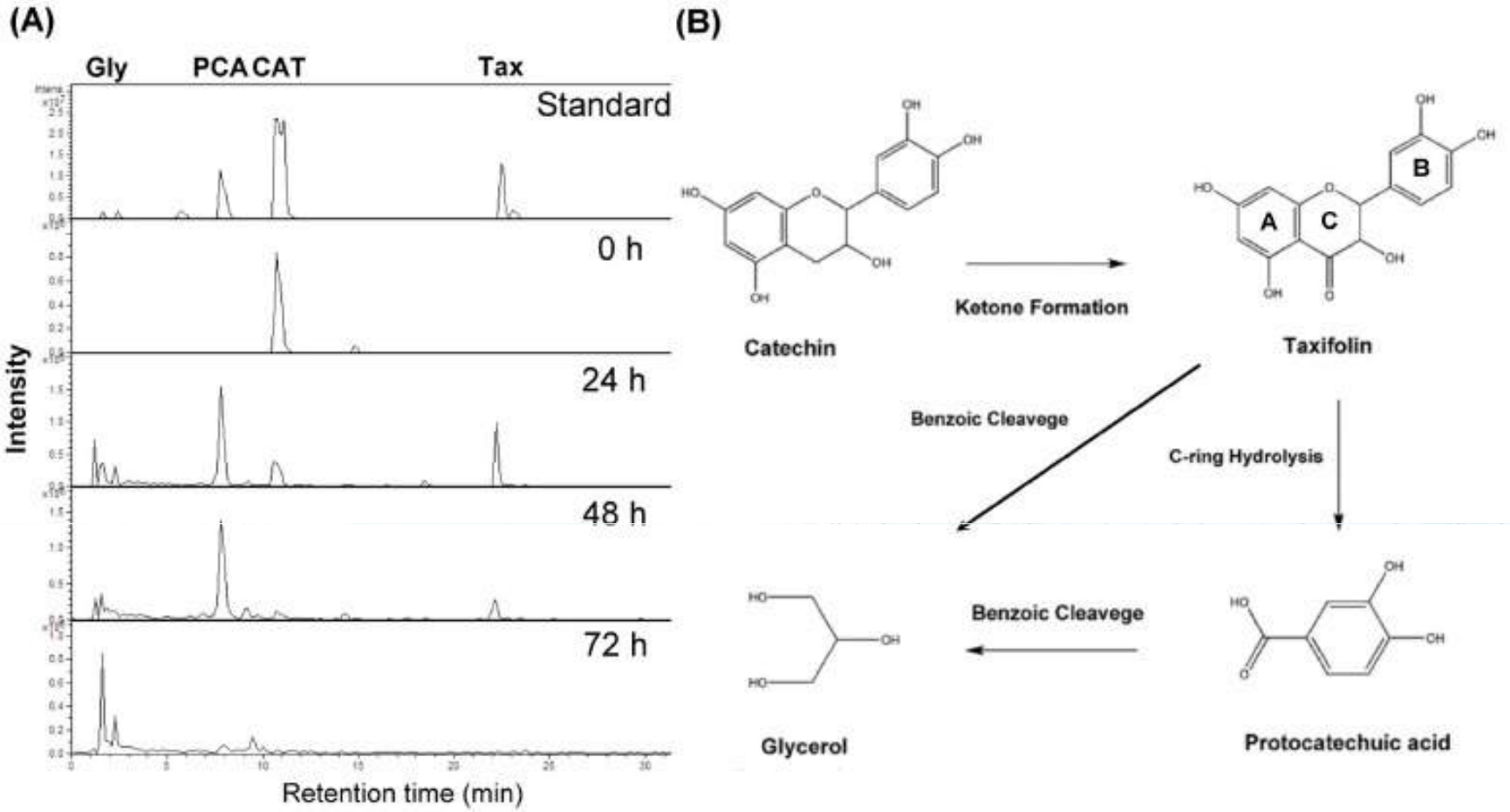


Figure 3. Metabolic pathway for (-)-catechin transformed by *Pseudomonas* sp. CRF3-Ps-1 was analysed by the LC-ESI-MS/MS method (A). (-)-Catechin (CAT) was transformed into taxifolin (Tax) via ketone formation during the first 24 h. Subsequently, C-ring hydrolysis occurred and generated protocatechuic acid (PCA) and glycerol (Gly). Finally, (-)-catechin was transformed into glycerol 72 h after incubation. The possible transformation hypothesis is also illustrated (B).

doi: 10.1371/journal.pone.0085162.g003

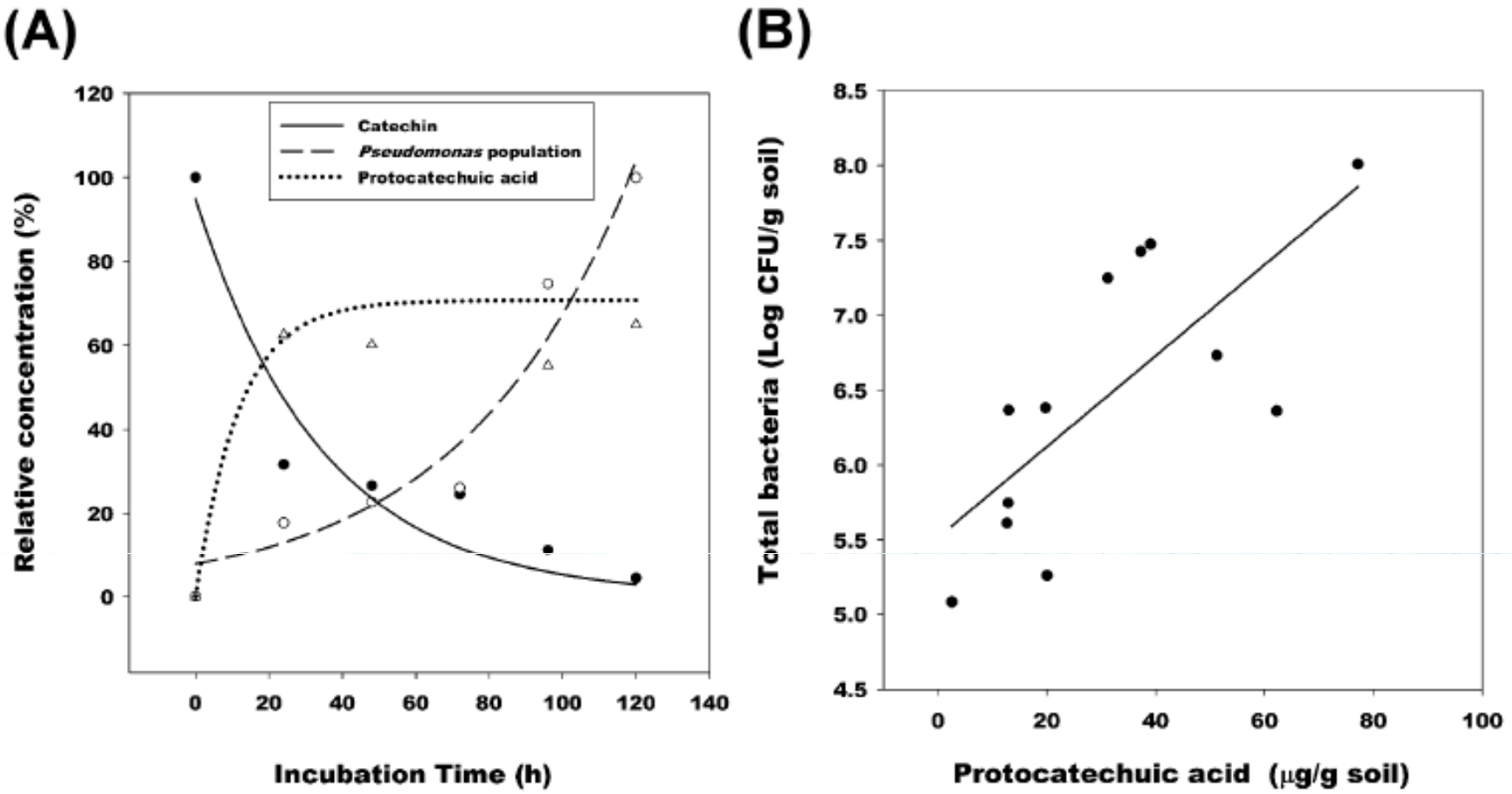


Figure 4. Relative concentrations of (-)-catechin and protocatechuic acid, and the bacterial population in the medium, during 120 h incubation with *Pseudomonas* sp. CRF3-Ps-1(A)(●) Relative concentration of catechin (r = -0.958, P = 0.0025, n = 6); (○) Bacterial population of *Pseudomonas* CRF3-Ps-1 (r = 0.974, P = 0.001, n = 6); (△) Relative concentration of protocatechuic acid (r = 0.874, P = 0.0226, n = 6). (B) Correlations between the concentration of protocatechuic acid and bacterial populations in the soil of *R. formosanum* (r = 0.734, P = 0.0066, n = 12).

doi: 10.1371/journal.pone.0085162.g004

Table 1. The concentration of allelochemicals in the soil of *Rhododendron formosanum* in Sunlinksea and Dasyueshan.

Study Site	Season	Catechin (μg/g soil)	Protocatechuic acid (μg/g soil)	Precipitation (mm/month)
Sunlinksea				
	Winter	5.30 ± 2.6	35.79 ± 2.3	35.67 ± 6.5
	Summer	178.36 ± 63.3*	14.07 ± 5.7*	199.77 ± 32.3**
	Typhoon	760	<1	635
Dasyueshan				
	Winter	6.94 ± 2.9	63.5 ± 7.5	27.37 ± 5.6
	Summer	237.44 ± 89.5	12.82 ± 0.1	224.93 ± 16.0***
	Typhoon	579	<1	726

Results are the mean ± SE values of 3 experiments. Asterisks indicate significant difference (*$p < 0.05$, **$p < 0.01$, ***$p < 0.005$) between seasons.

doi: 10.1371/journal.pone.0085162.t001

Table 2. The recovery value and half-life of standard (-)-catechin or protocatechuic acid in the soil from rhizosphere of *R. formosanum*.

Compound	(-)-Catechin	Protocatechuic acid
Added concentration (μg/g soil)	**200**	**25**
Measured mass concentration (μg/g soil)	180.2 ± 10.2	22.9 ± 1.1
Recovery (%)	90.1	91.5
RSD* (%)	5.0	4.4
Half-life in soil (hr)	4.8 ± 1.2	>120

* RSD: relative standard deviation

doi: 10.1371/journal.pone.0085162.t002

Table 3. Inhibitory effects of (-)-catechin and protocatechuic acid on the radicle length, germination, and ETR_{max} of *L. sativa* at the EC_{50} concentration.

	EC_{50} (mM)		
	Radicle	**Germination**	**ETR_{max}**
(-)-Catechin	10.8 ± 1.8	9.2 ± 1.3	> 20
Protocatechuic acid	4.4 ± 0.3*	4.3 ± 0.4*	3.2 ± 0.03

Results are the mean ± SE values of 3 experiments. Asterisk indicates significant difference ($p < 0.05$) between treatments.

doi: 10.1371/journal.pone.0085162.t003

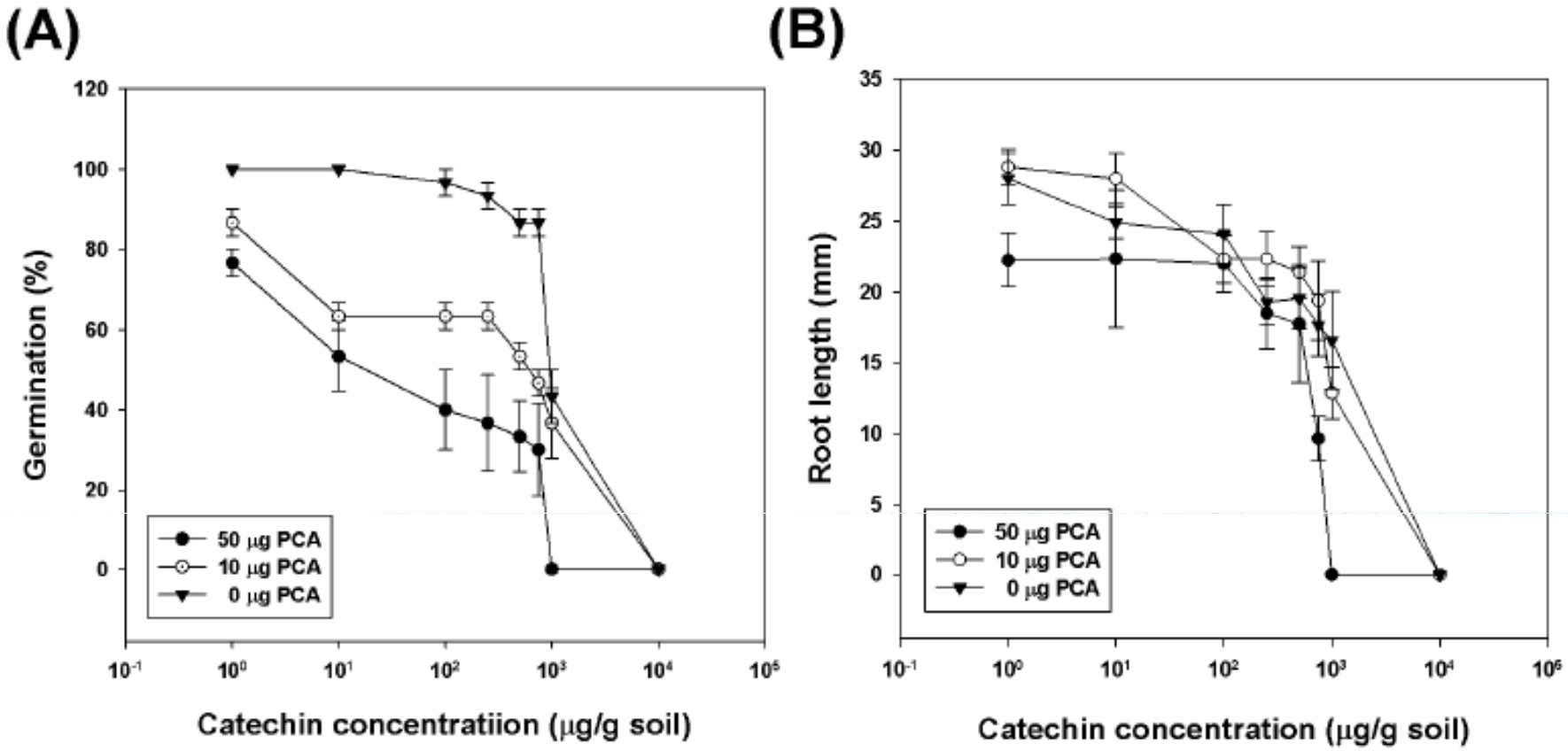


Figure 5. The phytotoxic effects of (-)-catechin on the seed germination (A) and radicle growth (B) of *Lactuca sativa* at different concentrations in combination with 0 μg, 10 μg and 50 μg protocatechuic acid. Error bars represent the standard errors of the mean.

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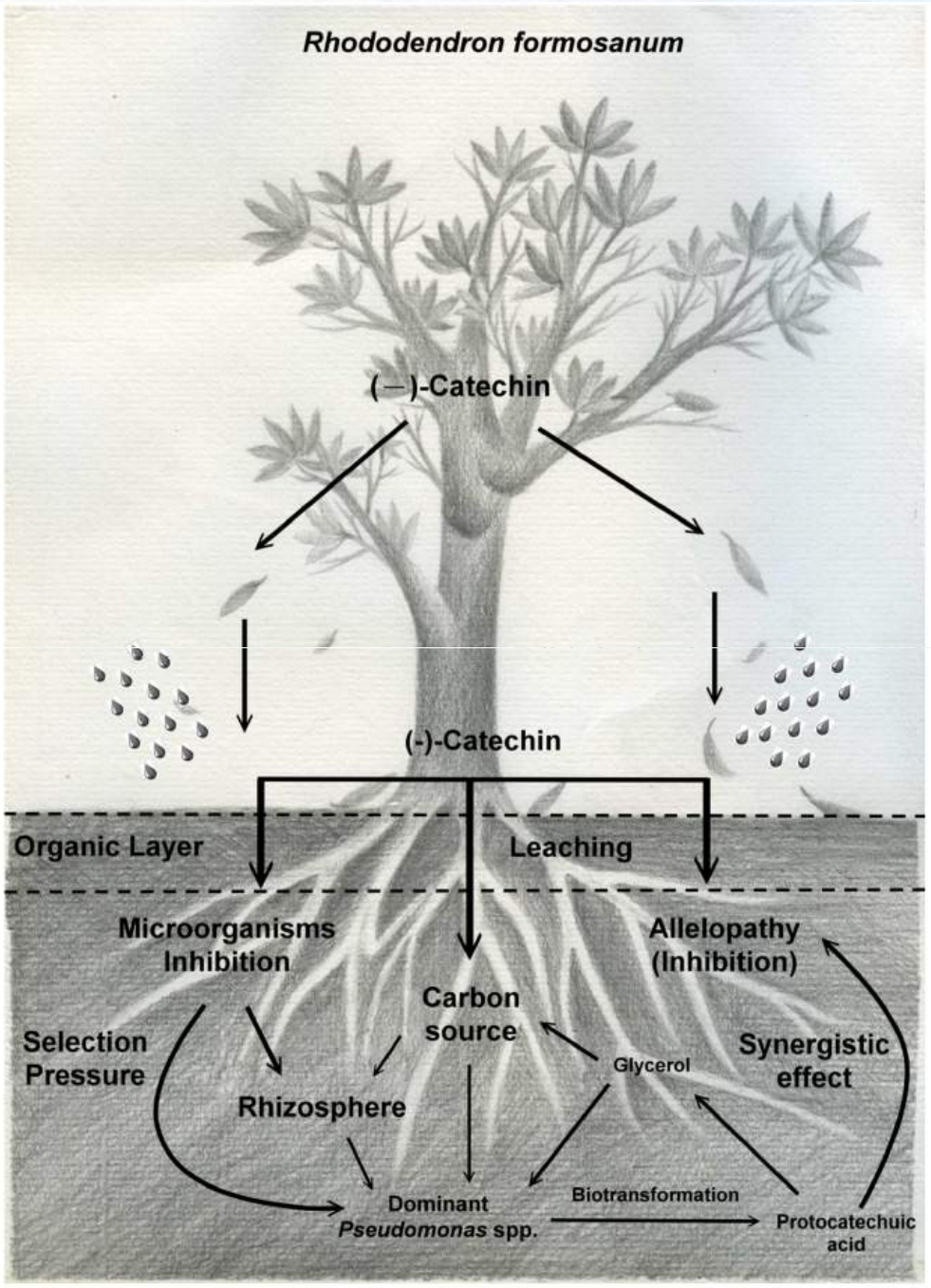

Summary

- Plants produce an array of chemicals and that many of these chemicals leach into the rhizosphere and have allelopathic effects on soil conditions, neighbouring plants, and microorganisms.
- Allelochemicals are utilized as nutrient sources and lead to an increase in the population of the selected microorganisms.
- Intermediate compounds of biotransformation have synergistic allelopathic effects with the original compounds

Plant Natural Products Can be Used as Pharmaceutical Agent

- **Anticancer**
- **Antioxidant**
- **Antibiotic**

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Chemical Constituents of *Rhododendron formosanum* Show Pronounced Growth Inhibitory Effect on Non-Small-Cell Lung Carcinoma Cells

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Lung cancer and Apoptosis

- **Lung cancer is the second most frequent type of cancer**
- **Nonsmall-cell lung cancer (NSCLC) accounts for ~80% of primary lung cancers**
- **Apoptosis, a major form of cell death, is regulated by programmed cellular signaling pathways**
- **Apoptosis is associated with characteristic morphological changes including the formation of apoptotic bodies, chromatin, and nuclear condensation and DNA fragmentation**

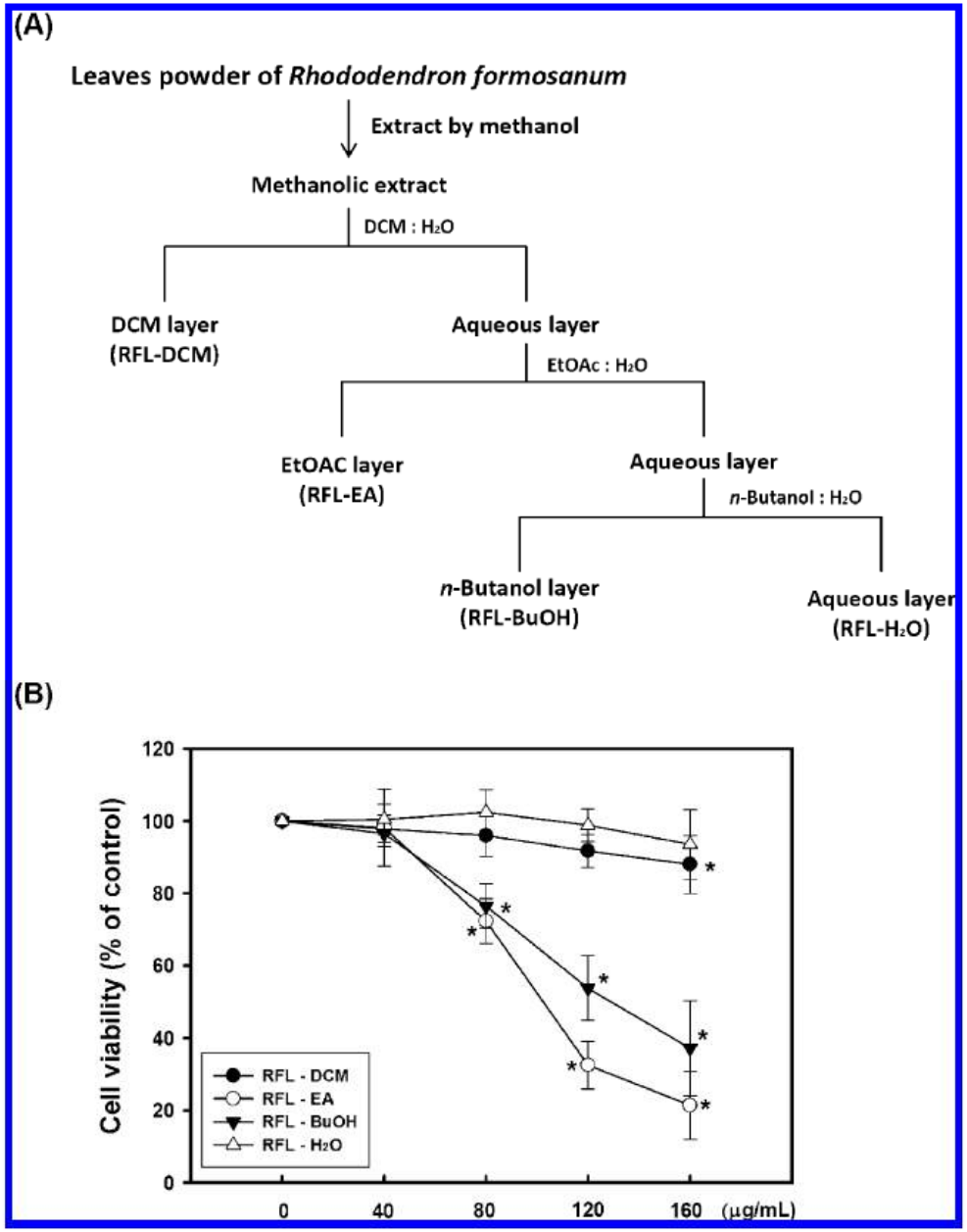


Figure 1. Antiproliferation activities of different fractions from *R. formosanum*. (A) Leaf powder of *R. formosanum* was extracted by methanol and partitioned into four fractions including dichloromethane, ethyl acetate, *n*-butanol, and water to get RFL-DCM, RFL-EA, RFL-BuOH, and RFL-H_2O fractions. (B) A549 cells were treated with the four fractions (40–160 μg/mL) for 24 h. Cell viability was then determined using the MTT assay. This experiment was repeated three times. The data represent the mean ± SD. Values were significantly different from the control group: ∗, $P < 0.05$.

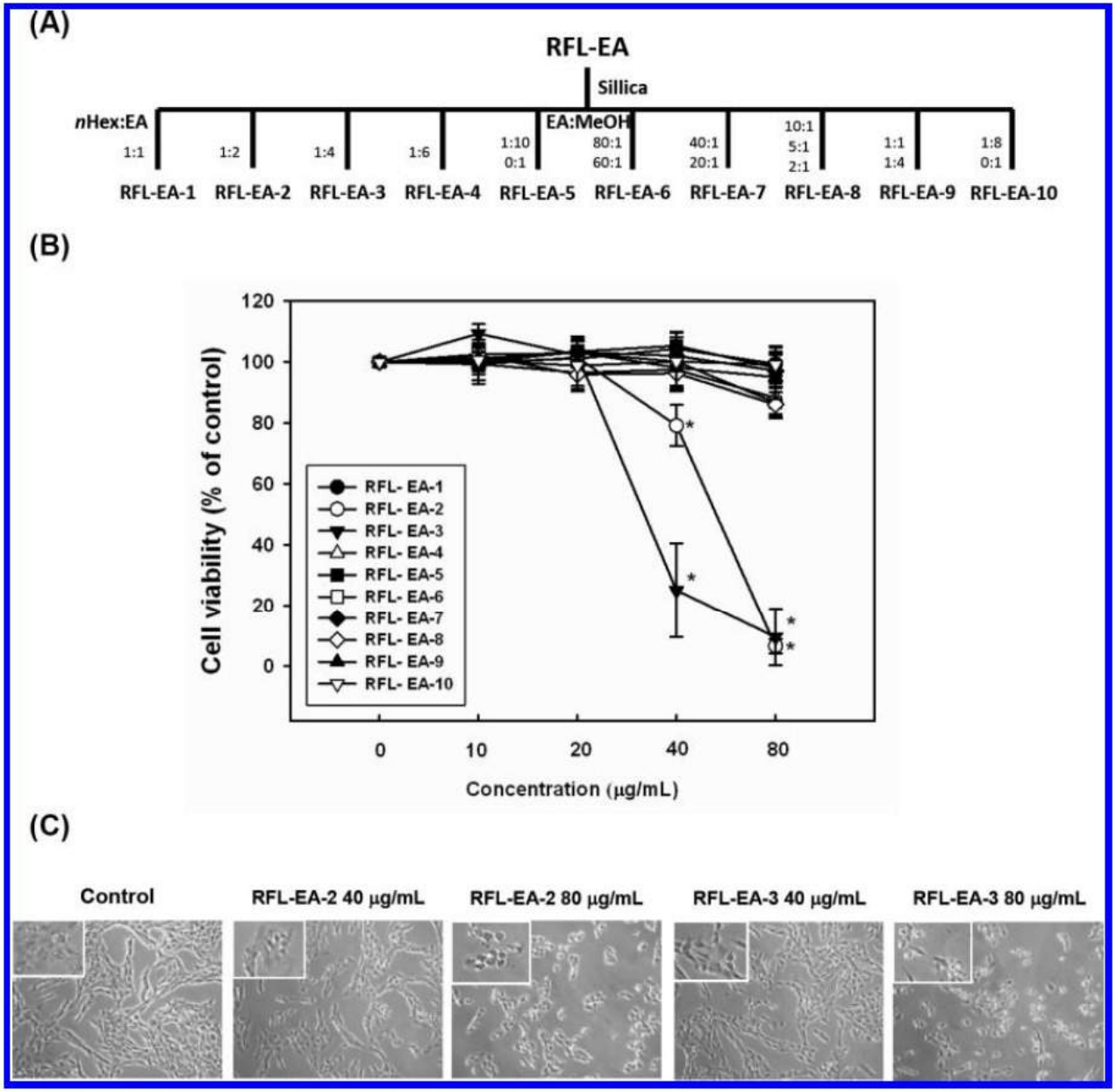


Figure 2. Antiproliferation activities of different RFL-EA fractions against NSCLC A549 cells. (A) RFL-EA was injected into a silica column and eluted with *n*-hexane, ethyl actate, and methanol at different combination rates to get 10 fractions. (B) A549 cells were treated with the 10 fractions (10−80 μg/mL) for 24 h. Cell viability was then determined using the MTT assay. This experiment was repeated three times. The data represent the mean ± SD. Values were significantly different from the control group: ∗, $P < 0.05$. (C) A549 cells were treated with RFL-EA-2 (40−80 μg/mL) and RFL-EA-3 (40−80 μg/mL) for 24 h, and cell morphology was observed by photomicroscope.

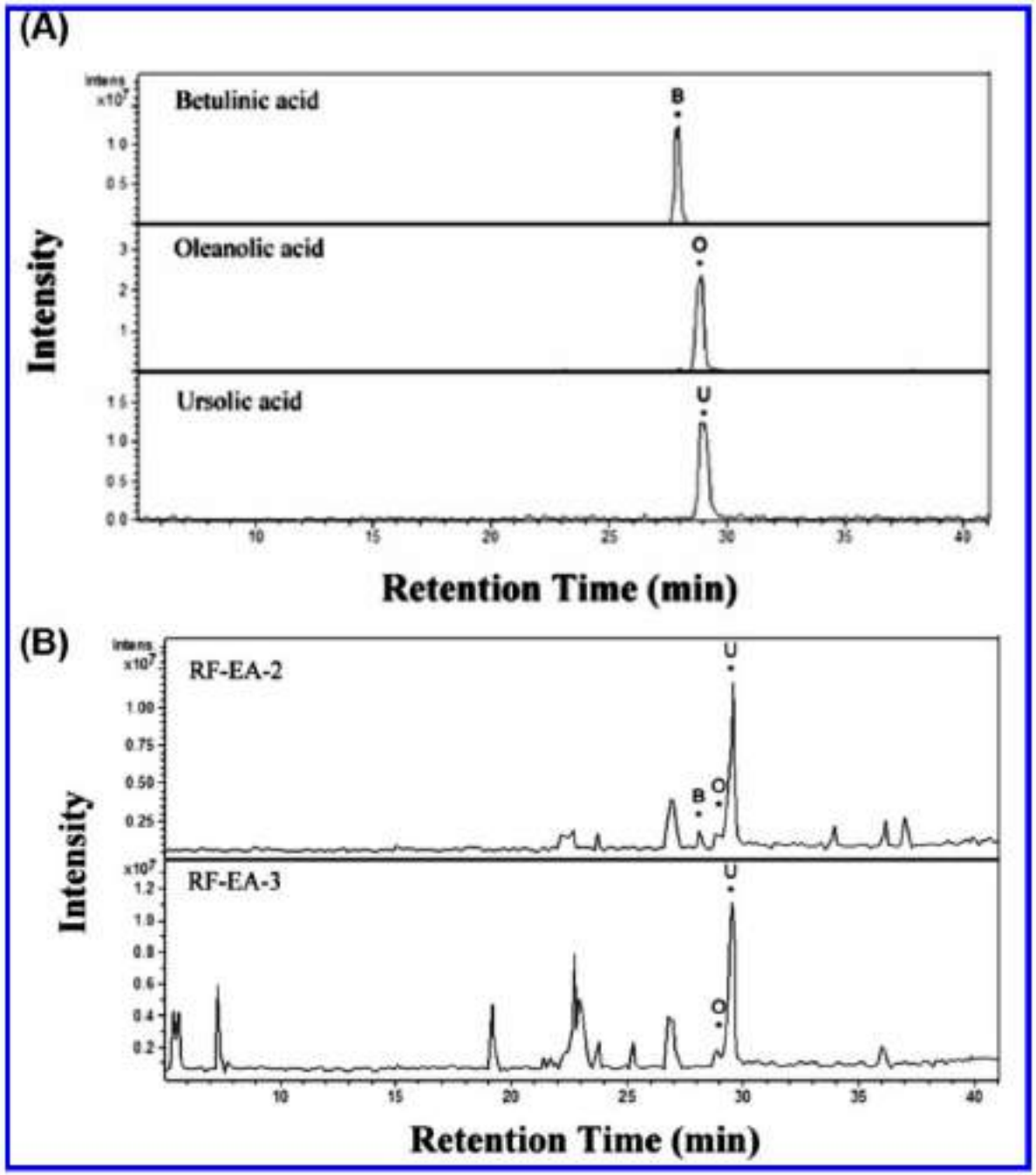


Figure 3. Quantification of triterpenoids by LC-ESI-MS/MS analysis. (A) Individual standard triterpenoids including betulinic acid (B), oleanolic acid (O), and ursolic acid (U) were subjected to LC-ESI-MS/MS analysis for chemical identification and quantification. Panel B shows quantification of the betulinic acid, oleanolic acid, and ursolic acid from RFL-EA-2 and RFL-EA-3 fractions.

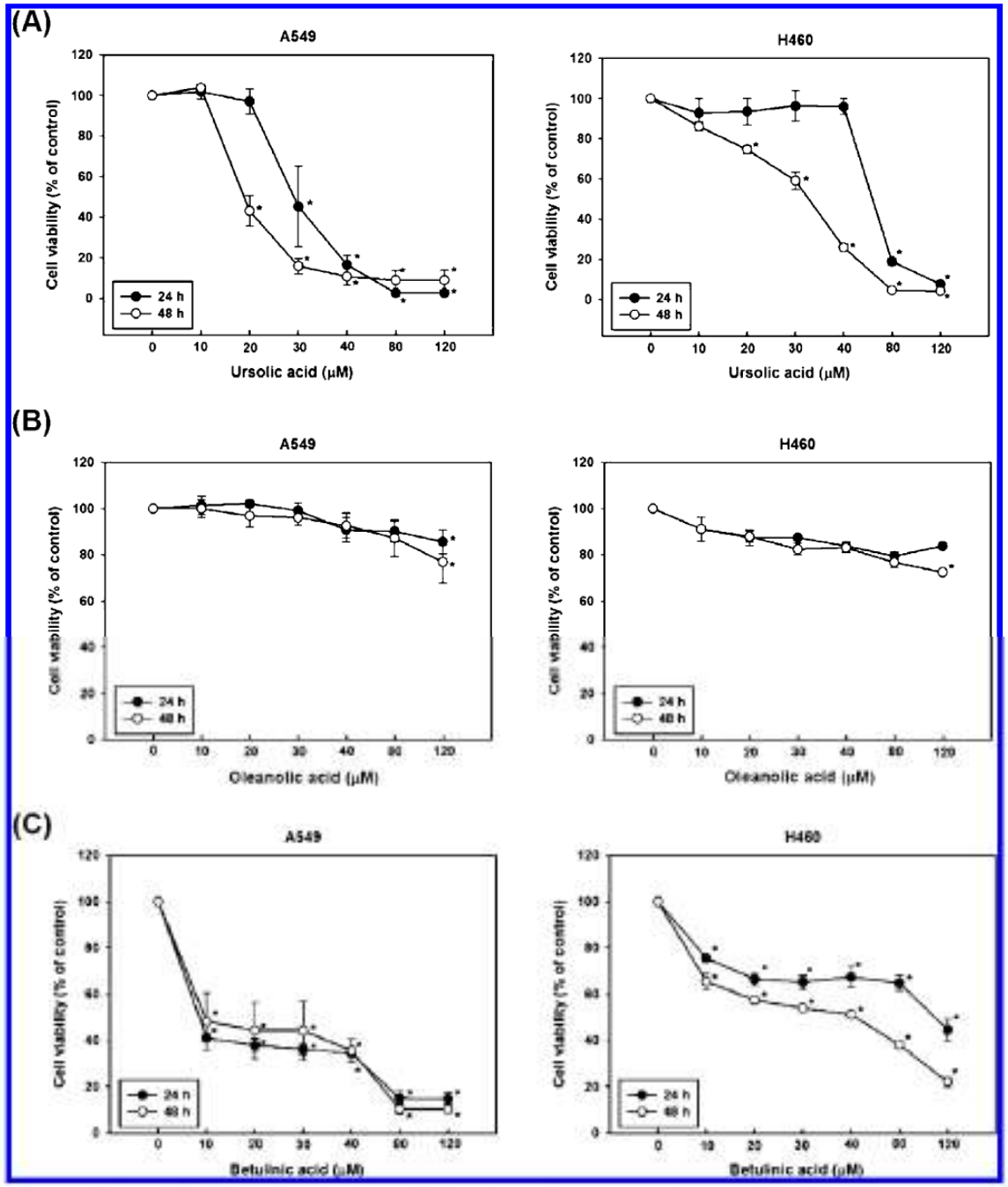


Figure 4. Antiproliferation effect of ursolic acid, oleanolic acid, and betulinic acid against NSCLC cells. A549 cells and H460 cells were treated with (A) ursolic acid, (B) oleanolic acid, and (C) betulinic acid at concentrations of 10–160 μM for 24 or 48 h. Cell viability was then determined using the MTT assay. This experiment was repeated three times. The data represent the mean ± SD. Values were significantly different from the control group: *, $P <$ 0.05.

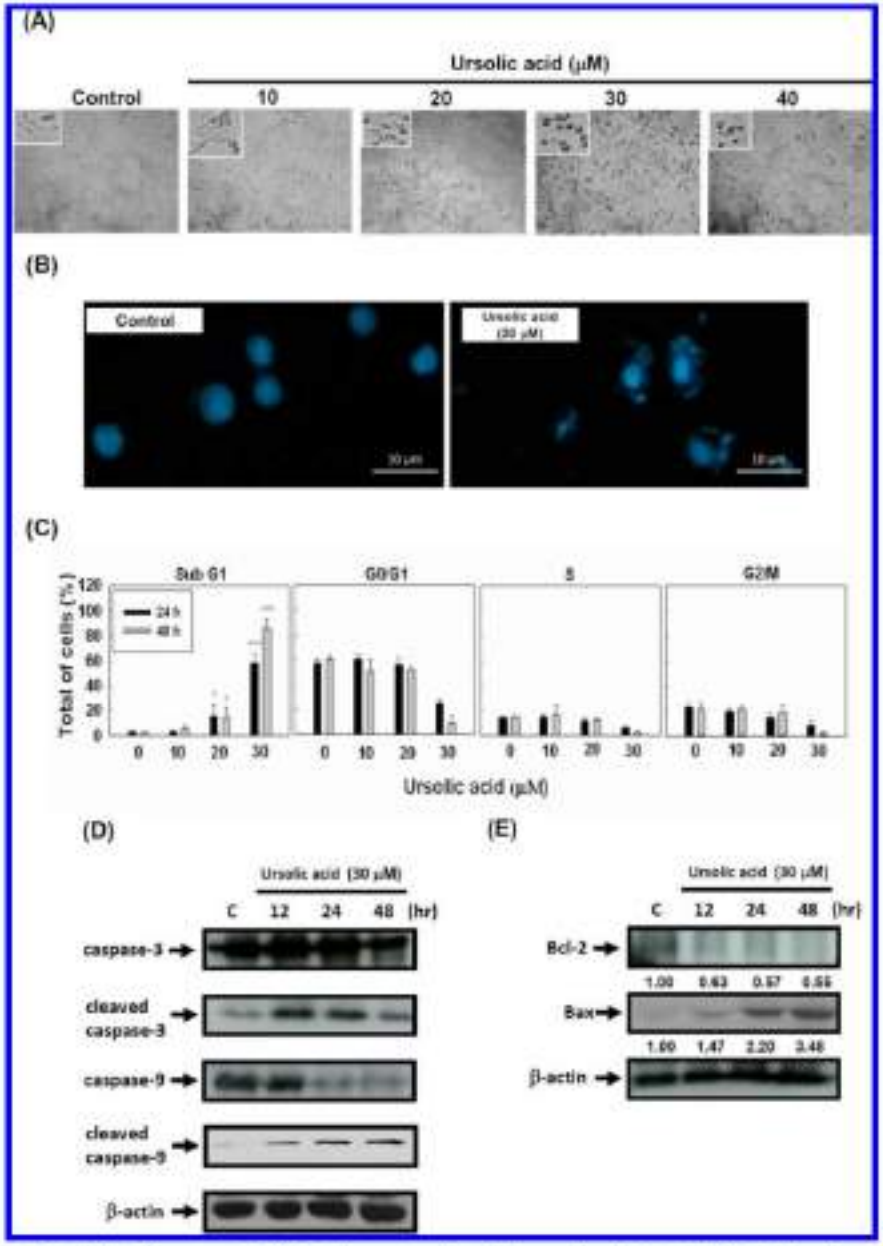


Figure 5. Ursolic acid induced A549 cell apoptosis. (A) A549 cells were incubated with ursolic acid (10–40 μM) for 24 h, and morphology was observed by photomicroscope; (B) the morphology of cell nuclei was observed by fluorescence microscope. (C) To observe cell cycle statements and apoptosis levels, cell were stained with PI and measured by flow cytometry. (D) A549 cells were treated with ursolic acid (30 μM) for the indicated time. Cells were then harvested and lysed for the detection of caspase-3, cleaved caspase-3, caspase-9, cleaved caspase-9, and β-actin. (E) A549 cells were treated with ursolic acid (30 μM) for the indicated times. Cells were then harvested and lysed for the detection of Bcl-2, Bax, and β-actin. Western blot data presented are representative of those obtained in at least three separate experiments. The values below the figures represent change in protein expression of the bands normalized to β-actin.

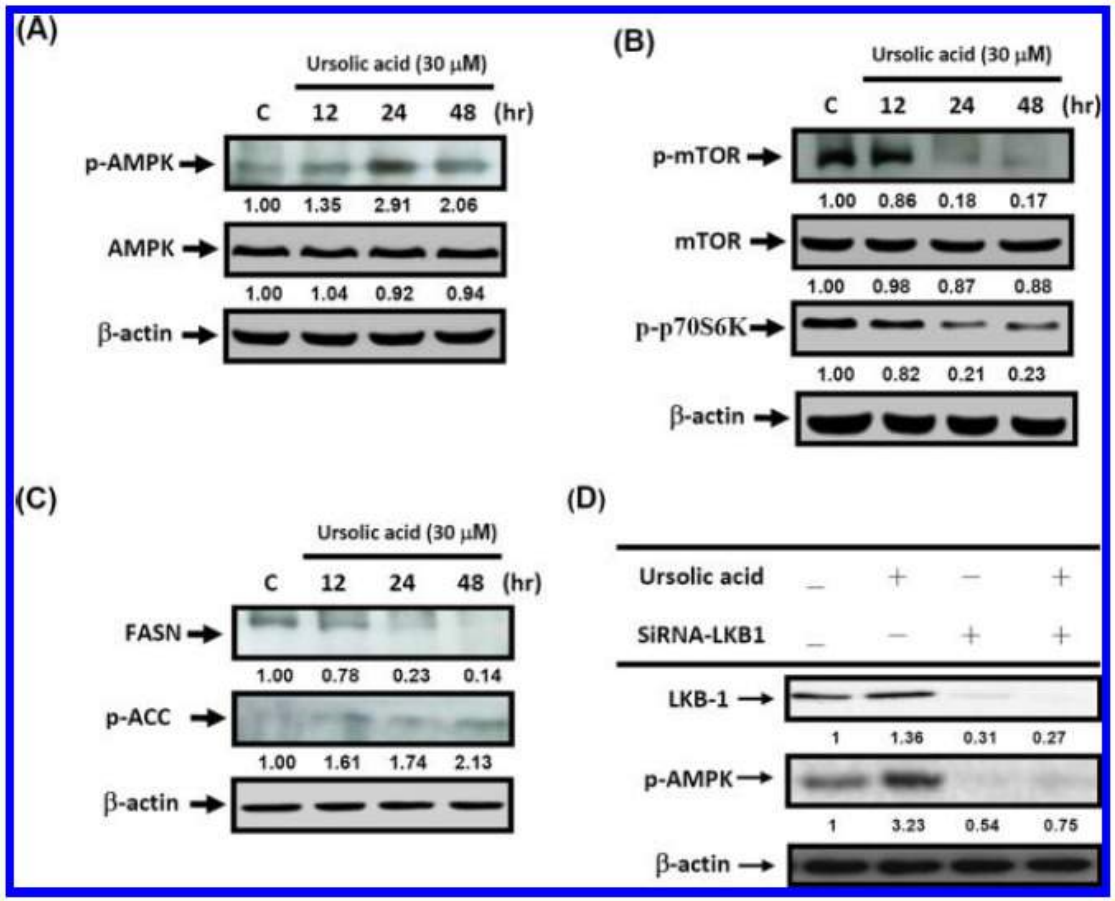
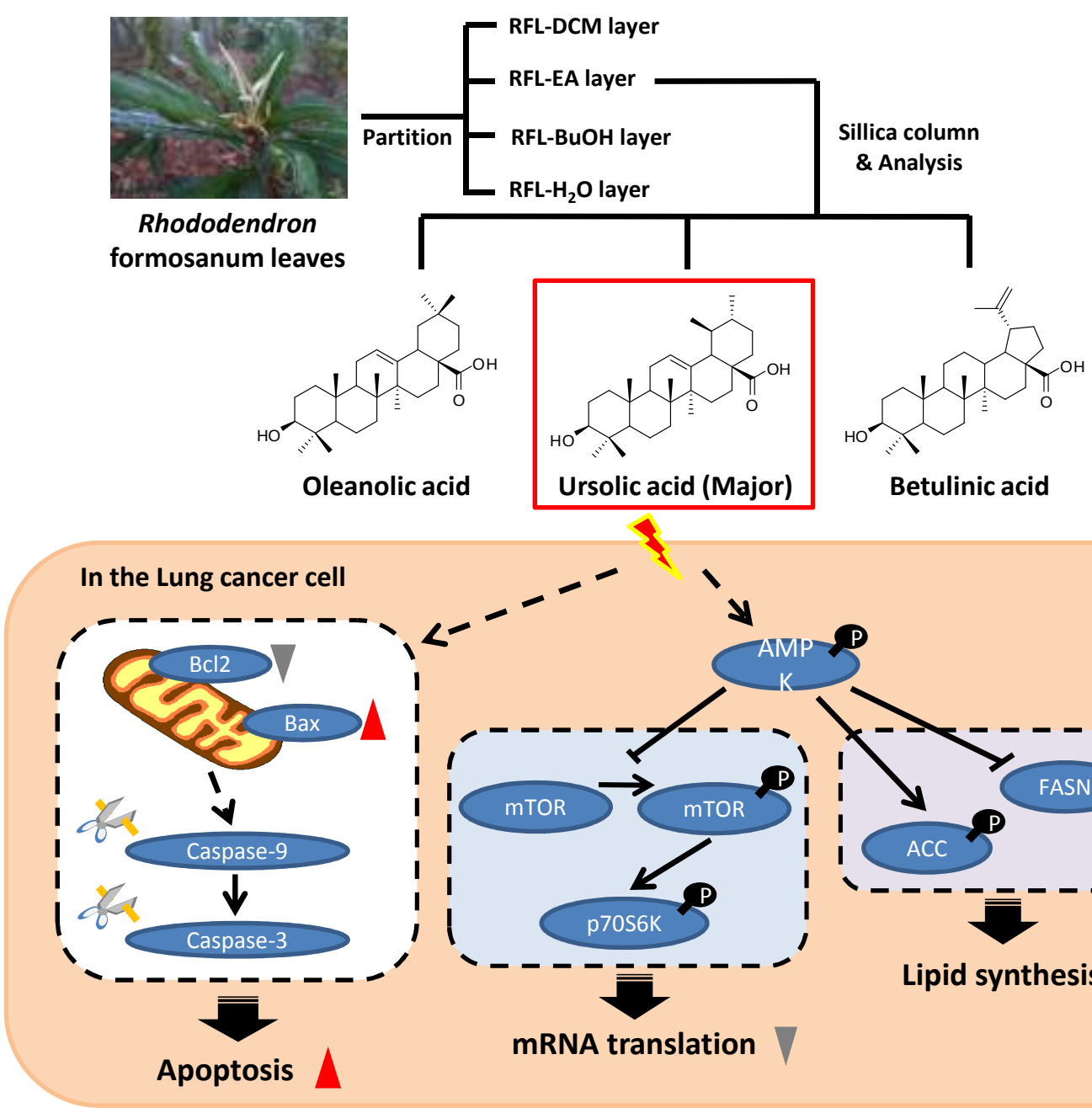


Figure 6. Ursolic acid decreases general mRNA translation and the activity of fatty acid synthesis via activation of AMPK. (A) A549 cells were treated with ursolic acid (30 μM) for the indicated times. Cells were then harvested and lysed for the detection of phosphorylated AMPK (Thr 172), AMPK, and β-actin. (B) A549 cells were treated with ursolic acid (30 μM) for the indicated times. Cells were then harvested and lysed for the detection of phosphorylated mTOR (Ser 2448), mTOR, phosphorylated p70S6K, and β-actin. (C) A549 cells were treated with ursolic acid (30 μM) for the indicated times. Cells were then harvested and lysed for the detection of FASN, phospho-ACC (Ser79), and β-actin. (D) A549 cells were transfected with 50 nmol/L LKB1-siRNA using Oligofectamine. A total of 24 h after transfection, cells were treated with ursolic acid (30 μM) for 24 h. After harvesting, cells were lysed and prepared for Western blotting analysis using antibodies against LKB-1, phosphorylated AMPK (Thr 172), and β-actin. Western blot data presented are representative of those obtained in at least three separate experiments. The values below the figures represent change in protein expression of the bands normalized to β-actin.


Rhododendron formosanum leaves
Partition
RFL-DCM layer
RFL-EA layer
RFL-BuOH layer
RFL-H2O layer
Sillica column & Analysis
Oleanolic acid
Ursolic acid (Major)
Betulinic acid
In the Lung cancer cell
Bcl2
Bax
Caspase-9
Caspase-3
Apoptosis
AMP K
mTOR
mTOR
p70S6K
mRNA translation
FASN
ACC
Lipid synthesis

Molecules **2015**, *20*, 12787-12803; doi:10.3390/molecules200712787

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Article

Structure Elucidation of Procyanidins Isolated from *Rhododendron formosanum* and Their Anti-Oxidative and Anti-Bacterial Activities

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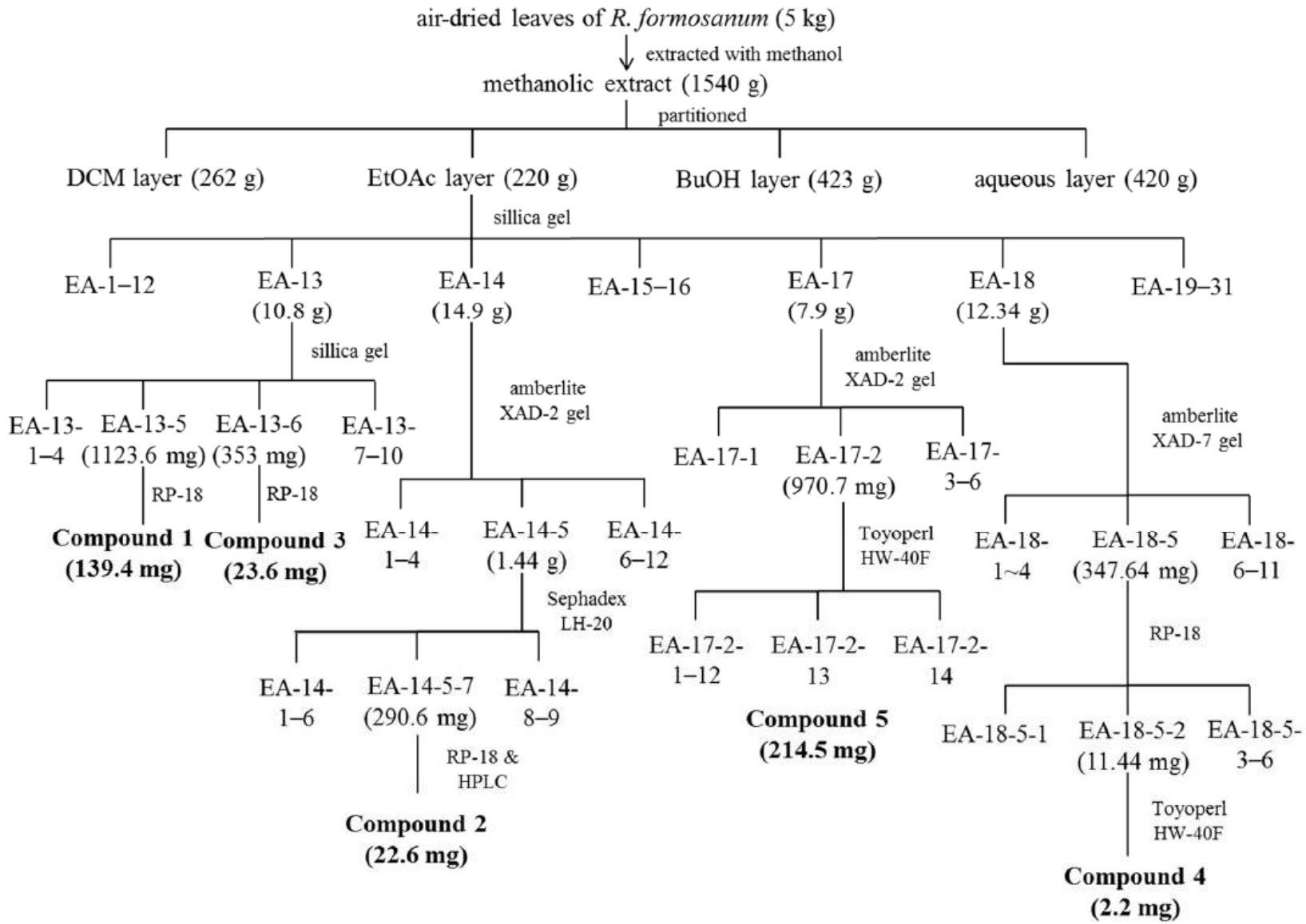
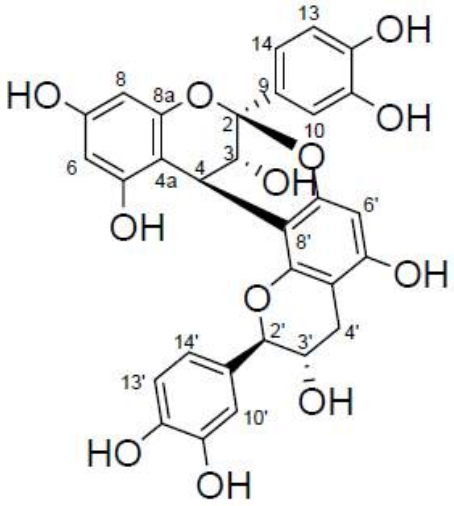
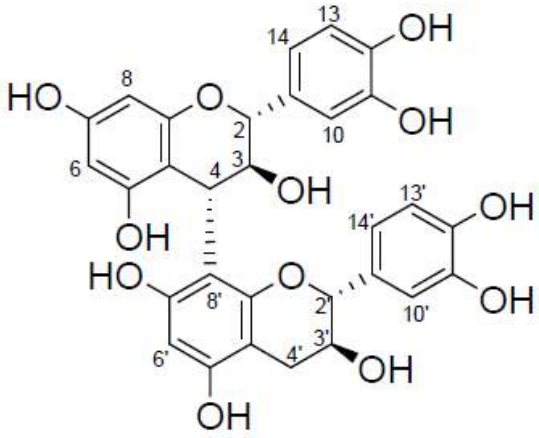


Figure 5. Purification flow chart of procyanidins isolated from *R. formosanum*.



Procyanindin A1



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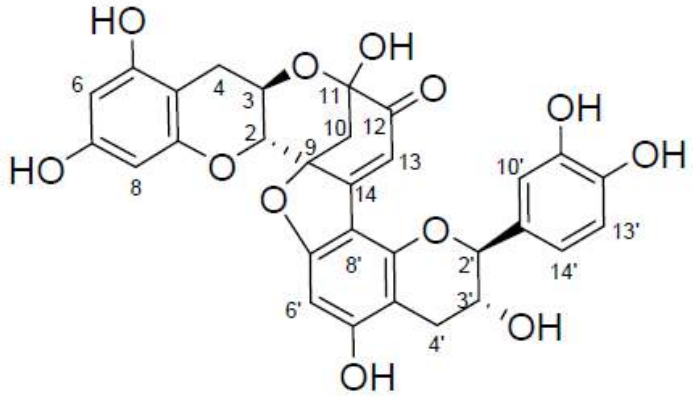
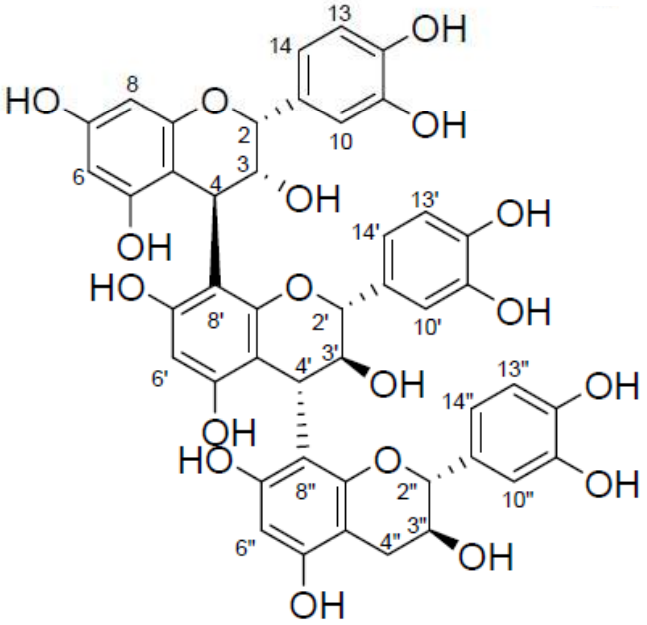
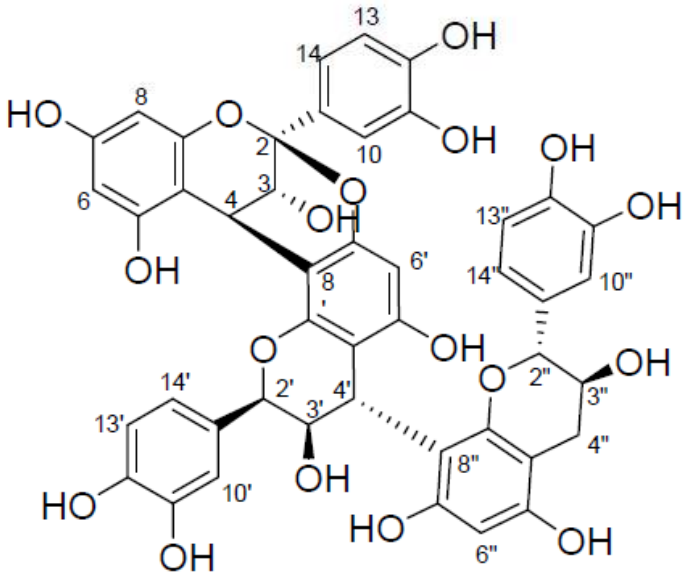


Rhodonidin A



Procyanindin C4



Cinnamtannin D1

Figure 1. Chemical structure of procyanidines from *R. formosanum*

Table 2. The minimum inhibitory concentration (μg/mL) of antibiotics and natural procyanidins for different bacterial pathogens.

Pathogens	Minimum Inhibitory Concentration (μg/mL)							
	Antibiotics and Procyanidins							
	Ap *	Tet	Met	1	2	3	4	5
Staphylococcus aureus	16	8	+N.D.	64	64	4	>128	>128
Enterococcus faecalis	2	4	N.D.	>128	>128	>128	>128	>128
Listeria monocytogenes	1	2	N.D.	64	>128	>128	>128	>128
Bacillus cereus	128	4	N.D.	64	>128	>128	>128	>128
Escherichia coli	4	0.5	N.D.	>128	>128	>128	>128	>128
Salmonella enterica	1	8	N.D.	>128	>128	>128	>128	>128
Pseudomonas aeruginosa	512	32	N.D.	>128	>128	>128	>128	>128
Helicobacter pylori **	N.D.	N.D.	2	>256	>256	>256	>256	>256

* Ap: ampicillin; Tet: tetracycline; Met: metronidazole; **1**: procyanidin A1; **2**: procyanidin B3; **3**: rhodonidin A; **4**: procyanidin C4; **5**: cinnamtannin D1; ** *H. pylori* was tested by minimum bactericidal concentration method. +N.D.: not determined.

Table 3. The antioxidant activities of the procyanidins from leaves of *R. formosanum* using the DPPH free radical-scavenging assay (IC_{50}, μM) and CUPric reducing antioxidant capacity (CUPRAC) method (TEACs).

Compounds	Total OH Groups	Average OH/unit	Antioxidant Activity	
			IC_{50}/DPPH (μg/mL)	CUPRAC (TEACs)
Trolox	2	2	61.12	1.00
(−)-Catechin	5	5	27.07	2.74
1	9	4.5	20.89	1.75
2	10	5	8.55	4.87
3	7	3.5	13.06	1.96
4	15	5	6.26	3.48
5	14	4.7	3.29	2.93

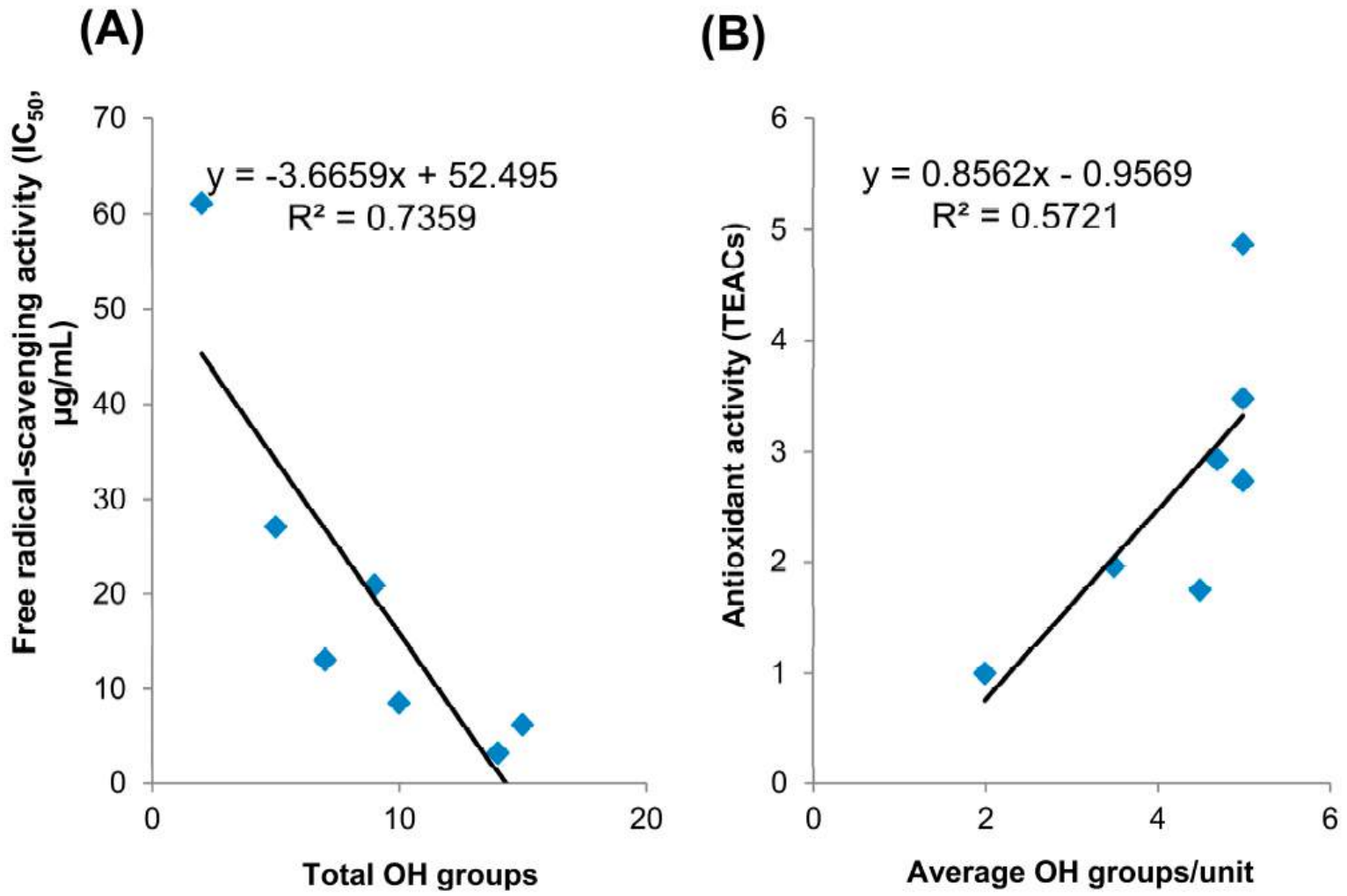


Figure 4. Correlations of total OH groups with free radical-scavenging activity (**A**) and average OH groups/unit with antioxidant activity (**B**).

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Cinnamtannin D1 from *Rhododendron formosanum* Induces Autophagy via the Inhibition of Akt/mTOR and Activation of ERK1/2 in Non-Small-Cell Lung Carcinoma Cells

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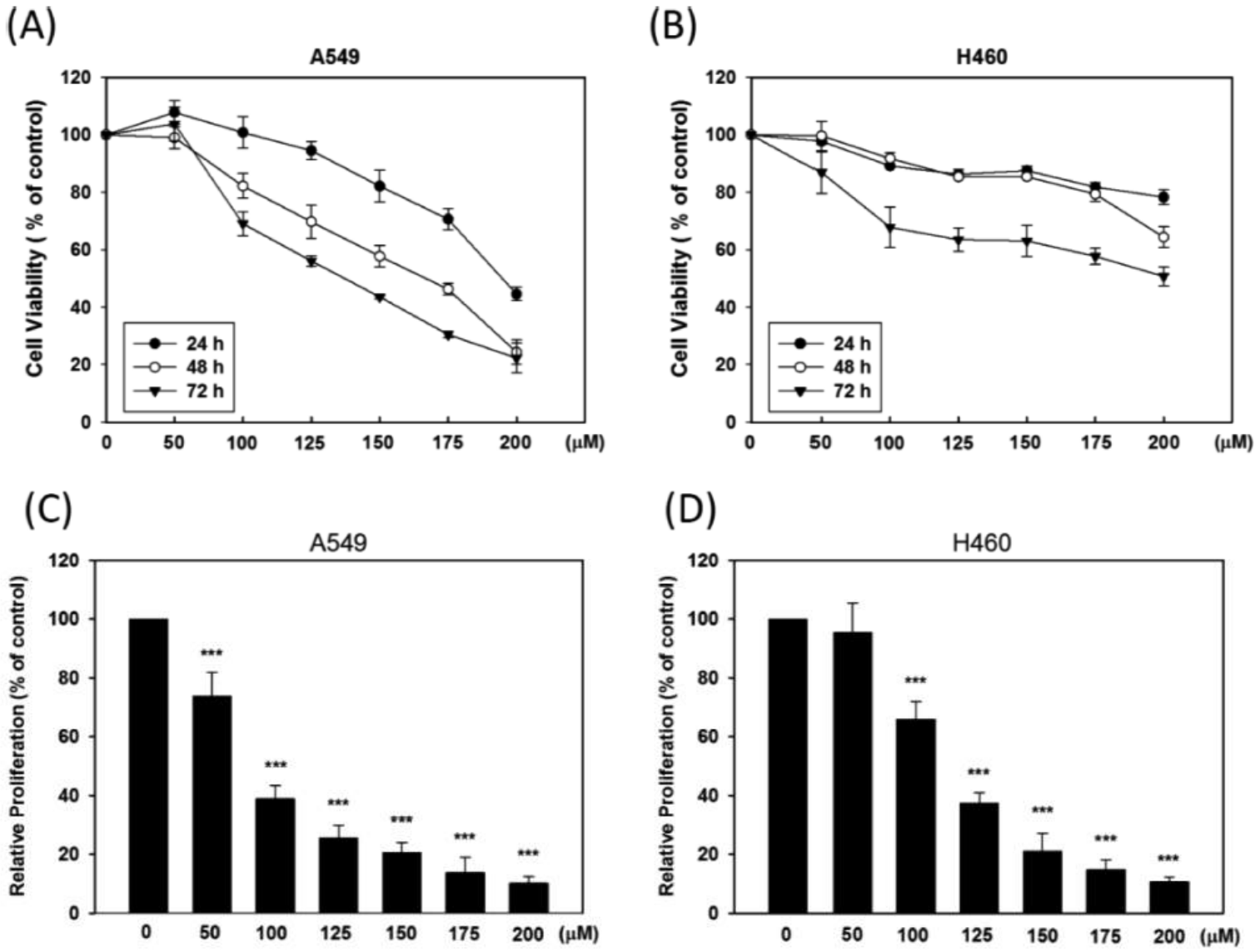


Figure 2. Effect of CNT D1 on antiproliferation activity. (A) A459 and (B) H460 cells were treated with CNT D1 at different concentrations for 24, 48, and 72 h and measured by the MTT assay. After (C) A459 and (D) H460 cells were treated with CNT D1 for 72 h, the cell proliferation ability of A549 and H460 cells was measured by the BrdU cell proliferation assay. These experiments were repeated three times. The data represent the mean ± SD.

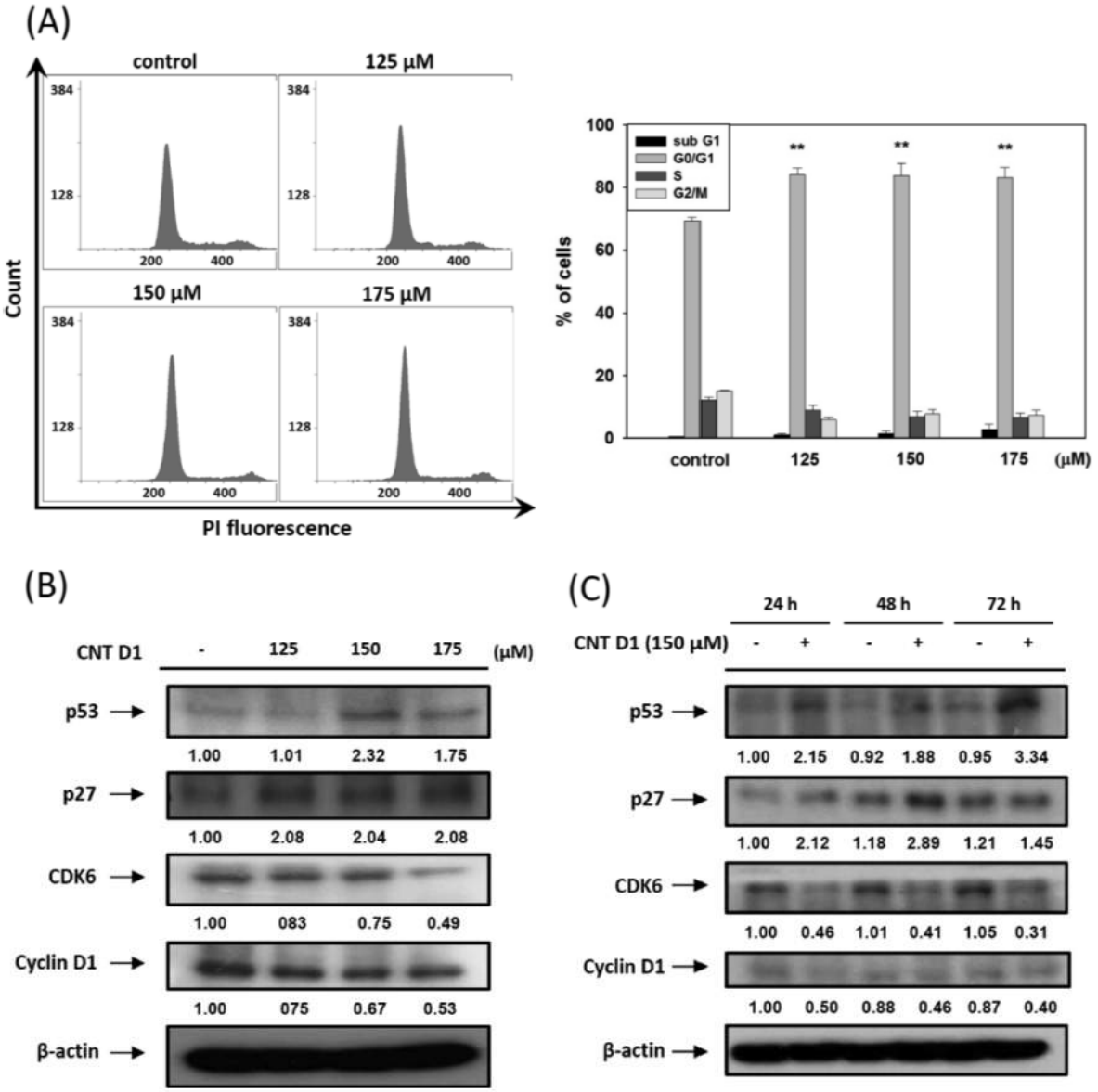


Figure 3. Effect of CNT D1 on cell cycle distribution and related protein expression. (A) To observe the state of the cell cycle, A549 cells were treated with CNT D1 at 125, 150, and 175 μM for 72 h. Cells were stained with PI and measured by flow cytometry. Results are described as means ± SD, and the statistical significance between the control group and different experimental groups is presented with asterisks (*, $P < 0.05$; **, $P < 0.005$; ***, $P < 0.001$). (B) A549 cells were treated with CNT D1 at 125, 150, and 175 μM for 72 h. (C) A549 cells were treated with CNT D1 (150 μM) for the indicated time. The protein expression of p53, p27, CDK6, and cyclin D1 was measured by Western blot. Western blot data presented are representative of those obtained in at least three separate experiments. The values below the figures represent the change in protein expression of the bands normalized to β-actin.

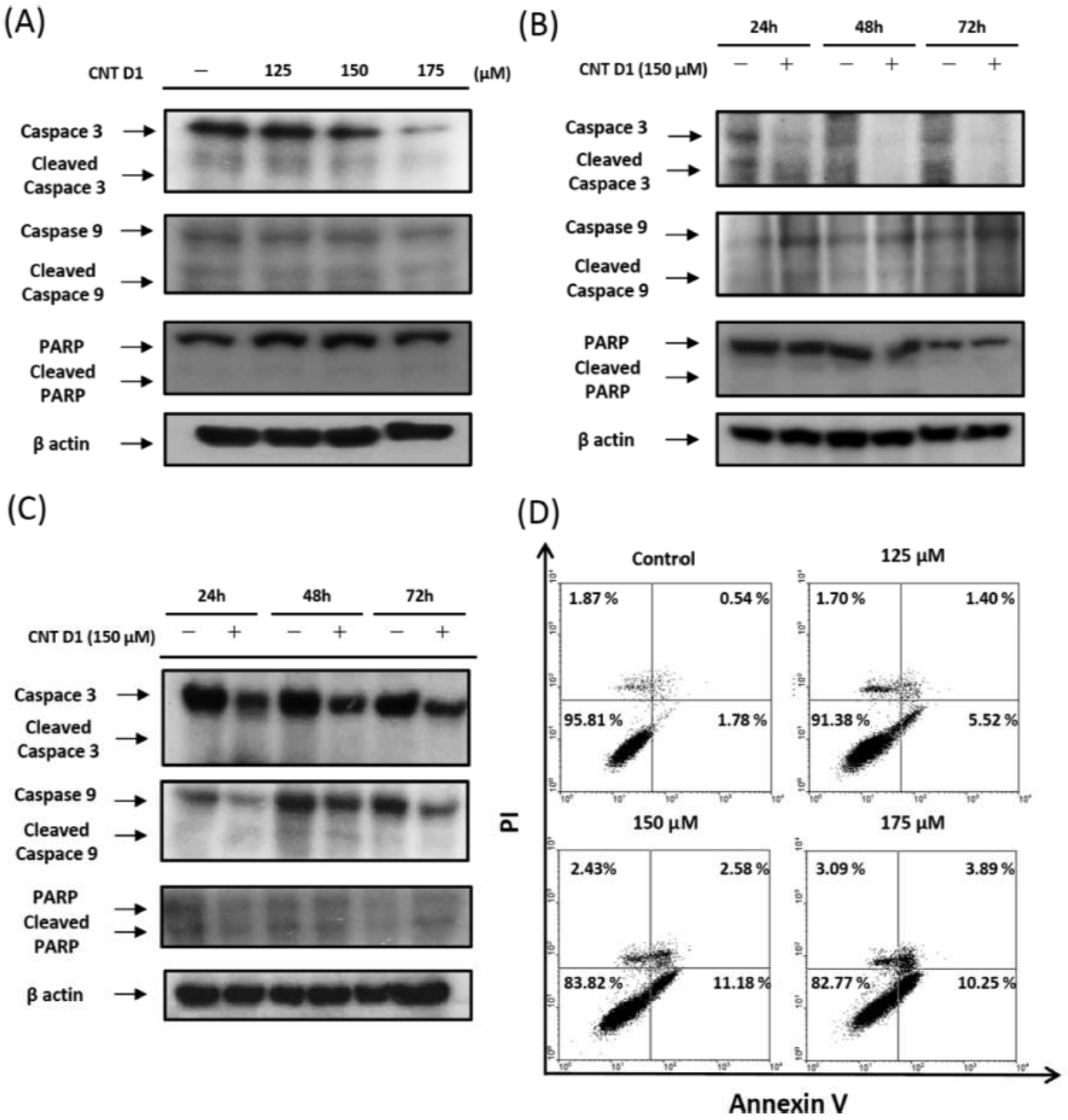


Figure 4. Effect of CNT D1 on cleavage of caspase 3, caspase 9, and PARP. (A) A549 cells treated with CNT D1 at 125, 150, and 175 μM for 72 h. (B) A549 and (C) H460 cells were treated with CNT D1 (150 μM) for the indicated time. The protein expression of caspase 3, cleaved caspase 3, caspase 9, cleaved caspase 9, PARP, and cleaved PARP were measured by Western blot. Western blot data presented are representative of those obtained in at least three separate experiments. The values below the figures represent the change in protein expression of the bands normalized to β-actin. (D) After A549 cells were treated with CNT D1 for 72 h, cells were stained with annexin V-FITC and PI and measured by flow cytometry.

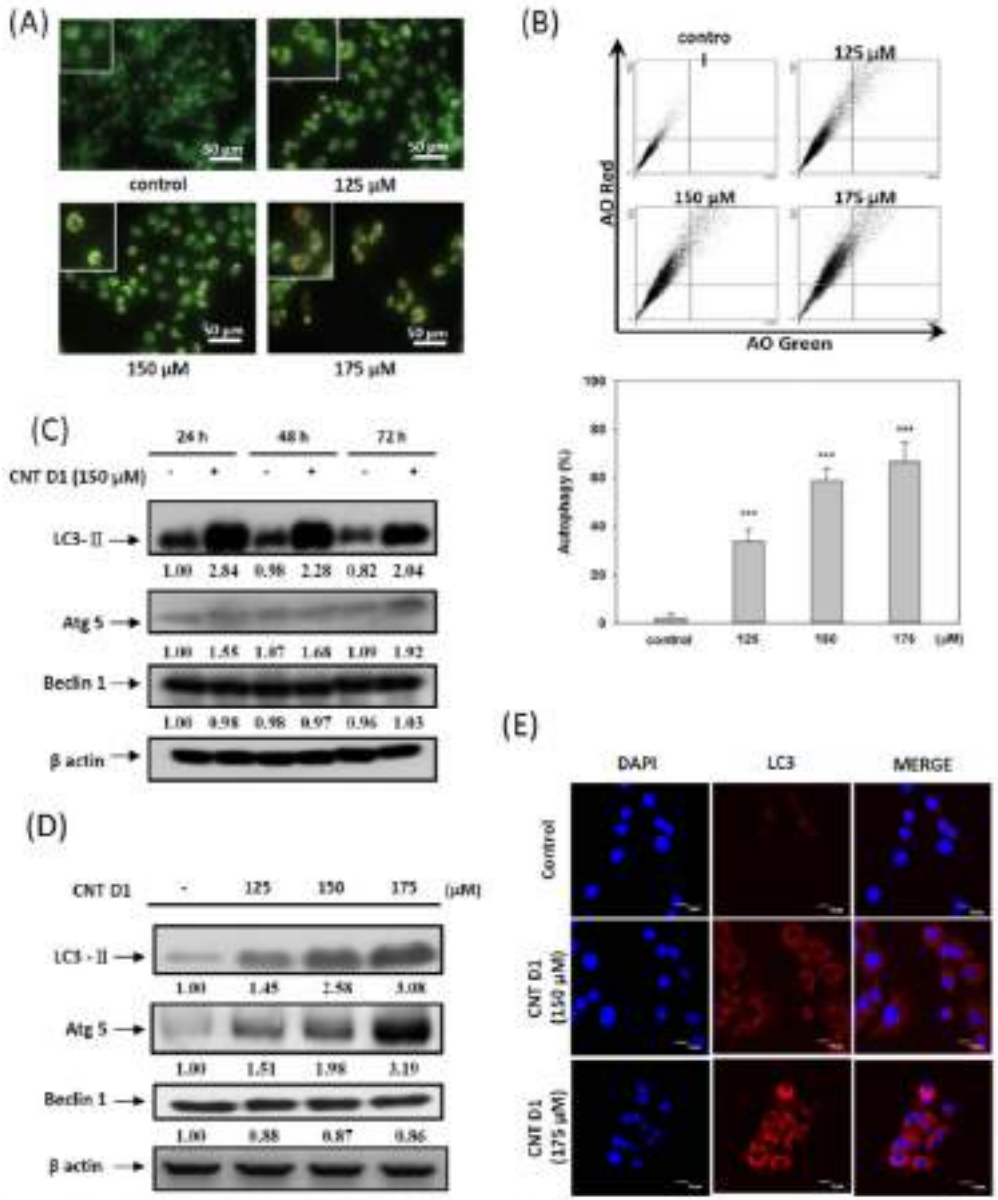


Figure 5. CNT D1-induced autophagy in A549 Cells. (A) A549 cells treated with CNT D1 at 125, 150, and 175 μM for 72 h and stained with AO. The autophagy of cell morphology was observed by fluorescence microscopy. Scale bar: 50 μm. (B) The AO-stained acidic vacuoles on CNT D1-treated A549 cells were measured by flow cytometry. Scale bar: 50 μm. Autophagy data are described as means ± SD, and the statistical significance between the control group and different experimental groups is presented with asterisks (*, $P < 0.05$; **, $P < 0.005$; ***, $P < 0.001$). (C) A549 cells treated with CNT D1 at 125, 150, and 175 μM for 72 h. (D) A549 cells were treated with CNT D1 (150 μM) for the indicated time. The protein expression of LC3 II, Atg 5, and Beclin 1 were measured by Western blot. Western blot data presented are representative of those obtained in at least three separate experiments. The values below the figures represent the change in protein expression of the bands normalized to β-actin. (E) Detecting the LC3 accumulation by immunofluorescence. A549 cells were stained with rabbit monoclonal anti-LC3 and PE-conjugated secondary antibodies (red), and nuclei were stained with DAPI (blue). Results were observed by confocal microscope. Scale bar: 10 μm.

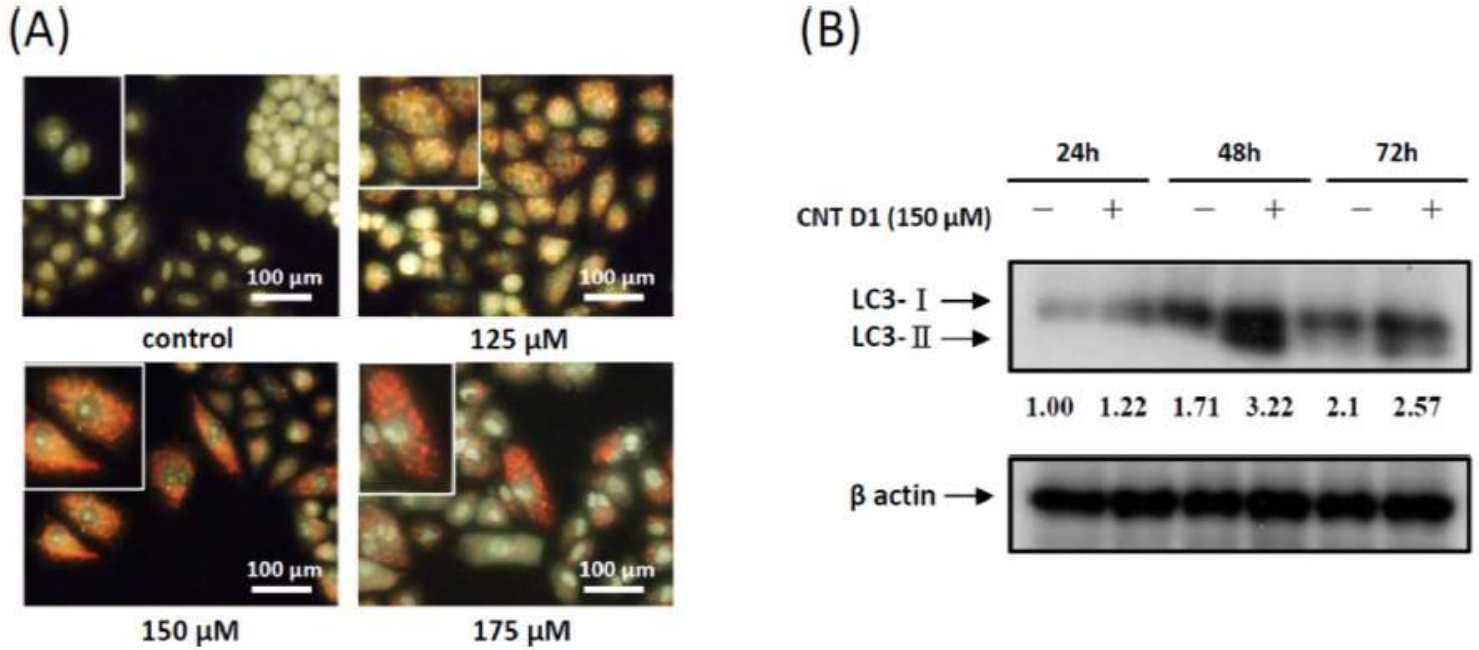


Figure 6. CNT D1-induced autophagy in H460 cells. (A) H460 cells treated with CNT D1 at 125, 150, and 175 μM for 72 h and stained with AO. The autophagy of cell morphology was observed by fluorescence microscopy. Scale bar: 100 μm. (B) H460 cells were treated with CNT D1 (150 μM) for the indicated time. The protein expression of LC3 II was measured by Western blot. Western blot data presented are representative of those obtained in at least three separate experiments. The values below the figures represent change in protein expression of the bands normalized to β-actin.

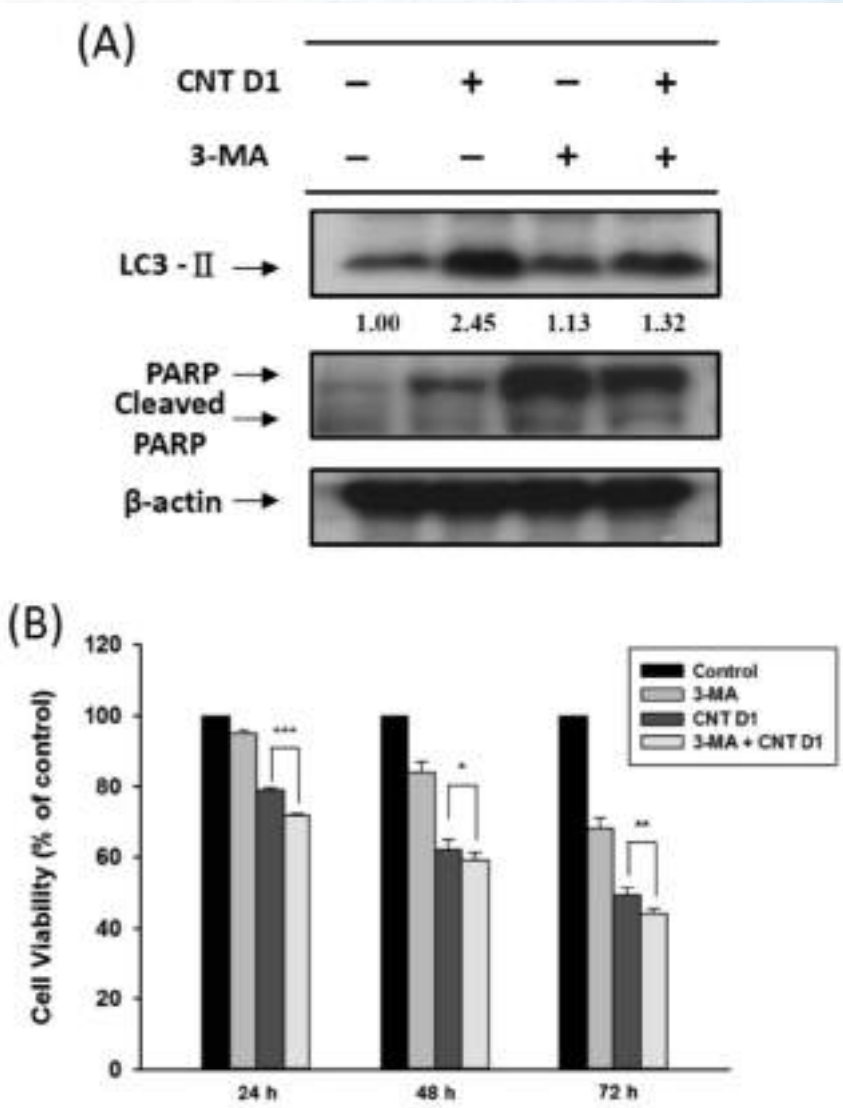


Figure 7. Autophagy inhibition by 3-MA. After A549 cells were pretreated with 1 mM 3-MA for 1 h, they were cotreated with 150 μM CNT D1 and 1 mM 3-MA for 72 h. (A) Protein expression of LC3 I, LC3 II, PARP, and cleaved PARP was determined by Western blot. (B) Cell viability was measured by MTT assay. Results are described as means ± SD, and between the CNT D1 group and 3-MA + CNT D1 group, the statistical significance is presented with asterisks (*, $P < 0.05$; **, $P < 0.005$; ***, $P < 0.001$).

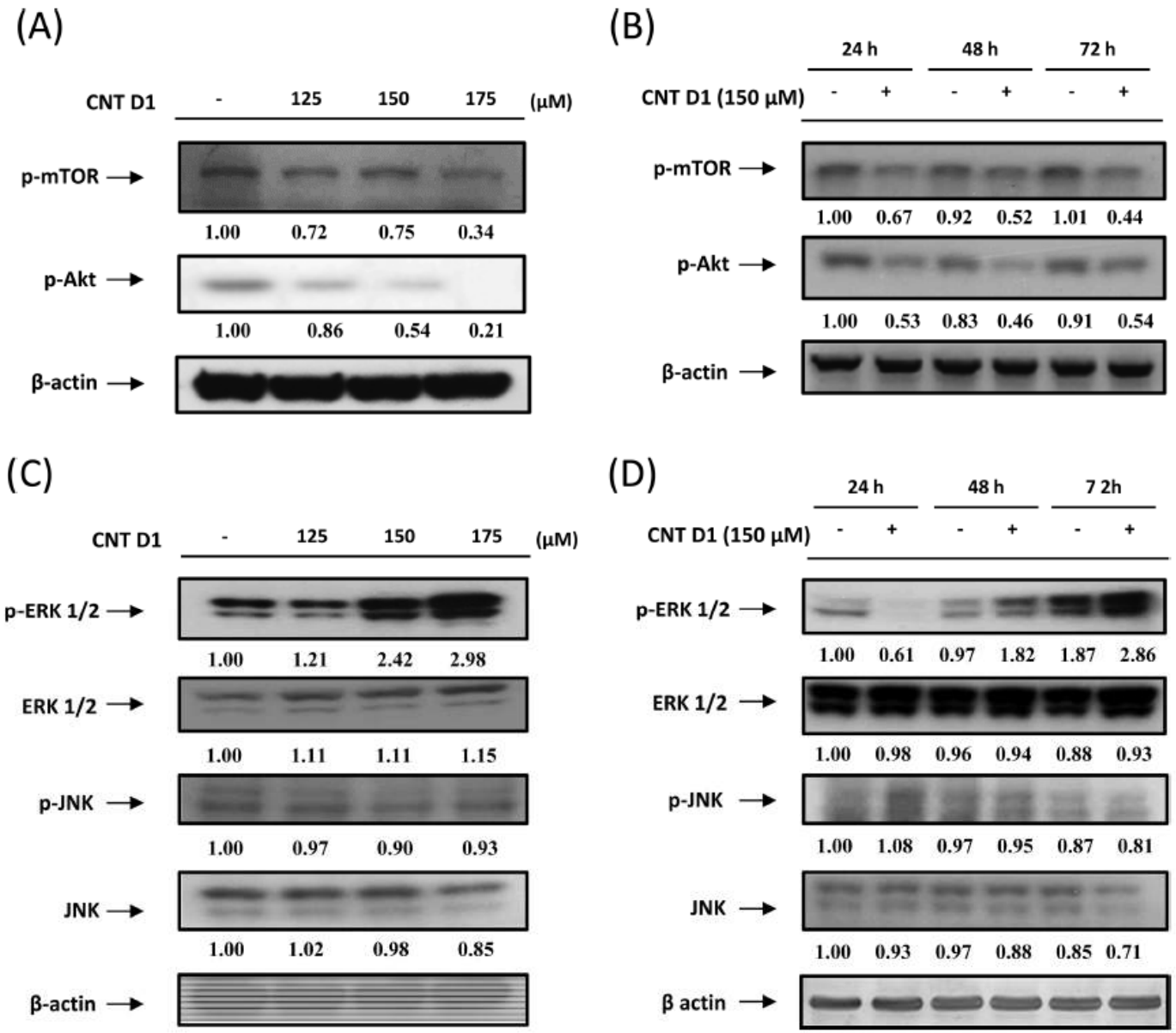
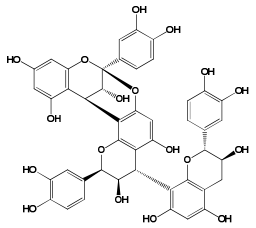


Figure 8. CNT D1 downregulated Akt/mTOR phosphorylation and upregulated ERK1/2 phosphorylation in A549 cells. (A) A549 cells treated with CNT D1 at 125, 150, and 175 μM for 72 h. (B) A549 cells were treated with CNT D1 (150 μM) for the indicated time. The protein expression of Akt and mTOR phosphorylation was measured by Western blot. (C) A549 cells treated with CNT D1 at 125, 150, and 175 μM for 72 h. (D) A549 cells were treated with CNT D1 (150 μM) for the indicated time. The protein expression of ERK1/2 and JNK phosphorylation was measured by Western blot. Western blot data presented are representative of those obtained in at least three separate experiments. The values below the figures represent the change in protein expression of the bands normalized to β-actin.



Cinnamtannin D1

NSCLC cells

Atg5 dependent

Akt/mTOR pathway

ERK1/2 pathway

p53, p27

CDK 6, Cyclin D1

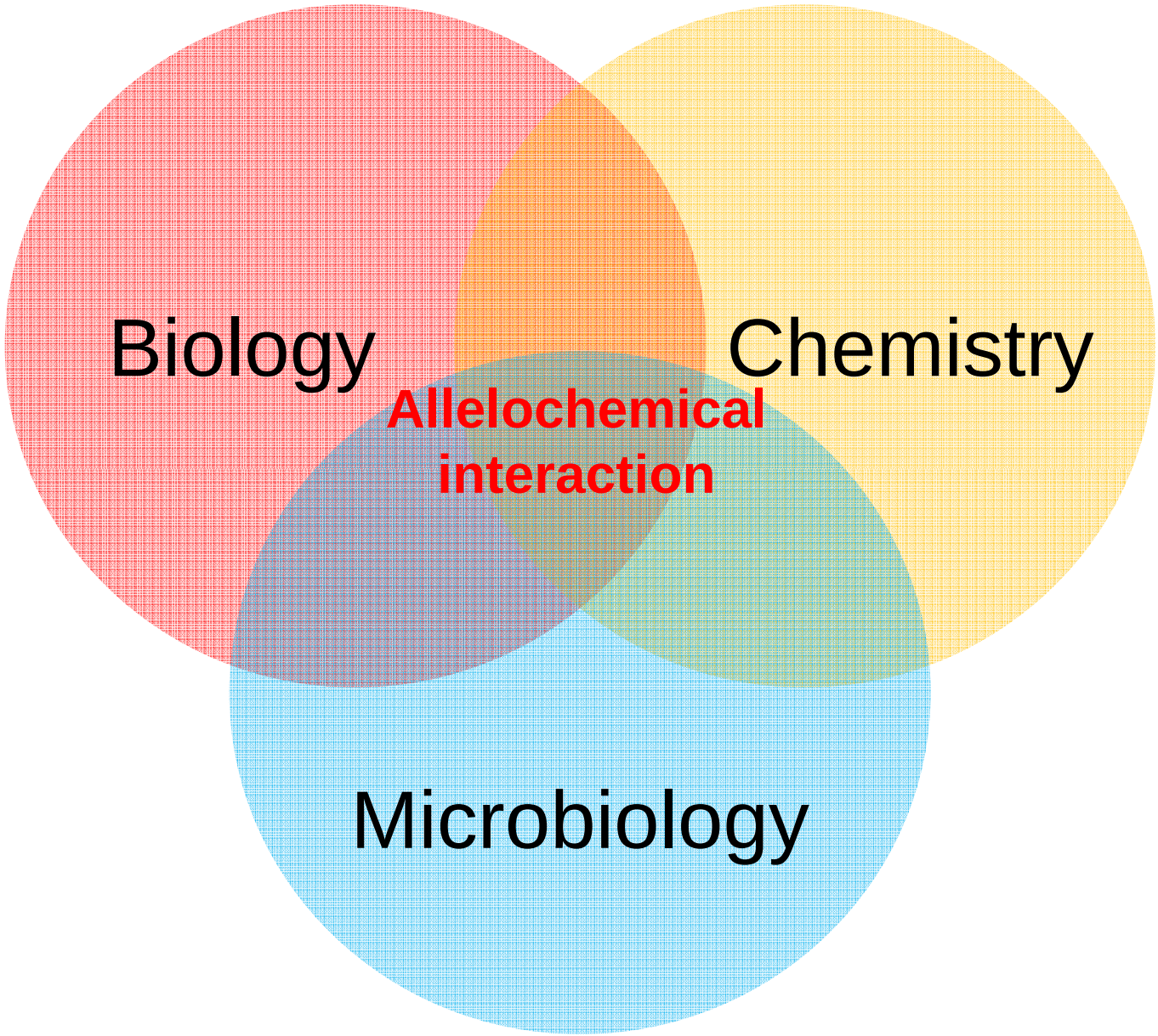
Autophagy (cytoprotection)

3-MA (autophagy inhibitor)

Apoptosis

G1-phase cell cycle arrest

Study on Chemical Ecology Needs Multiple Disciplines



Conclusions

- ***Rhododendron formosanum*** **released varieties of natural products, such as fatty acids, phenolics, flavonoids and terpenoids, etc.**
- **The aforementioned natural products possess allelopathic, allelochemical, and pharmaceutical properties.**
- **(-)-catechin and its polymers play a major role in allelopathy and pharmaceutical**
- **Catechin converted to protocatechuic acid through microbial transformation plays the most important role in allelopathic effect upon the growth of understory species**

Conclusions

- **Triterpenoids, namely ursolic acid, oleanolic acid, and betulinic acid, perform anticancer activity on non-small-cell lung carcinoma cells at low concentration of 20 μM.**
- **Procyanidins perform anticancer activity on lung cancer cells and antibacterial activity against certain pathogens.**
- **All natural products indicated above are highly potential to be developed into herbicides, fungicide, antibiotics, and pharmaceutical usage.**
- **Natural products from *Rhododendron formosanum* exhibit diverse functions of biodiversity which benefits to human well-being**

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- National Science Council, Taiwan
- Research center for biodiversity, China Medical University


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