College of Medicine & Health Sciences @ Sultan Qaboos University



Mesenchymal Stem Cells to Repair Vascular Damage after Chemotherapy: Past, Present and Future

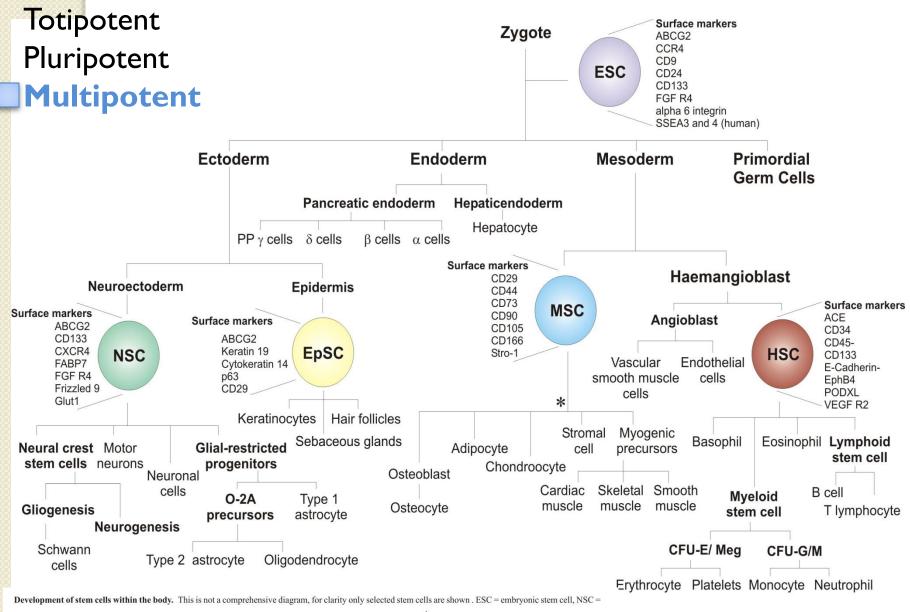
Cell Therapy 2014 Las Vegas, NV, USA

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What are MSCs?

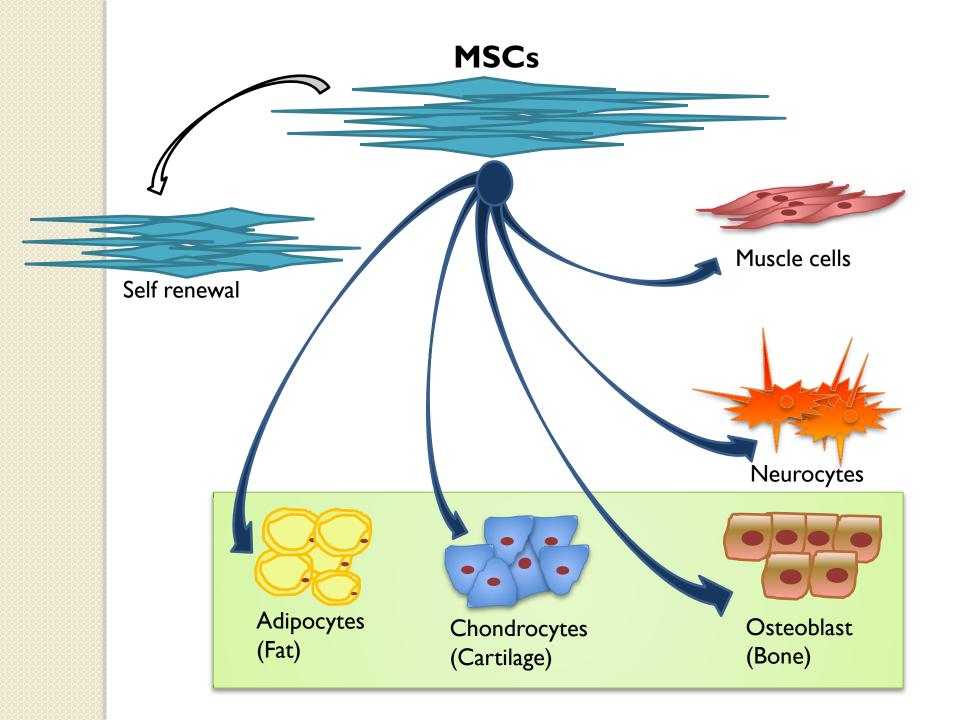
- Stem cell capacity
- Stromal property
- Non-hematopoietic cells
- Multipotent cells
- 0.001 0.01% of nucleated cells in BM
- Adhere to culture plate
- Express variable range of cell markers



 $neuronal stem \ cell, \ EpSC = epidermal \ stem \ cell, \ MSC = mesenchymal \ stem \ cell, \ HSC = haematopoeitic \ stem \ cell. \ * Differentiation \ of \ MSCs \ along \ neuronal \ lineages \ along \ neuronal \ ne$

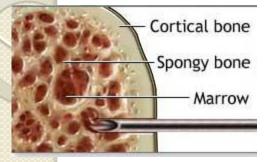
 $has also been demonstrated, see text for information. \ Modified from R\&D \ Systems \ website (http://www.rndsystems.com). \ Copyright BTR @ the second se$

http://www.york.ac.uk/res/bonefromblood/background/osteogenesis.html

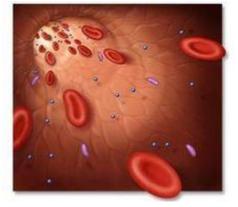


MSC Sources

Bone marrow



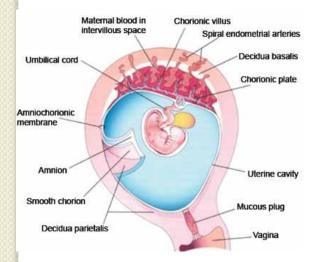
Peripheral blood



Cord blood



Placenta and fetal membrane



Adipose tissue



Common MSC Markers

| Negative Markers | Roles | | |
|-----------------------|---|--|--|
| CD34 | Primitive hematopoietic cells and endothelial cells | | |
| CD45 | Leukocytes | | |
| CDIIb and CDI4 | Monocytes and macrophages | | |
| CD 79α & CD19α | B cells | | |
| HLA Class II | Antigen presenting cells and lymphocytes | | |
| Positive Markers | | | |
| CD73/5'-Nucleotidase | Catalyzes production of extracellular adenosine from AMP | | |
| CD90/Thy I | Wound repair, cell-cell and cell-matrix interactions | | |
| CD I 05/Endoglin | Vascular homeostasis; modulates TGF- β functions via interaction with TGF- β receptors (RI & RII) | | |

MSC Clinical Applications

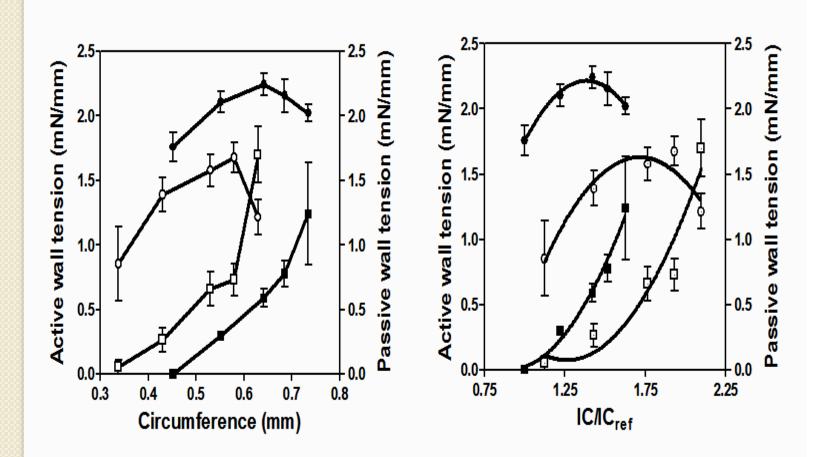
Hematopoietic Stem Cell Transplantation Enhance engraftment Reduce graft versus host disease Solid Organ Transplantation Improved graft function **Reduced** rejection Repair Vascular Damage Chemotherapy **Radiotherapy** Others

Chemotherapy and Vascular Toxicity

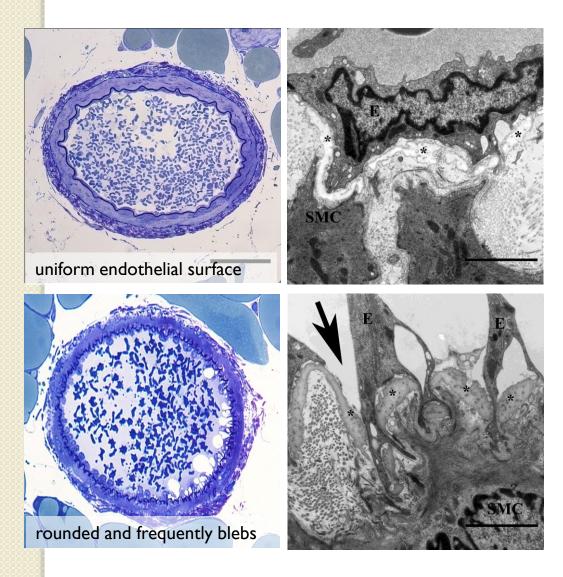
TABLE 1. Cardiotoxicity Profiles of Chemotherapeutic Agents

| | | - | | |
|--|---|--|---|---|
| Drug Class/Name, Generic (Brand) | Cardiac Adverse Events | Relative Frequency of Specific Adverse Effect* | Relative Frequency of Therapeutic Use† | Comment |
| Anthracyclines/anthraquinolo | ones | | | |
| Doxorubicin (Adriamycin) Daunorubicin (Cerubidine) Epirubicin (Ellence, Pharmorubicin) Idarubicin (Idamycin) | CHF/LV dysfunction | +++ | +++ | Risk of CHF is cumulative dose and schedule dependent; LV dysfunction is secondary to free radical production; increased risk for young/elderly, after mediastinal XRT, female gender, history of cardiac disease; continuous infusion, liposomal delivery systems, or use of dexrazoxane can reduce toxicity; when appropriately administered, incidence of LV dysfunction is $<\!5\%$ |
| Mitoxantrone (Novantrone) | CHF/LV dysfunction | ++ | + | Anthraquinone derivative; low propensity for free radical production; myocarditis and arrhythmia can be seen acutely with infusion |
| Alkylating agents | | | | |
| Busulfan (Myleran) 🗲 🗕 | Endomyocardial fibrosis Cardiac tamponade | +++++ | + | |
| Cisplatin (Platinol) | lschemia Hypertension | ++ ++++ | +++ | |
| | CHF | ++ | | CHF risk is increased in elderly, after chest XRT, or after prior anthracyclines |
| Cyclophosphamide < (Cytoxan) | Pericarditis/ myocarditis CHF | +++ | +++ | Rare incidence of hemorrhagic myocarditis, more common with high dose CHF risk is increased with cumulative dose, in elderly, after chest XRT, or after prior anthracyclines |
| lfosfamide (lfex) | CHF Arrhythmias | ++ ++ | ++ | CHF risk is increased with cumulative dose, prior anthracyclines |
| Mitomycin (Mutamycin) | CHF | ++ | + | CHF risk is increased with cumulative dose, prior anthracyclines, chest XRT |
| | | | | |

Bu-Cy and Vascular Toxicity



Bu-Cy and Vascular Toxicity



Control •Unbroken endothelial cell-cell contact

Bu-CyDetaching of endothelial cellsCell-cell contacts were disrupted

Chemotherapy & Vascular Toxicity

- Cisplatin-based therapy
- Testicular cancer patients
- Before and after chemotherapy

| | | | | Tab | IC 2. VO | | Cinto | | |
|---------|---------|-----------------------|------------------|--------------------------------------|----------|----------------------|-------|-------------|-------|
| | Age | | | Platelets* (× 10 ⁹ /L) | | Fibrinogen* (g/L) | | ∨WF* (%) | |
| Patient | (years) | Event | Time | Before | After | Before | After | Before | After |
| 1 | 31 | MI | Day 15, course 2 | 393 | 673 | 5.0 | 6.4 | 170 | 339 |
| 2 | 37 | MI | Day 9, course 1 | 378 | 667 | 4.8 | 4.4 | 185 | 280 |
| 3 | 34 | DVT (leg) | At diagnosis | 348 | 264 | 3.9 | 3.1 | 104 | 139 |
| 4 | 23 | DVT (subclavian vein) | Day 22, course 4 | 239 | 117 | NE |) | 78 | 95 |
| 5 | 31 | DVT (subclavian vein) | Day 9, course 2 | 217 | 270 | NE |) | 86 | 140 |
| 6 | 28 | PE | Day 11, course 4 | 285 | 196 | 4.6 | 3.4 | 88 | 173 |
| 7 | 22 | PE | Day 15, course 2 | 755 | 245 | 7.3 | 4.4 | 133 | 145 |

Table 2. Vascular Events

MI, myocardial infarction; DVT, deep vein thrombosis; PE, pulmonary embolism; vWF, von Willebrand factor

MSC Source

• Rat BM

Injury induction

- Ligation of proximal left coronary artery
 - MI

MSC infusion

Infarct area (2 x 10⁶)

- Expressed muscle and endothelium markers
- Improved LV function

MSC Source

• Canine BM

Injury induction

- Ameroid constrictor placement
 - Chronic myocardial ischemia
- MSC infusion
 - Intramyocardial injections (10 x 10⁷)

- Improved ejection fraction & vascular density
- Increased vascularity
- Improved cardiac function
- MSC Differentiation
 - SMCs & ECs

MSC Source

- Porcine BM
- Injury induction
 - Occlusion followed by reperfusion of the anterior coronary artery
 - MI

MSC infusion

Intramyocardial injections (20×10^7)

- Improved myocardial blood flow
- Improved ventricular & late cardiac functions
 - Enhanced blood vessels maturation
 - Reduced endomyocardial apoptosis
 - Reduced infarct scarring

MSC Source

Porcine BM

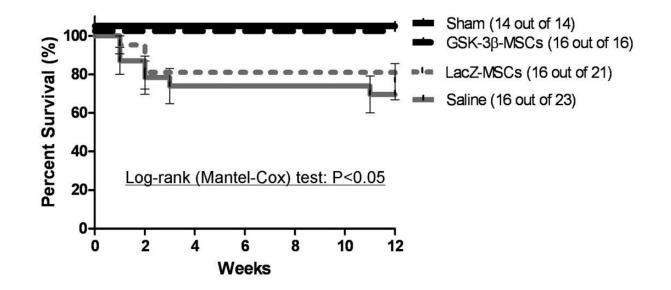
Injury induction

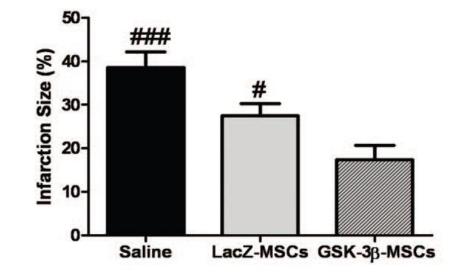
Carotid artery cannulation

- MI
- MSC infusion
 - Transendocardial injections (10 x 10⁷)

- Differentiated into cardiomyocytes and vascular structures
- Stimulated endogenous cardiac stem cells proliferation and differentiation
- Stimulated cardiomyocyte replication
- Reduced infarct size

MSC Source C57BI/6 mice BM • Express GSK-3 β Injury induction Carotid artery ligation • MI MSC infusion Border zone of MI (1.5x 10⁵) Outcomes • GSK-3 β increased MSCs survival GSK-3 β -MSCs induced cardiomyocyte differentiation and angiogenesis GSK-3 β -MSCs increased capillary density GSK-3 β -MSCs upregulated paracrine factors (VEGF-A) 0





MSC Source

C57BI/6 mice BM

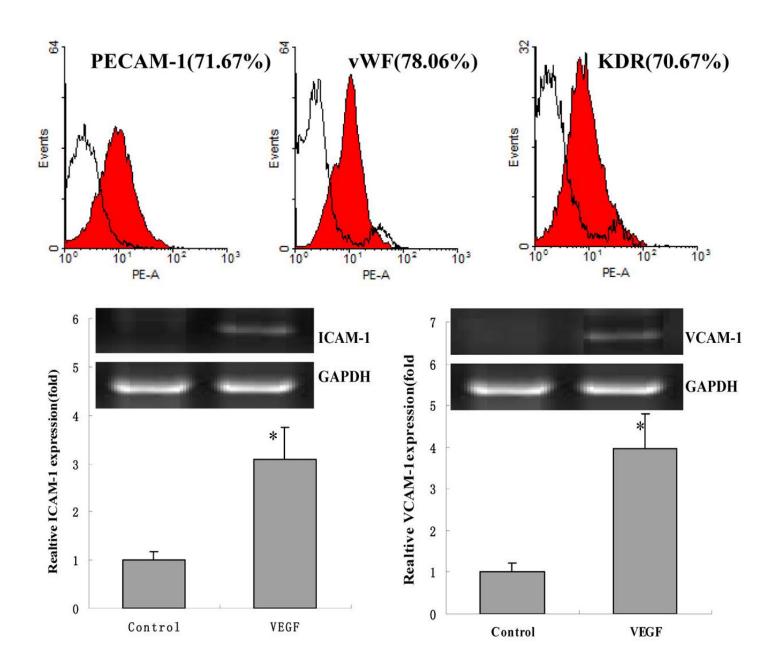
Injury induction

- Removal of endothelium with a flexible wire
 - Carotid artery injury

MSC infusion

• IV injection $(I \times I0^5)$

- Differentiated into neo-ECs in the injury
- Contributed to vascular remodeling
 - Expressed different markers
- Formed the tube-like structure



MSCs in clinical practice (Meta Analysis)

- Searched databases
 - PubMed, OVID, EMBASE, the Cochrane Library, and ClinicalTrials.gov
- Outcome
 - Improved LVEF in patients
- Efficacy of Cells in transplantation influenced by
 - Source of cells
 - Route of infusion
 - Type of injury
- BMCs & MSCs infusion is a safe and effective therapy to improve vascular repair

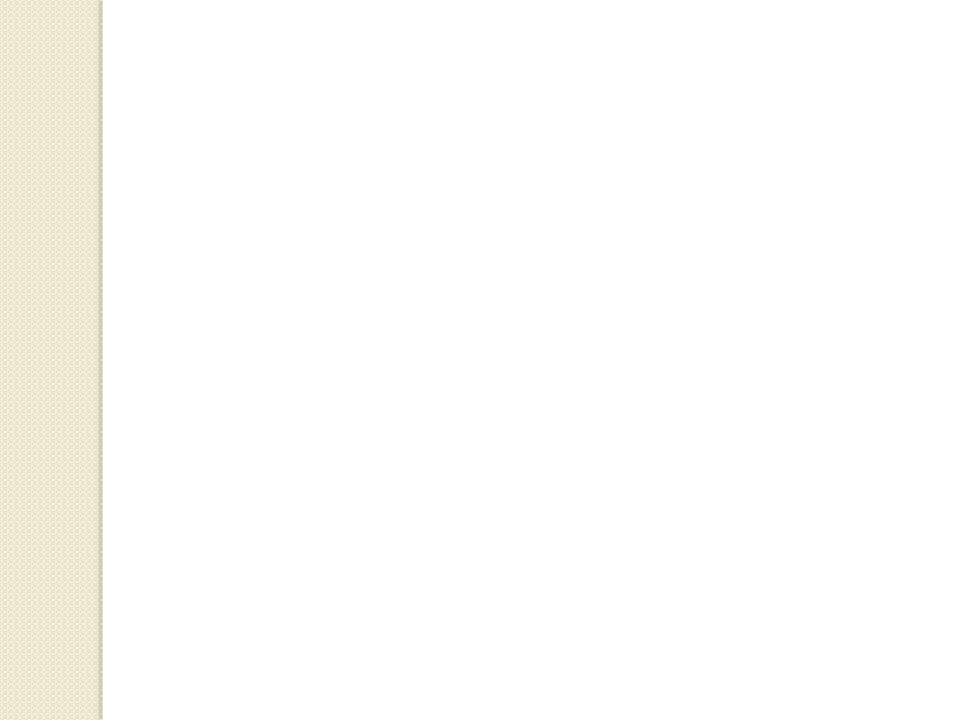
Precautions

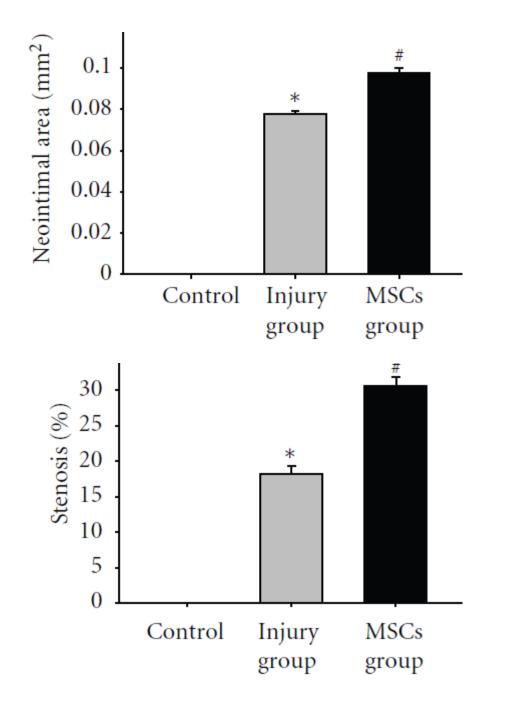
- Intra-myocardial calcification
- Vascular calcification
- Increase aortic stiffness
- Systolic hypertension
- Transient improvement
- Unregulated differentiation

Prospective

- MSC therapy under intensive investigation
 Establish clinical relevant vascular injury models
 Sources & doses
 - Timing
 - Protocols
- Collaboration
 - Type of disease
 - Data sharing

THANKS





Liao et al 2012

MSCs in clinical practice

TABLE 3: Ongoing clinical trials on MSCs: condition, intervention/dose, and followup in patients around the world (http://www.clinicaltrials.gov).

| World | Condition | Intervention | Time frame | Phase/Status |
|--------------------|---|---|-----------------|----------------------|
| | Chronic ischemic LV dysfunction secondary to MI | 10 and 20 intramyocardial injections of 2 million MSCs (low dose) or 20 million (high dose)/0.25–0.5 cm ³ for a total of 20 million or 200 million cells, respectively | 6–18 months | Phase I/II (unknown |
| Florida (IISA) | Chronic ischemic IV dysfunction and heart failure secondary to MI | Transendocardial injection of autologous human cells (bone marrow or mesenchymal). 40 million cells/mL delivered in either a dose of 0.25 mL/injection for a total of 100 million \times 10 injections or a dose of 0.5 mL/injection for a total of 200 million \times 10 injections | 6–18 months | Phase I/II (unknown |
| Florida (USA) | Chronic ischemic IV dysfunction secondary to MI | Transendocardial injection of autologous versus allogeneic MSCs. 40 million cells/mL delivered in either a dose of $0.5 \text{ mL/injection} \times 1$ injection for a total of 20 million, a dose of $0.5 \text{ mL/injection} \times 5$ injections for a total of 100 million, or a dose of $0.5 \text{ mL/injection} \times 10$ injections for a total of 200 million MSCs | 6–13 months | Phase I/II (active) |
| | Nonischemic dilated cardiomyopathy | Transendocardial injection of autologous versus allogeneic MSCs. 20 million cells/mL delivered in a dose of $0.5 \text{ mL/injection} \times 10$ injections for a total of 100 million of MSCs | 6–12 months | Phase I/II (active) |
| Maryland (USA) | Chronic ischemic IV dysfunction secondary to MI | 10 and 20 intramyocardial injections of 2 million MSCs (low dose) or 20 million (high dose)/0.25–0.5 cm ³ for a total of 20 million or 200 million of autologous human MSCs, respectively | 6–18 months | Phase I/II (unknown |
| | Chronic ischemic LV dysfunction secondary to MI | Transendocardial injection of autologous versus allogeneic MSCs. 40 million cells/mL delivered in either a dose of 0.5 mL/injection $\times 1$ injection for a total of 20 million, a dose of 0.5 mL/injection $\times 5$ injections for a total of 100 million, or a dose of 0.5 mL/injection $\times 10$ injections for a total of 200 million MSCs | 6–13 months | Phase I/II (active) |
| France (Europe) | Chronic myocardial ischemia; LV dysfunction | Transendocardial intramyocardial injections of 60 million autologous MSCs | 30 days–2 years | Phase I/II (active) |
| China (East Asia) | ST-elevation MI | Intracoronary human umbilical WJ-MSC transfer | 4 months–1 year | Phase II (active) |
| Korea (East Asia) | Acute MI | Intracoronary injection of single dose of autologous bone-marrow-derived MSCs (I million) cells/kg | 6 months | Phase II (completed) |
| India (South Asia) | ST-elevation acute MI | A Single Dose of Intravenous infusion of Allogenic MSCs | 6 months | Phase I/II (active) |
| | | | | |

Elnakish et al 2012

| Cultivation | Adherence to plastic in standard culture conditions | | | |
|----------------------------------|---|--|--|--|
| Phenotype | Positive expression (≥95%) CD 73 CD 90 CD 105 | Negative expression (≤2%) CD 14 or CD 11b CD 19 or CD 79α CD 34 CD 45 HLA-DR | | |
| <i>In vitro</i> differentiation: | Under specific stimulus, cells will differentiate into osteoblasts, adipocytes, and chondroblasts | | | |

Table 2. Basic criteria for defining human MSCs

Cancer Gene Therapy (2014), 12 – 23