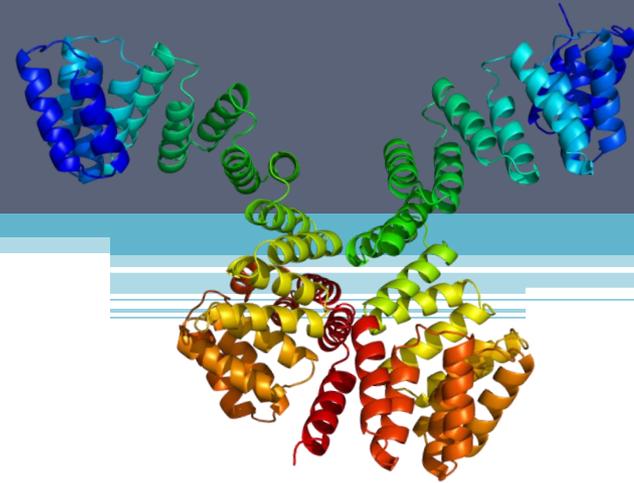
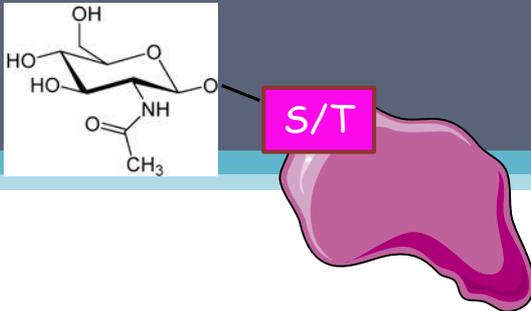


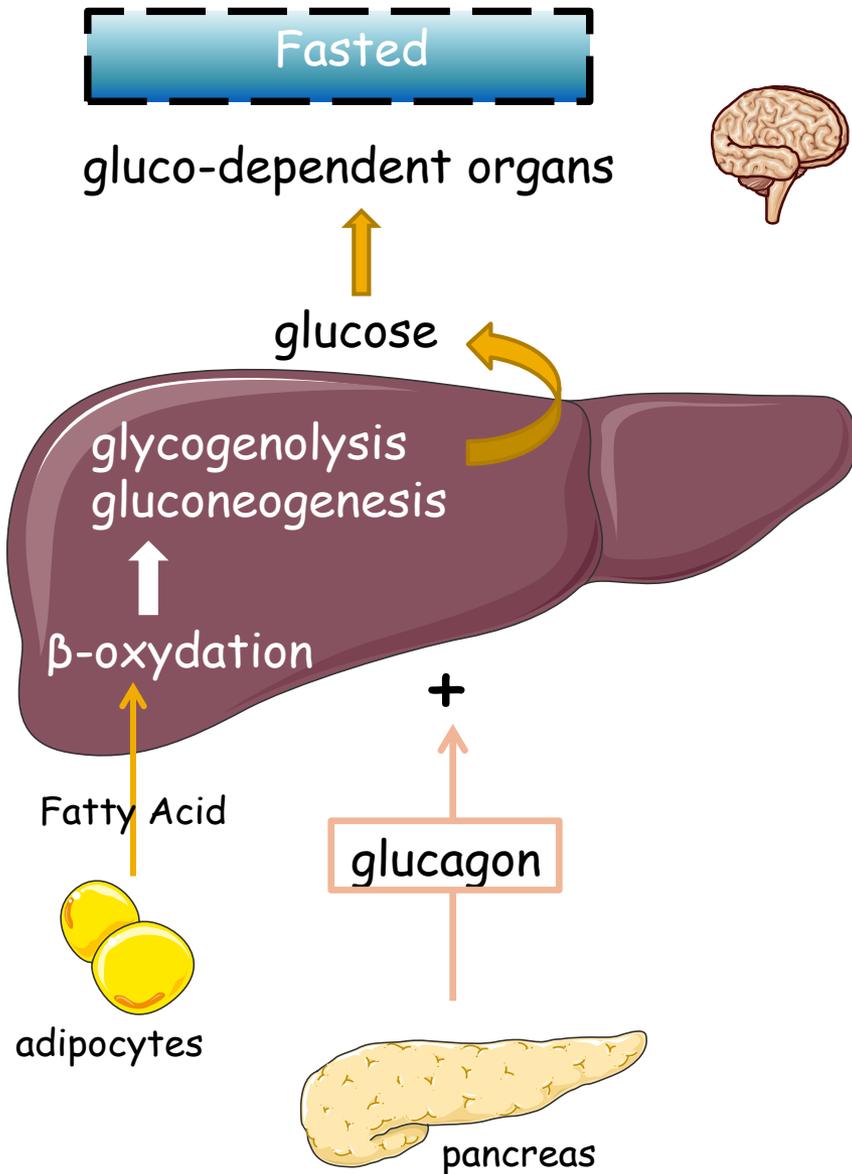
Regulation of hepatic Fatty Acid Synthase properties by O-GlcNAcylation *in vivo* and *ex vivo*



Baldini Steffi, Anne-Marie Mir, Marlène Mortuaire, Céline Guinez and Tony Lefebvre

CNRS-UMR 8576, Unit of Structural and Functional Glycobiology, FRABio FR3688, Lille 1 University, 59655 Villeneuve d'Ascq, France

Control of blood glucose by the liver



Hepatic glucose production

Control of blood glucose by the liver

Fasted

In the fed state

gluco-dependent organs



Glucose absorption



glucose

glucose

glycogenolysis
gluconeogenesis

glycogenogenesis
glycolysis

β -oxydation

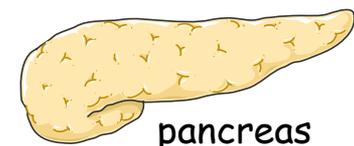
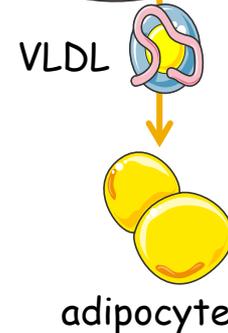
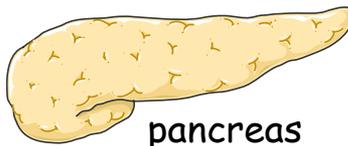
lipogenesis

Fatty Acid

glucagon

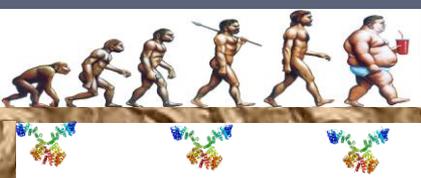
VLDL

insulin

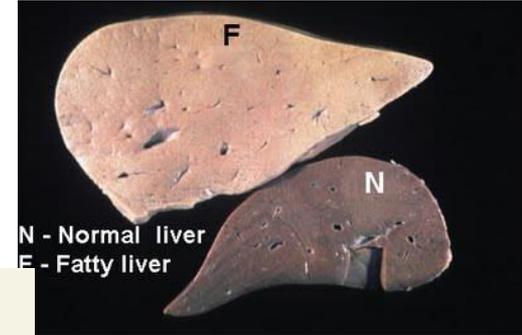
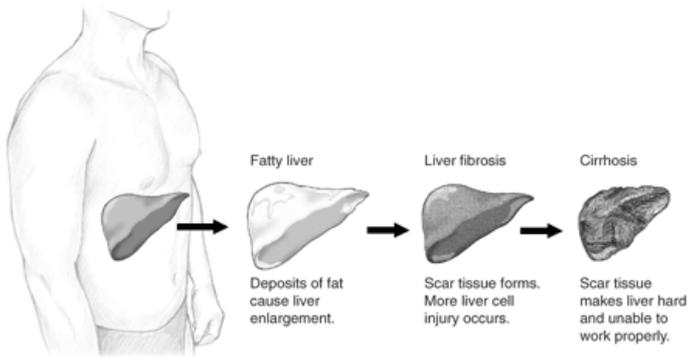


Hepatic glucose production

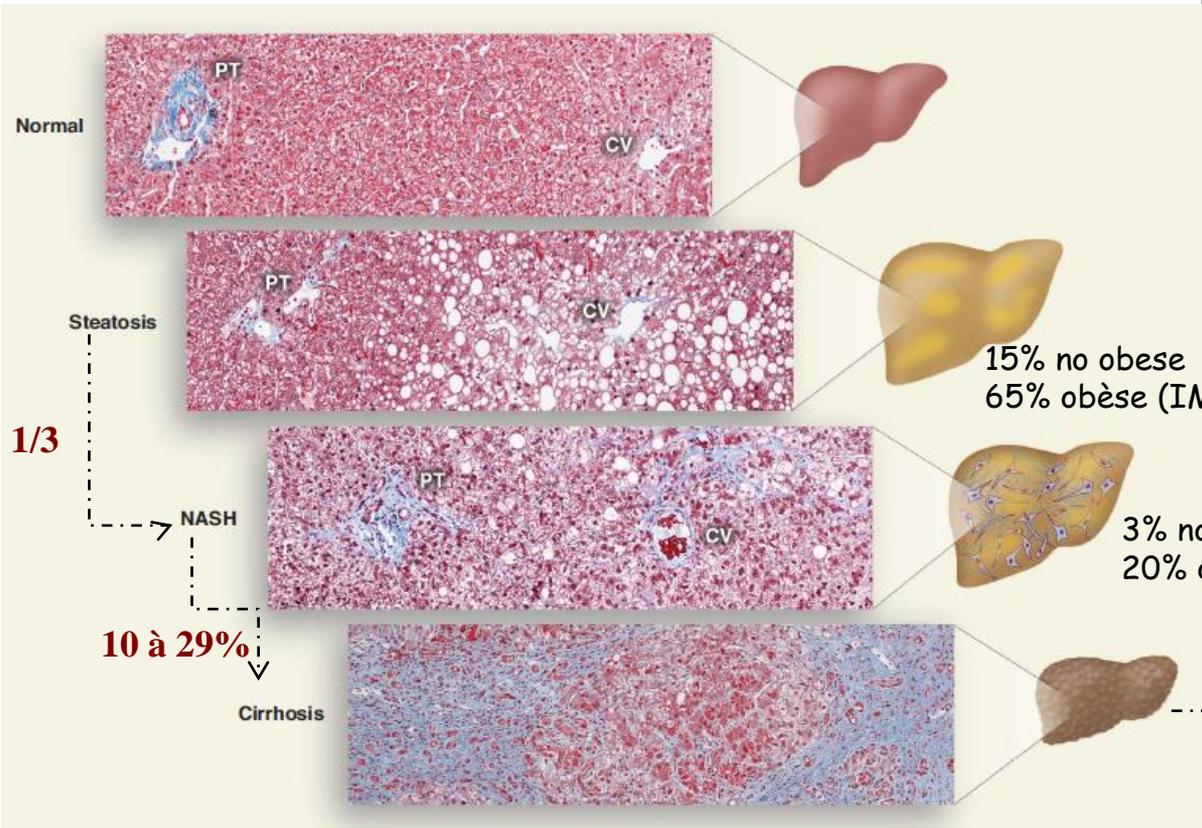
Hepatic glucose uptake



Dysregulation of glucido-lipidic metabolism and hepatic steatosis



The fat represents at least 5 to 10% of the liver weight



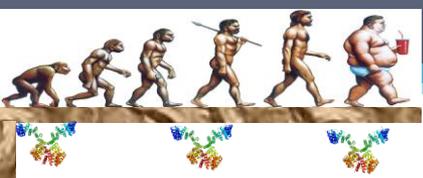
15% no obese
65% obese (IMC 30-40)

3% no obese
20% obese

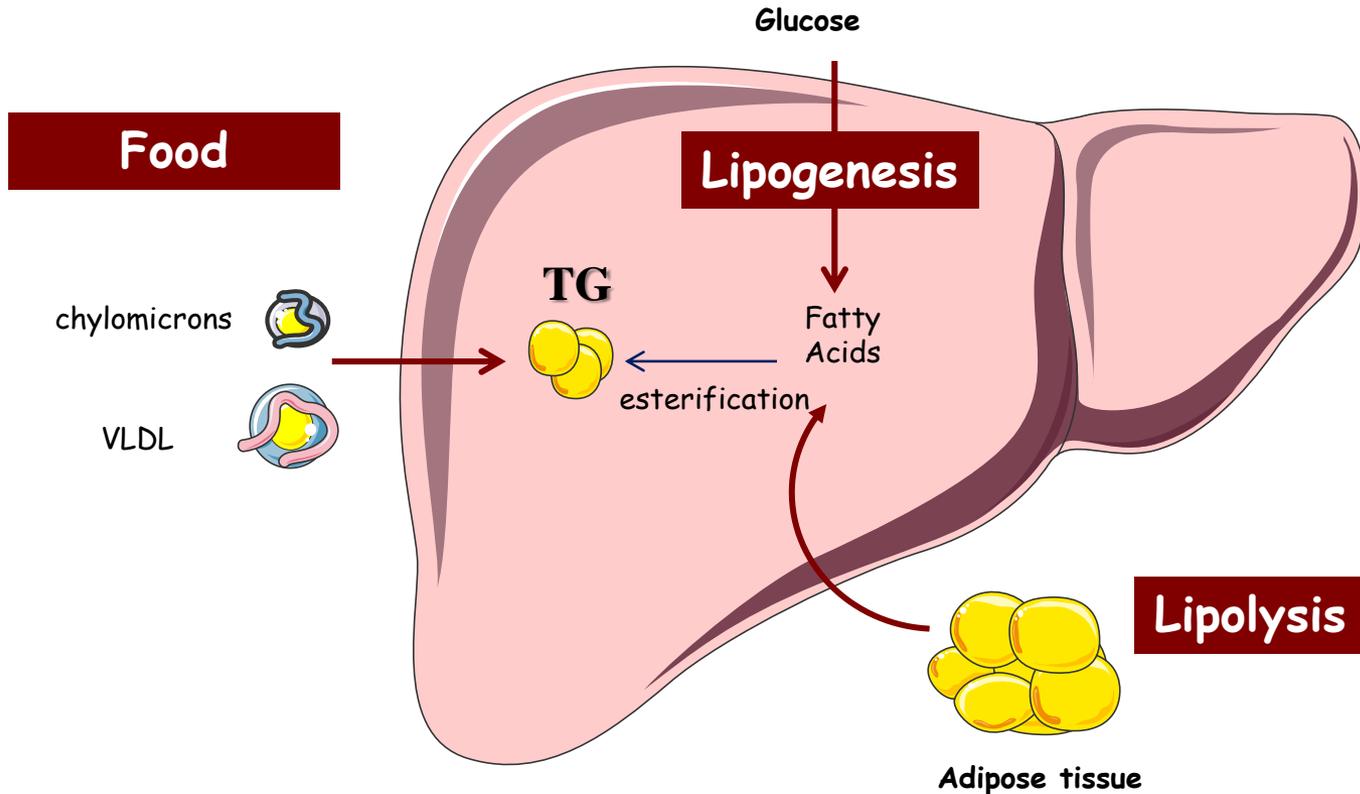
4 à 27% → **CHC**

1/3

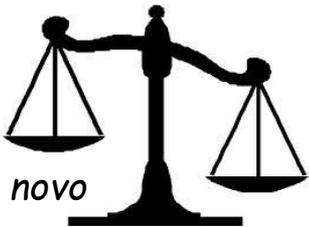
10 à 29%



The causes of hepatic steatosis

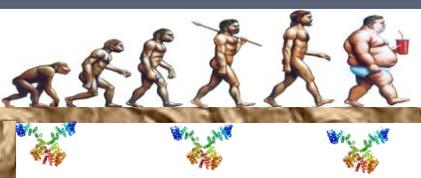


Supply

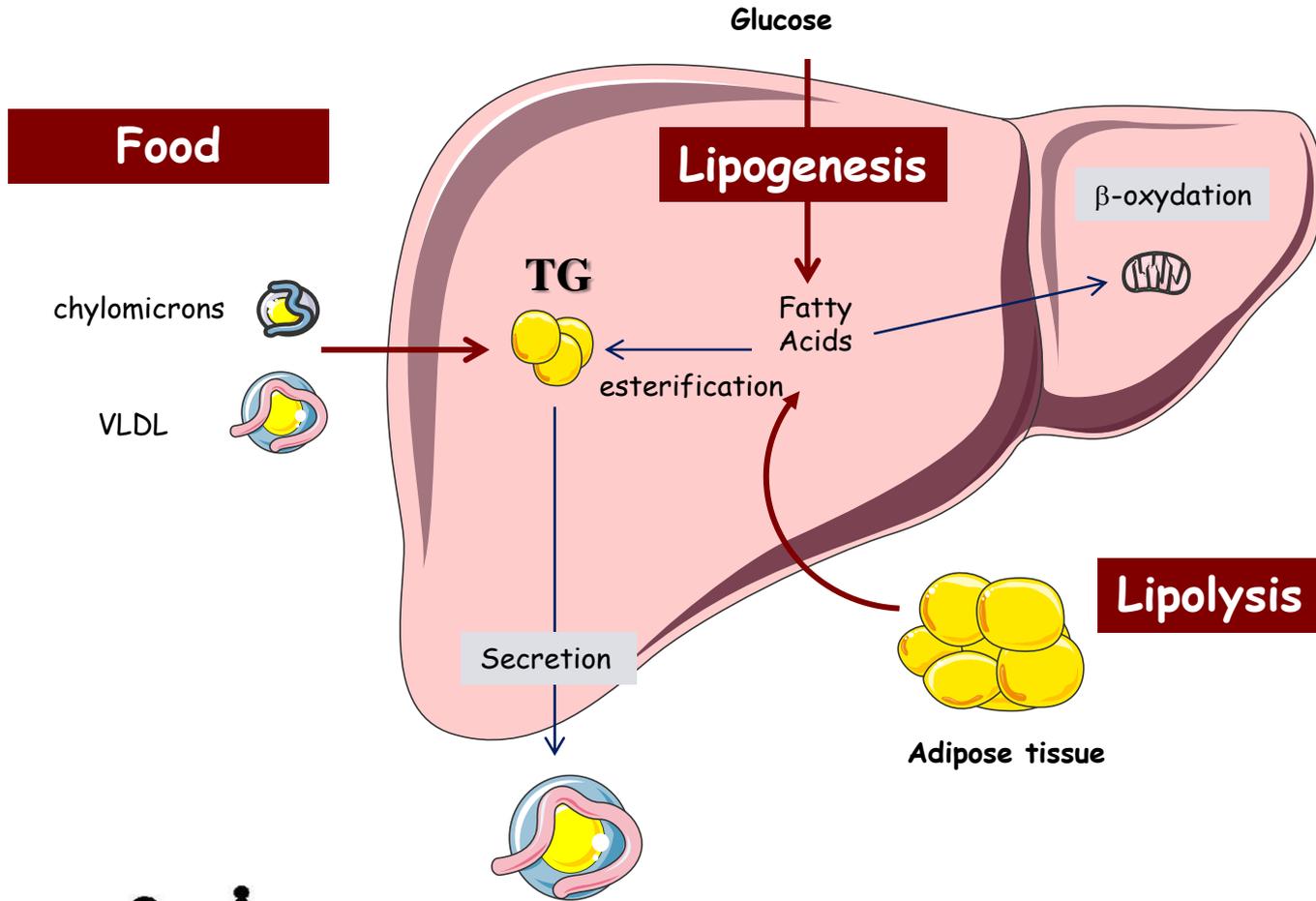


Lipolysis
Lipogenesis de novo
Food

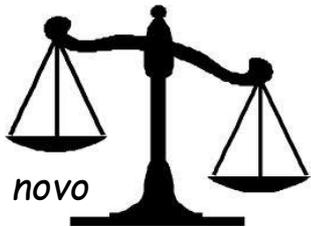
STEATOSIS



The causes of hepatic steatosis



Supply

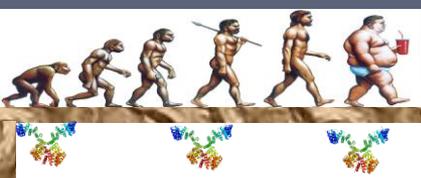


Use

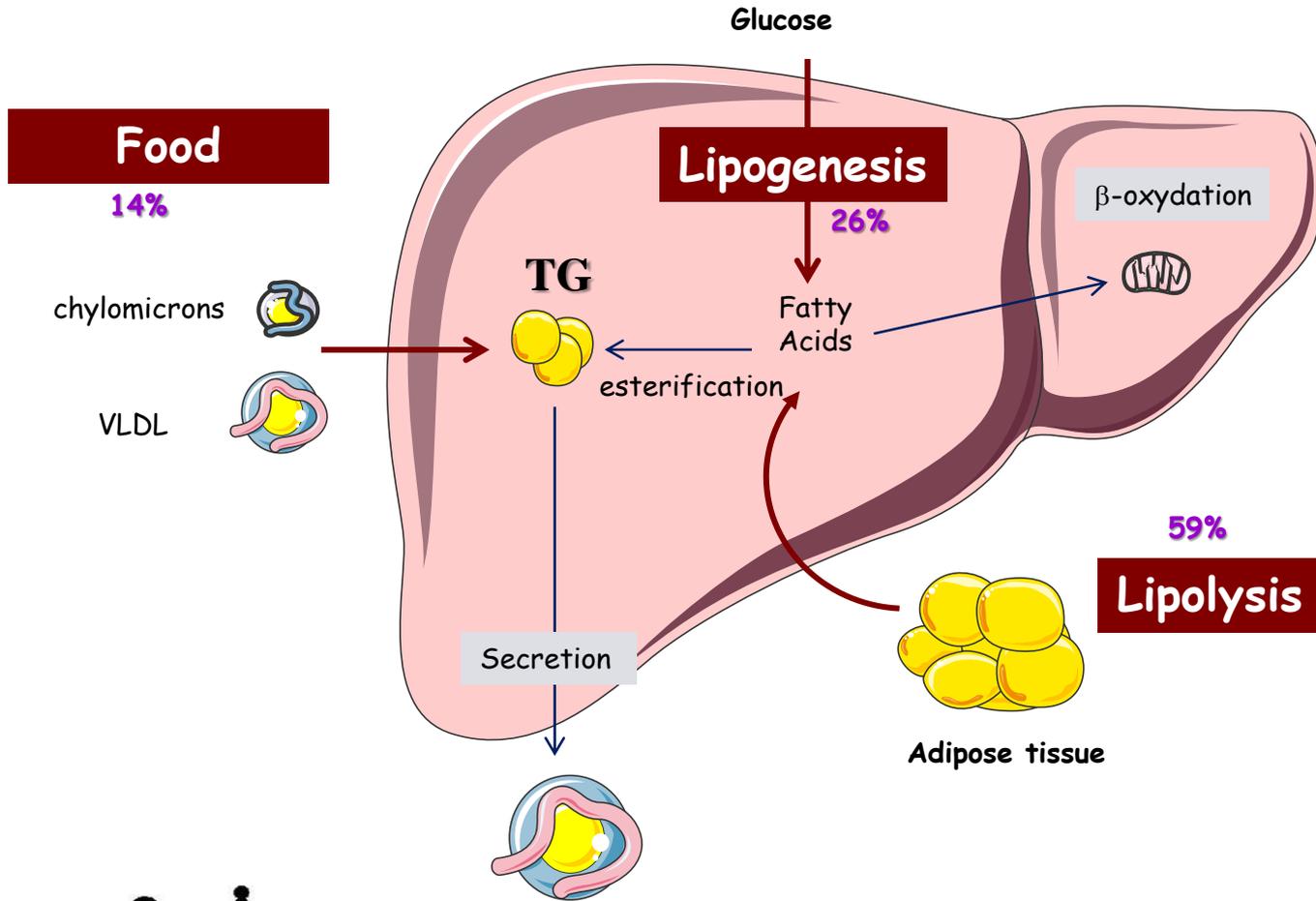
STEATOSIS

Lipolysis
Lipogenesis de novo
Food

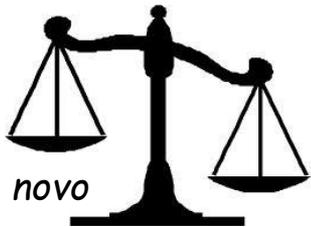
β -oxydation
Secretion



The causes of hepatic steatosis



Supply



STEATOSIS

Use

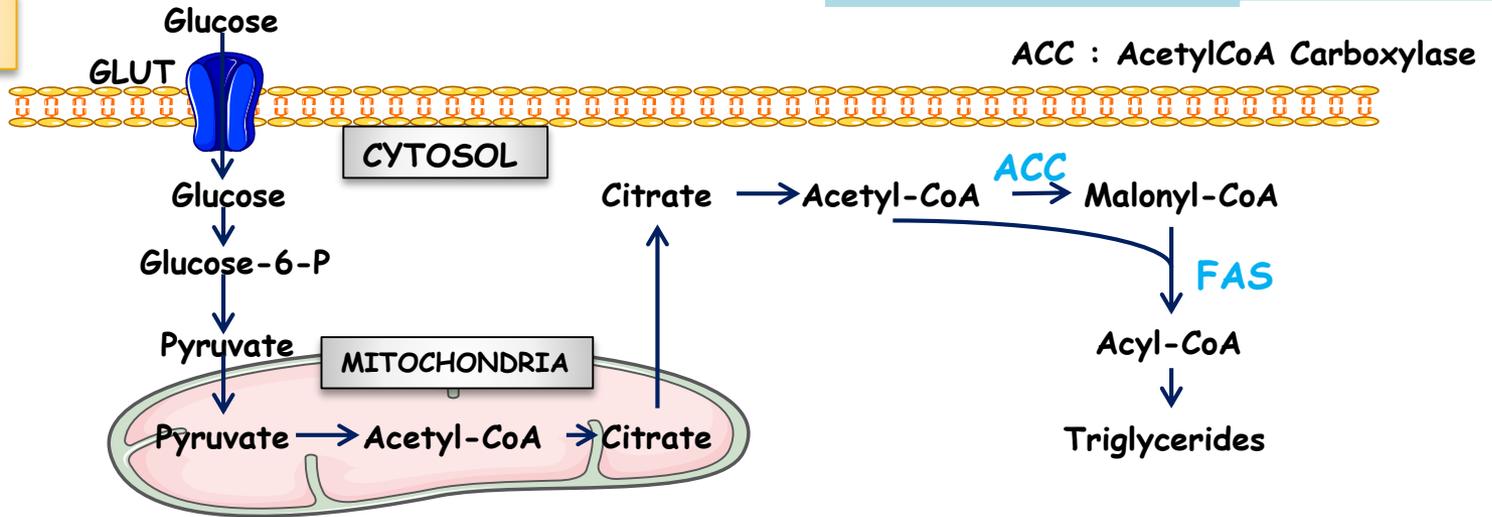
β-oxidation
Secretion

Donnelly et al., 2005

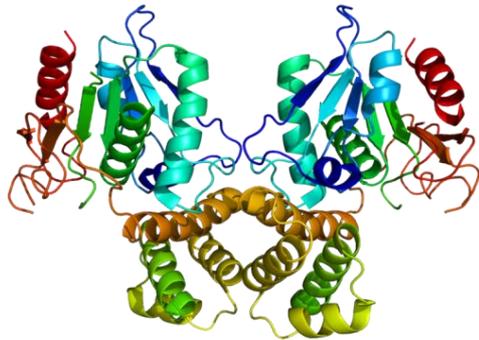
Hepatic steatosis : Fatty acids of triglycerides come from
59% of the lipolysis
26% of the lipogenesis
14 % of the food

Lipogenesis and Fatty Acid Synthase

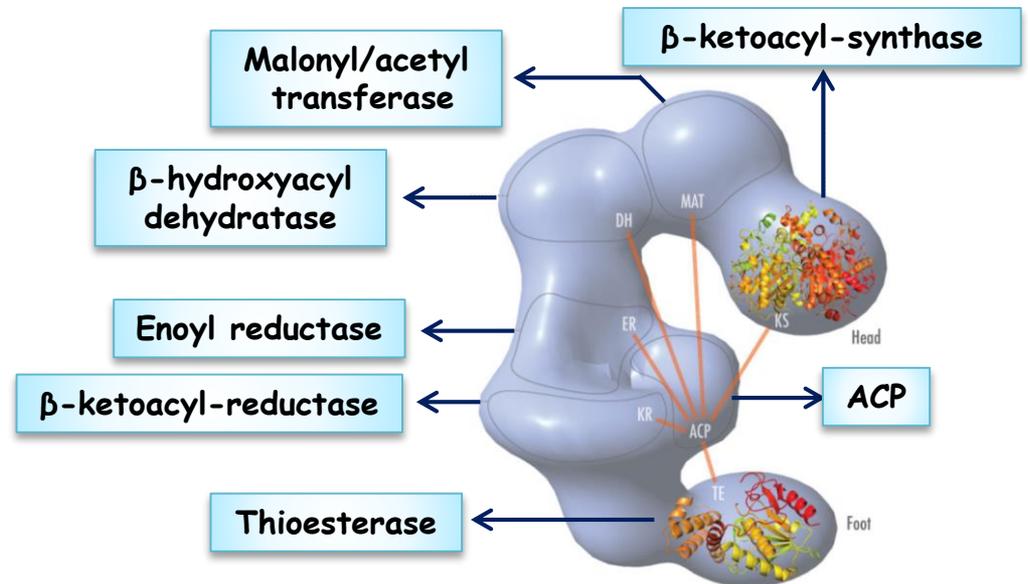
Lipogenesis



Fatty Acid Synthase



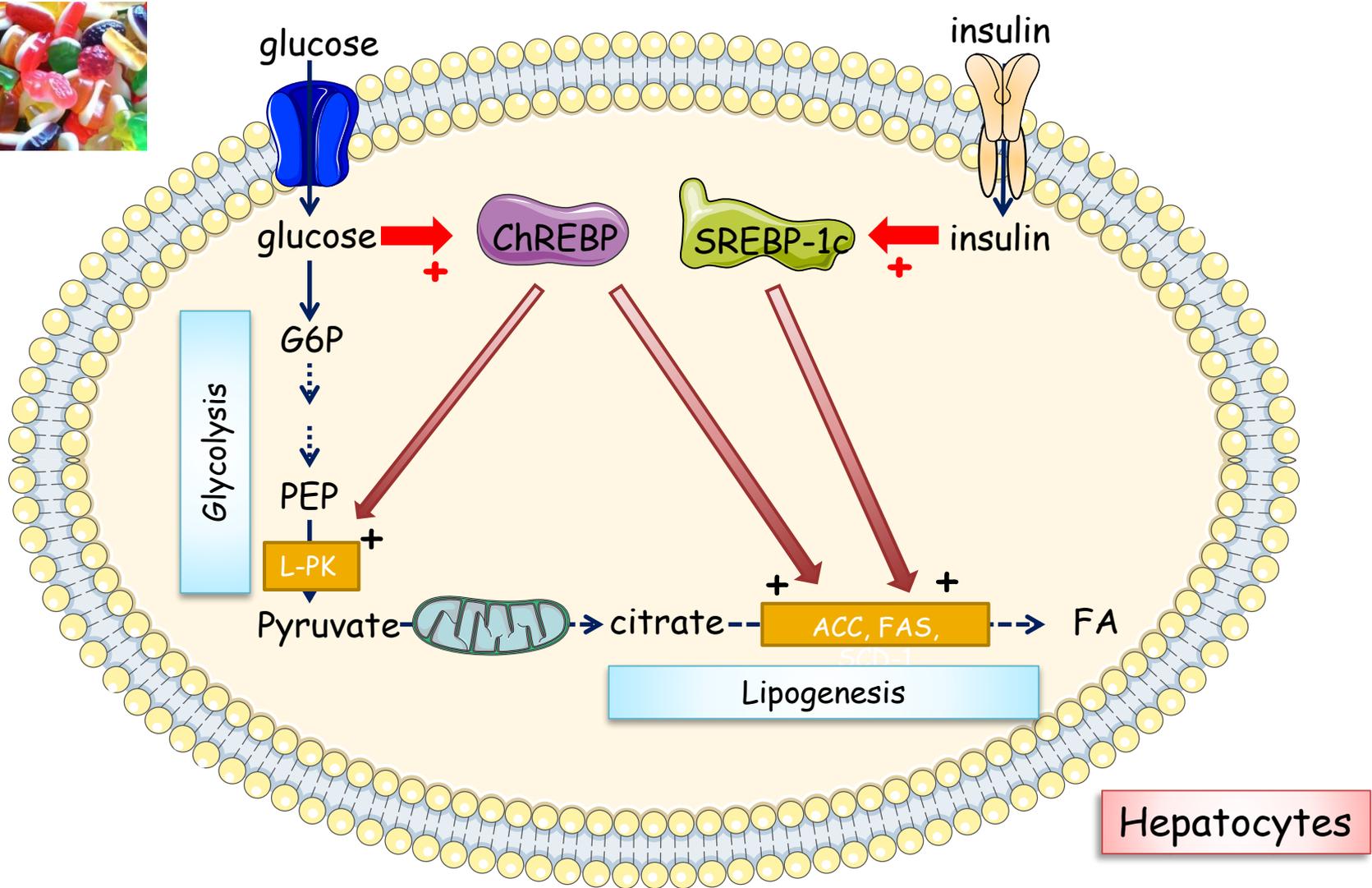
- 2 subunits
- 7 activities
- liver, adipose tissue, mammary gland



Overall reaction : $\text{acetylCoA} + 7 \text{ malonylCoA} + 14 \text{ NADPH, H}^+ \rightarrow \text{palmitoylCoA (C16:0)} + 7\text{CO}_2 + 14 \text{ NADP}^+ + 7\text{CoA}$

Regulation of FAS

➤ At the transcriptional level by the transcription factors, ChREBP and SREBP-1c



➤ At the post translational level by phosphorylation

Functions of O-GlcNAcylation

... Hundreds proteins bearing O-GlcNAc have been identified

Enzymes of metabolism

Proteins of stress

Proteins of cytoskeleton

Transcription Factors

OGT/OGA

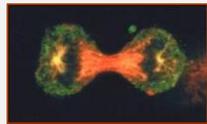
Kinases/phosphatases

Proteasome

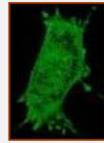
Proteins of pore nuclear

Ribosomals proteins

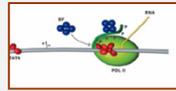
... Involved in many cellular processes



Cell cycle



Apoptosis



Translation



Transcription



Development



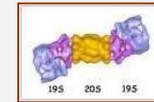
Signaling



Cell trafficking



Cellular architecture



Proteins degradation

... dynamism disturbed in certain diseases



Type 2 diabetes



Neurological diseases



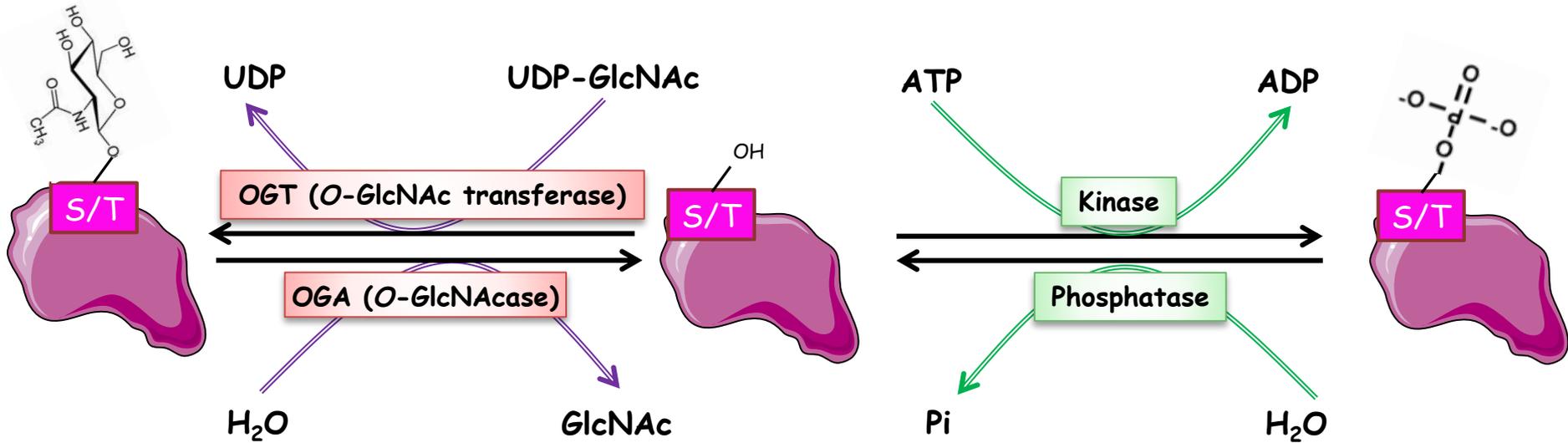
Cardiovascular diseases



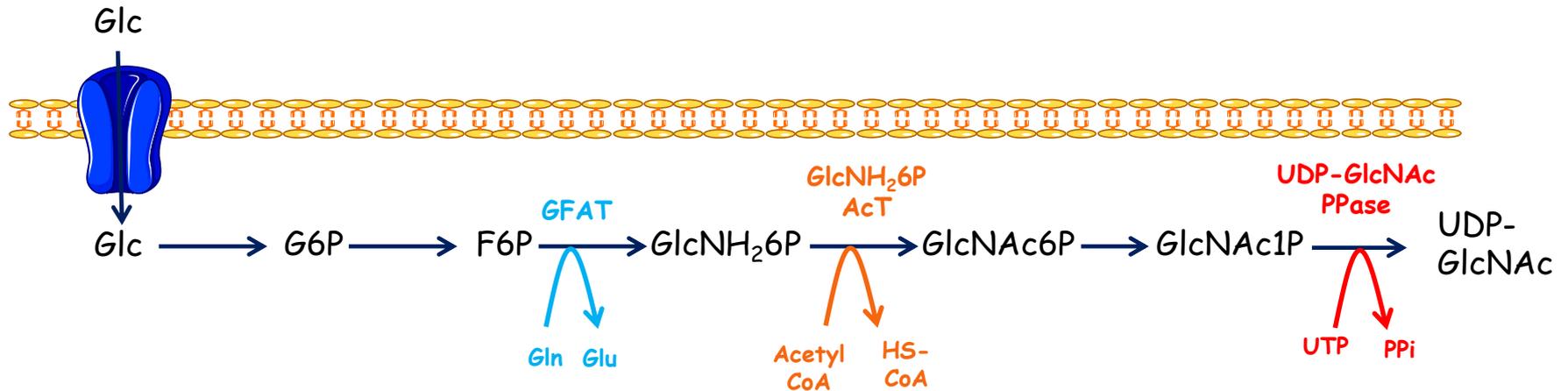
Cancers

O-N-acetylglucosinylation (O-GlcNAcylation)

O-GlcNAcylation

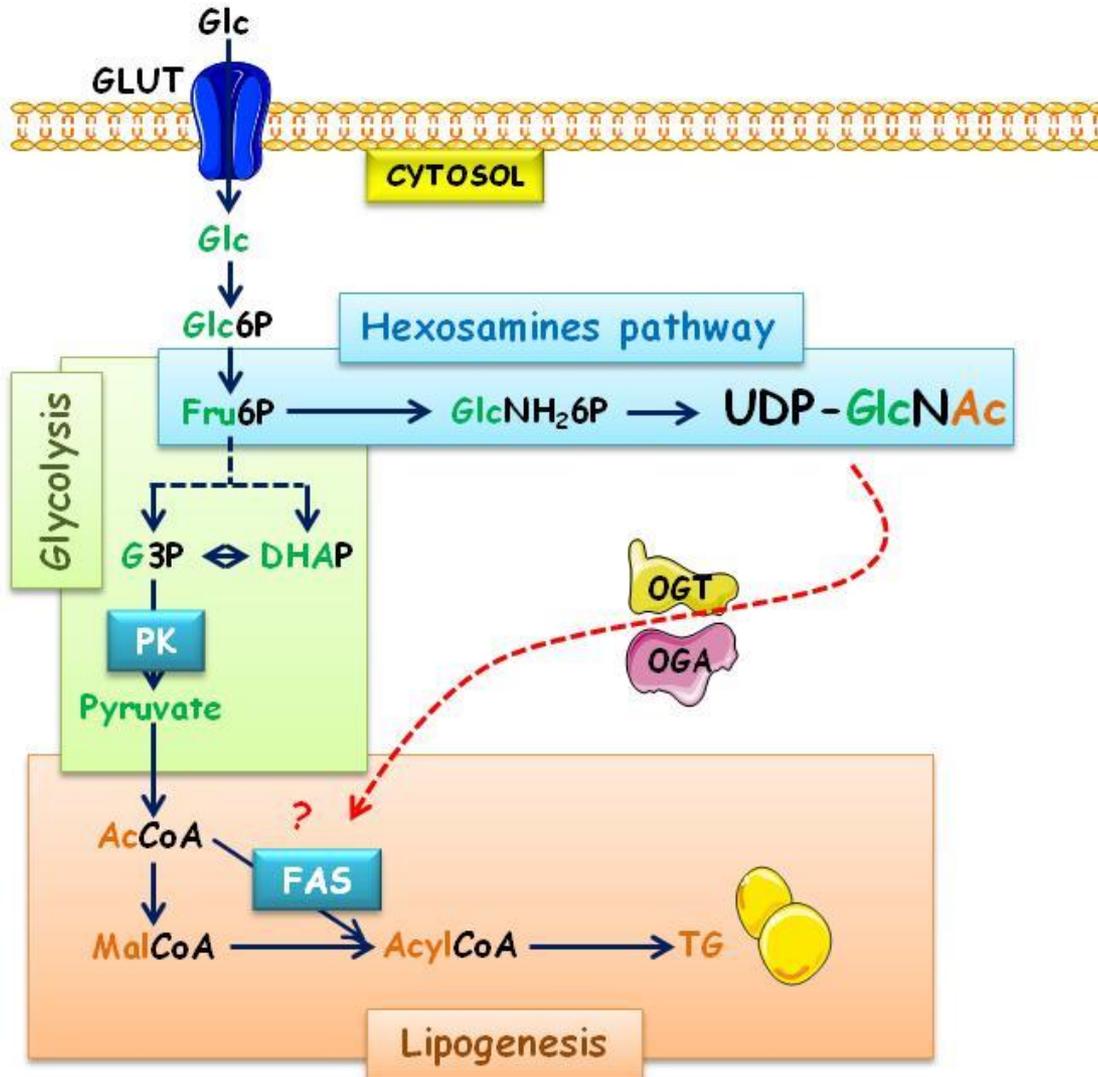


The Hexosamine Biosynthesis Pathway (HBP)



Hypothesis

- FAS expression and *O*-GlcNAcylation level depend on glucose concentrations



Relation between *O*-GlcNAcylation, expression and activity of FAS

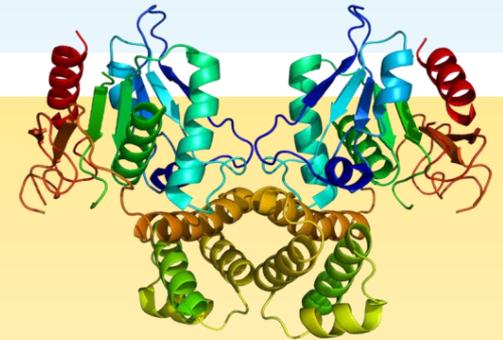


➤ Is FAS *O*-GlcNAcylated ?



□ Relations between FAS *O*-GlcNAcylation and nutritional conditions

➤ Do *O*-GlcNAcylation levels interfere with:

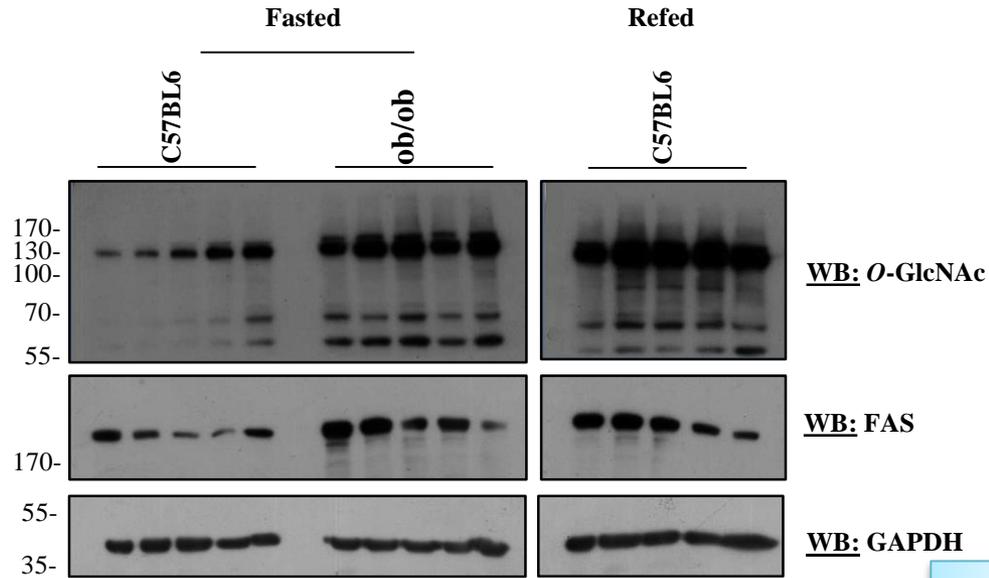


□ FAS expression

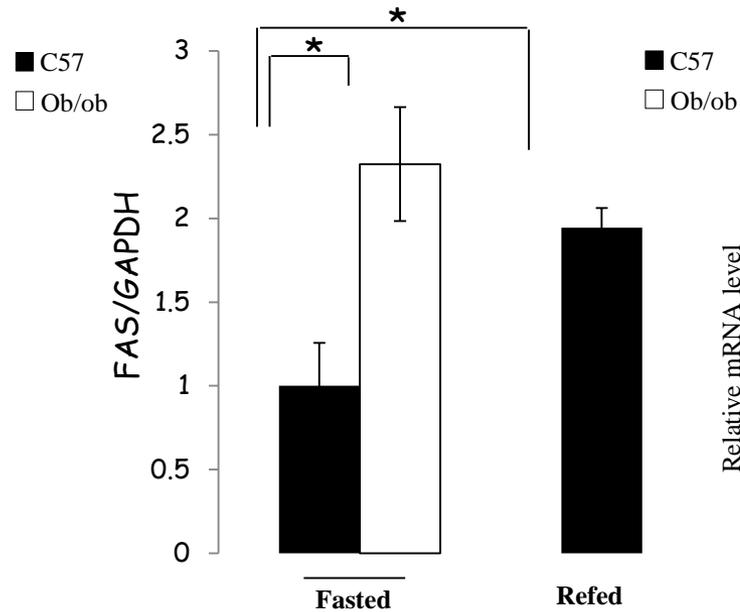
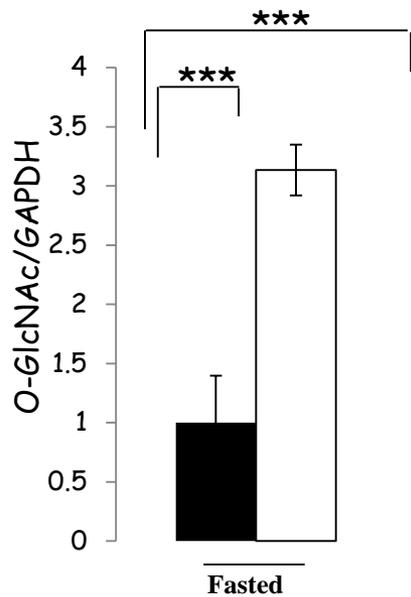
□ FAS stabilization

□ FAS activity

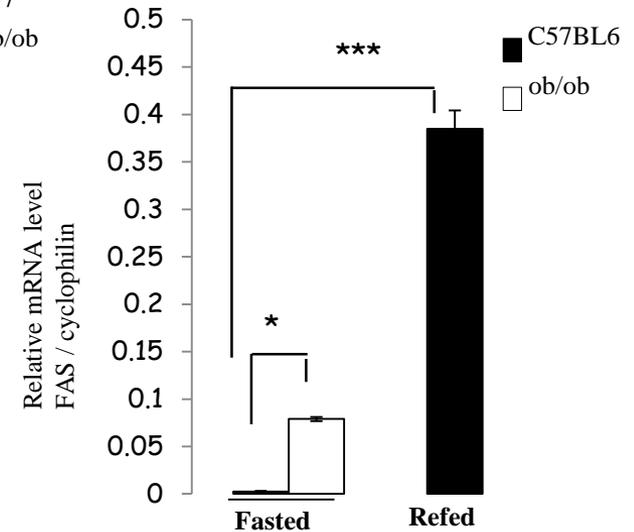
O-GlcNAcylated protein levels and FAS expression



Quantification



FAS ARNm



O-GlcNAcylation levels and FAS expression in physiopathological models

Model 2 : Use of mice C57BL6 fed a chow diet or fed a High Carbohydrate Diet.



12 weeks



Chow Diet (CD)



65% carbohydrate, 24%
protein, 11% fat

Fasted



Group1 : CD Fasted

Fasted



HCHO



Group2 : CD Refed

Fasted



Group3 : HCD Fasted

High Carbohydrate Diet (HCD)



75% carbohydrate, 22%
protein, 3% fat

Fasted

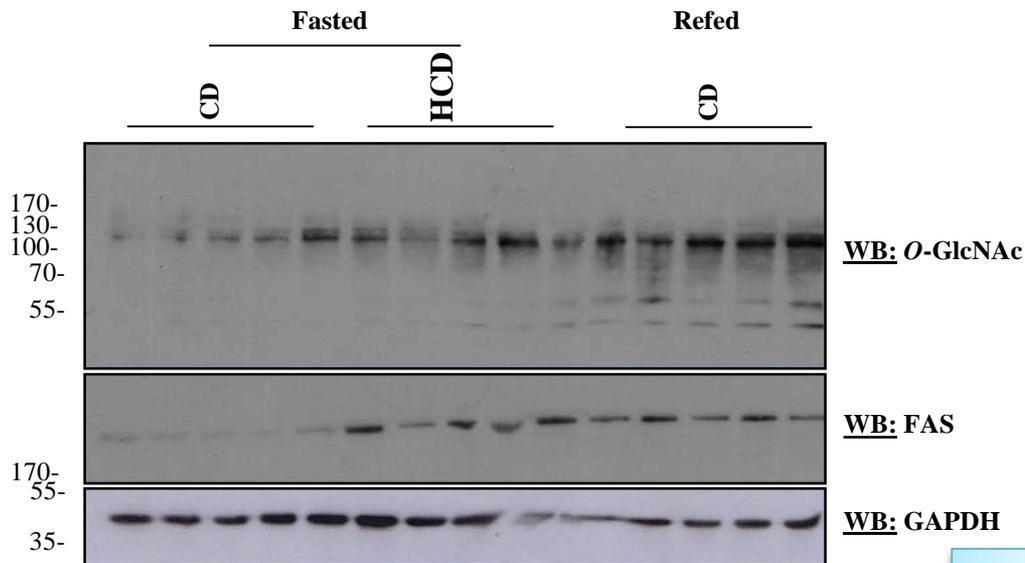


HCHO

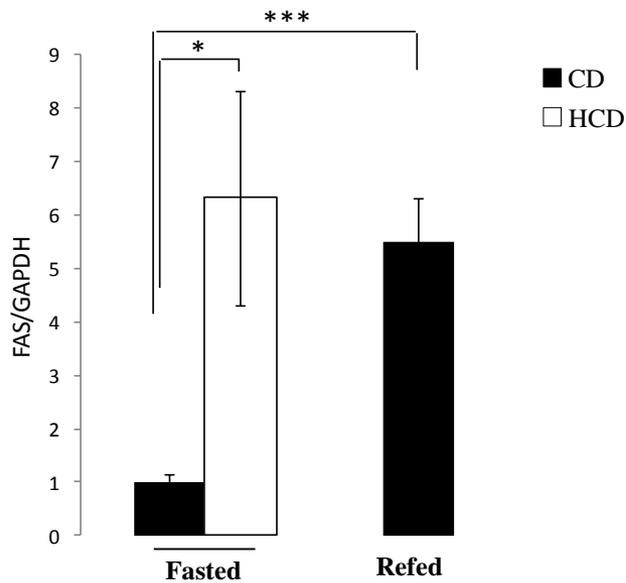
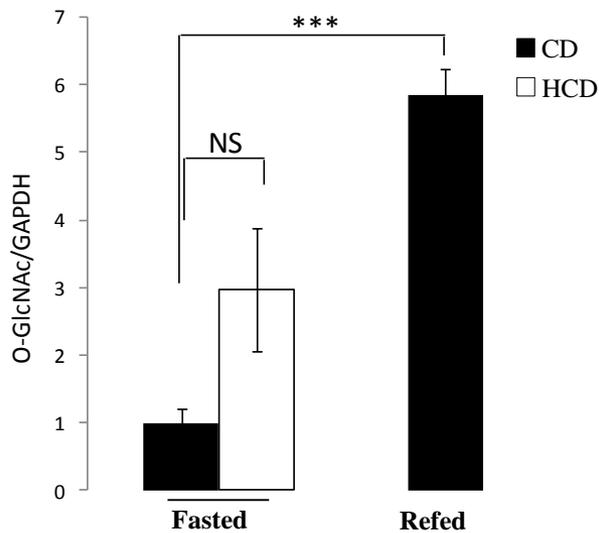


Group4 : HCD Refed

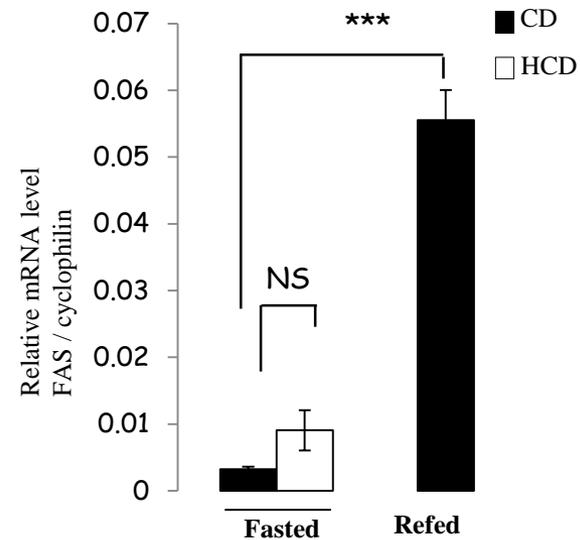
O-GlcNAcylated protein levels and FAS expression



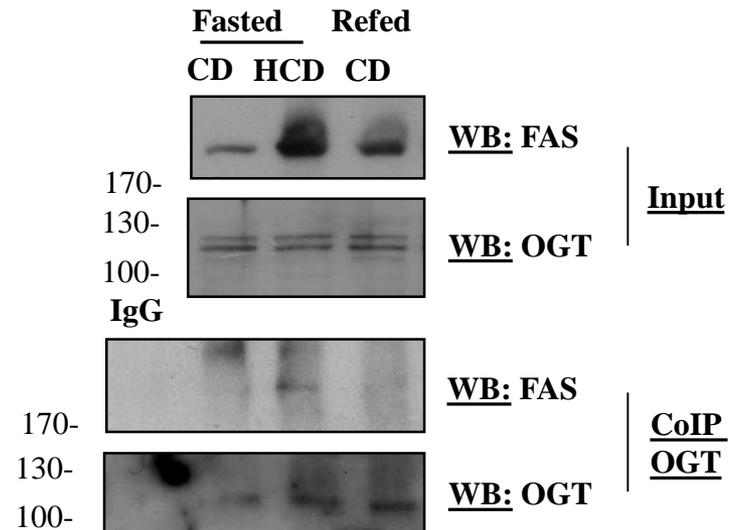
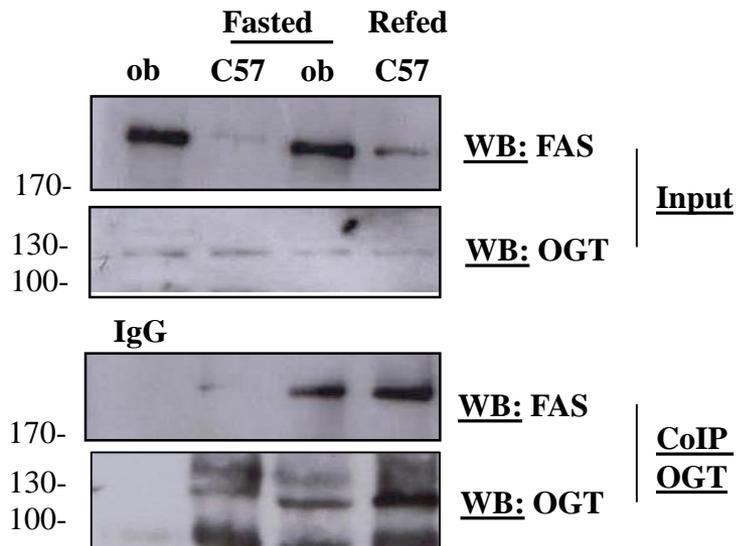
Quantification



FAS ARNm



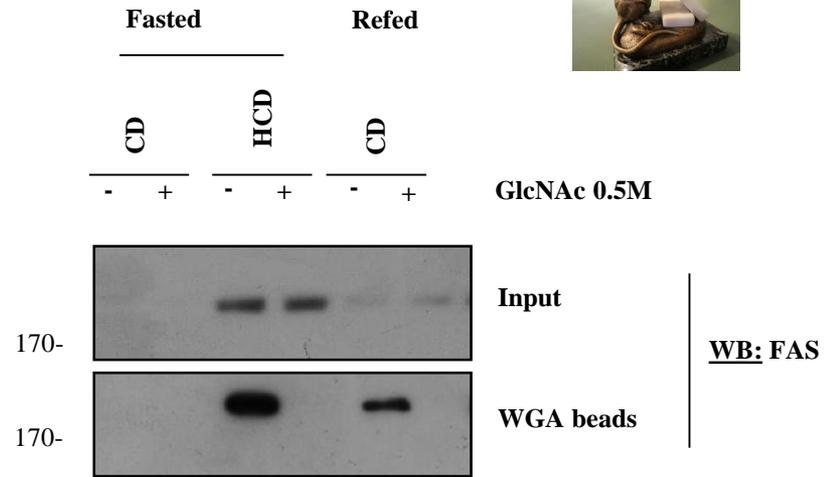
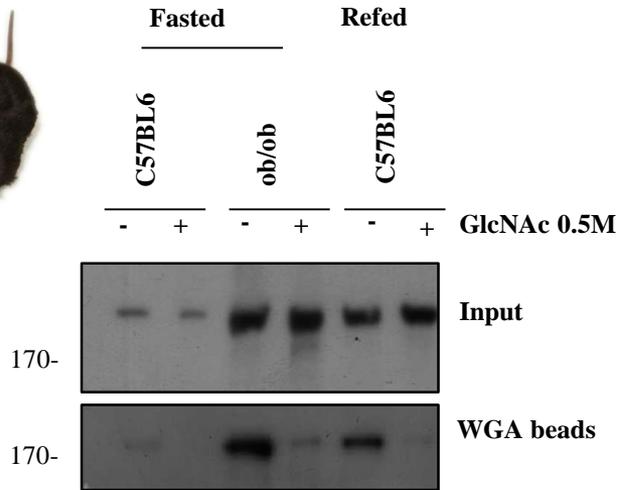
Interaction FAS/OGT



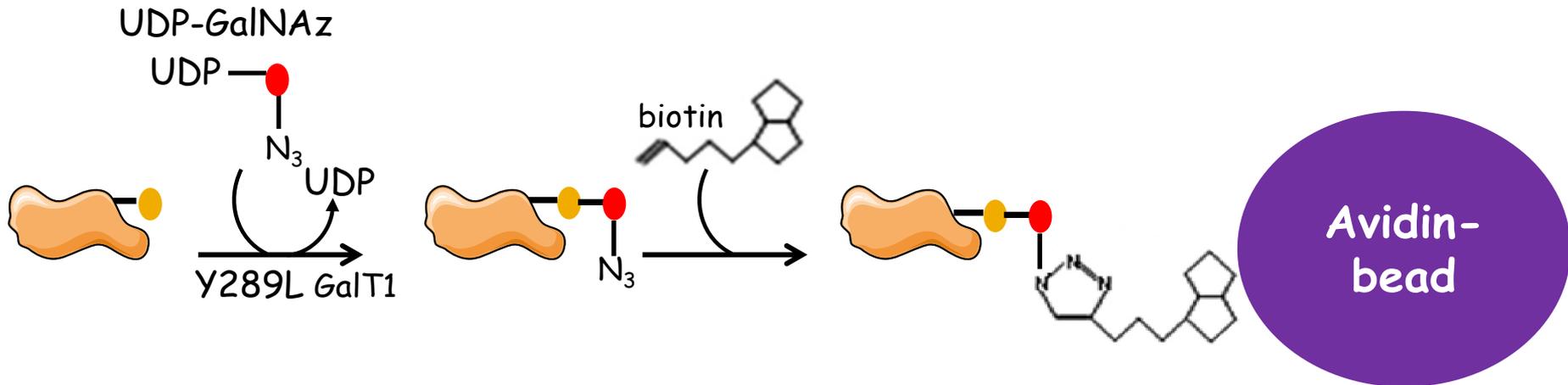
➔ FAS and OGT are partners of interaction

FAS O-GlcNAcylation

Use of lectins : Wheat Germ Agglutinin

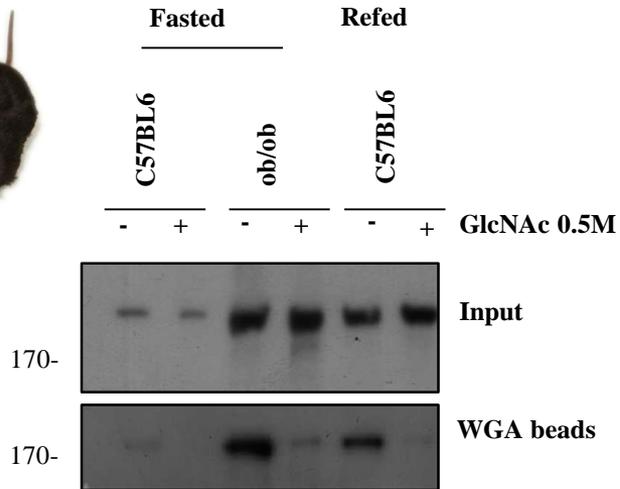


click chemistry

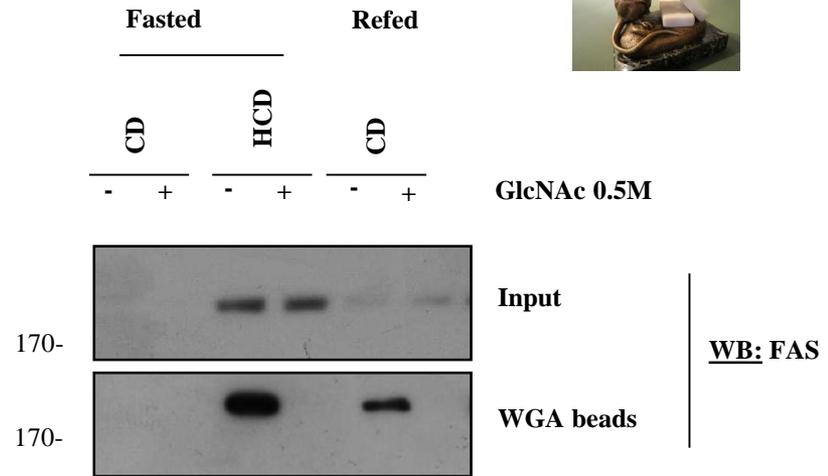


FAS O-GlcNAcylation

Use of lectins : Wheat Germ Agglutinin

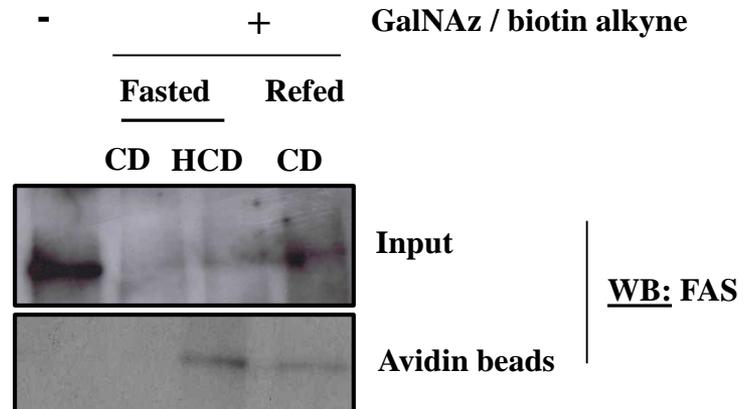
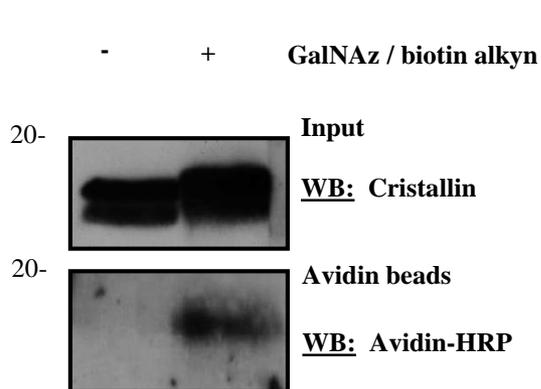


WB: FAS



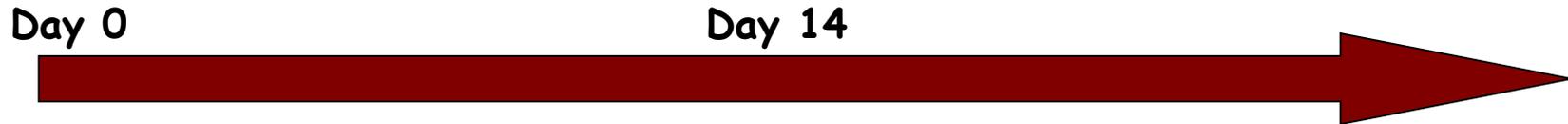
WB: FAS

click chemistry

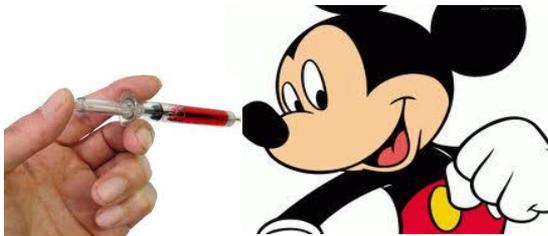
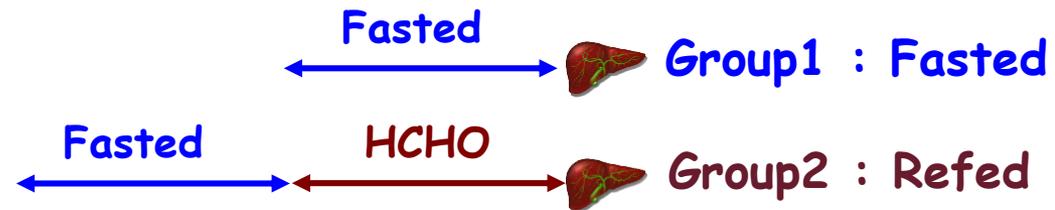


O-GlcNAcylation levels and FAS expression in physiopathological models

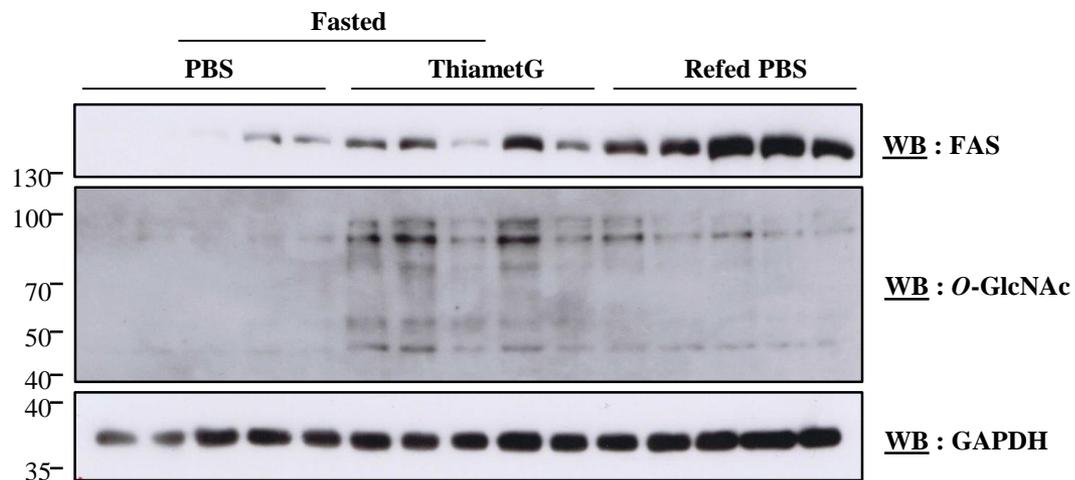
Model 3 : Use of mice C57BL6 presenting an inhibition of OGA



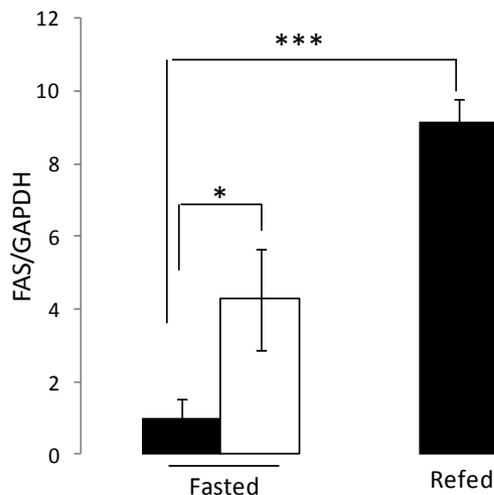
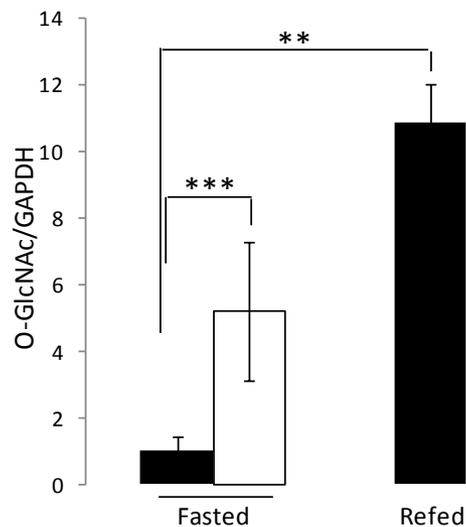
Daily injection of PBS/ThiametG
20 mg/ kg/ day



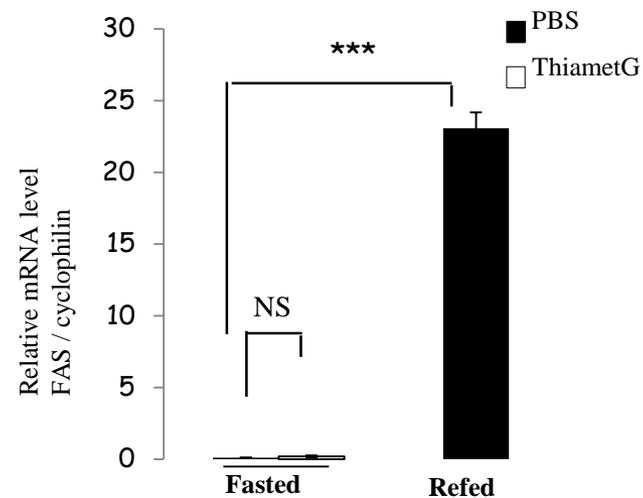
O-GlcNAcylated protein levels and FAS expression



Quantification



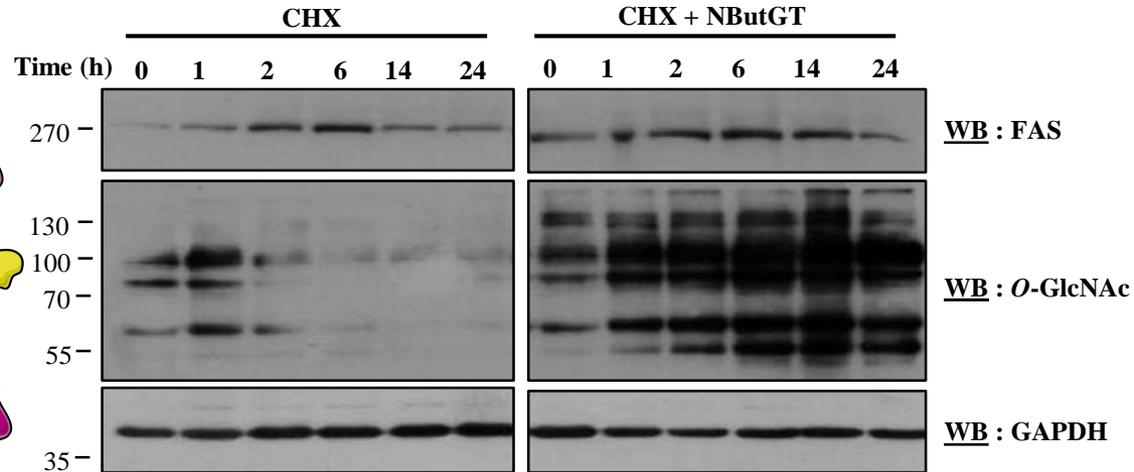
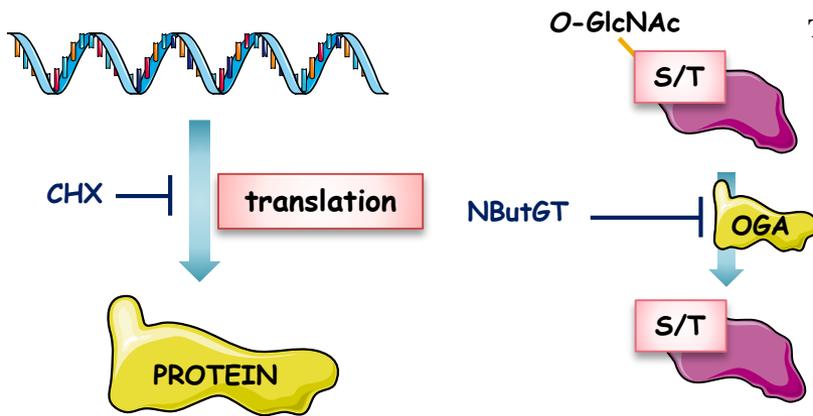
FAS ARNm



Role of *O*-GlcNAcylation on FAS stabilisation

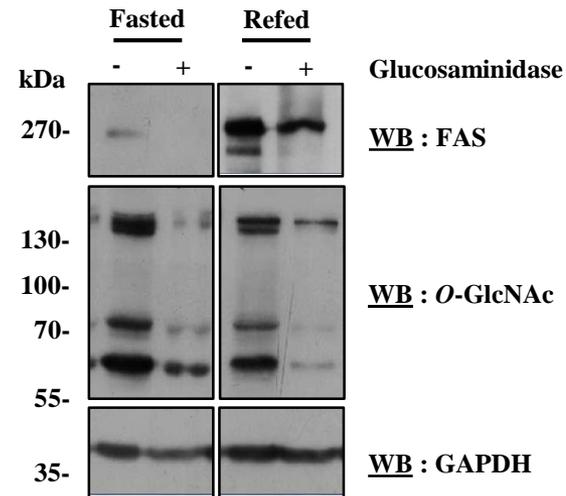
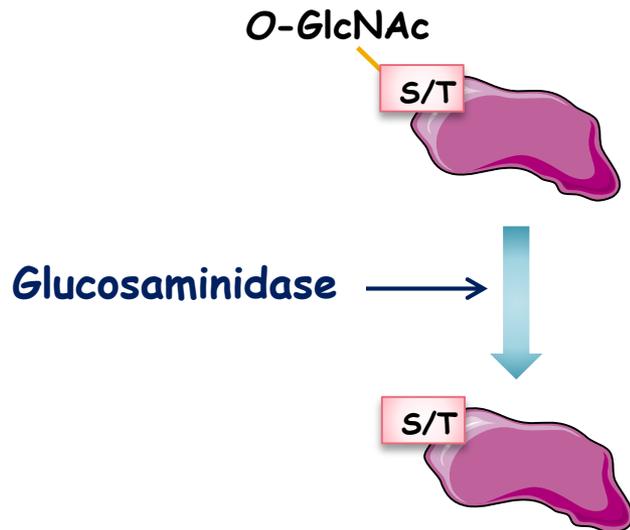
HepG2

Kinetic with cycloheximide



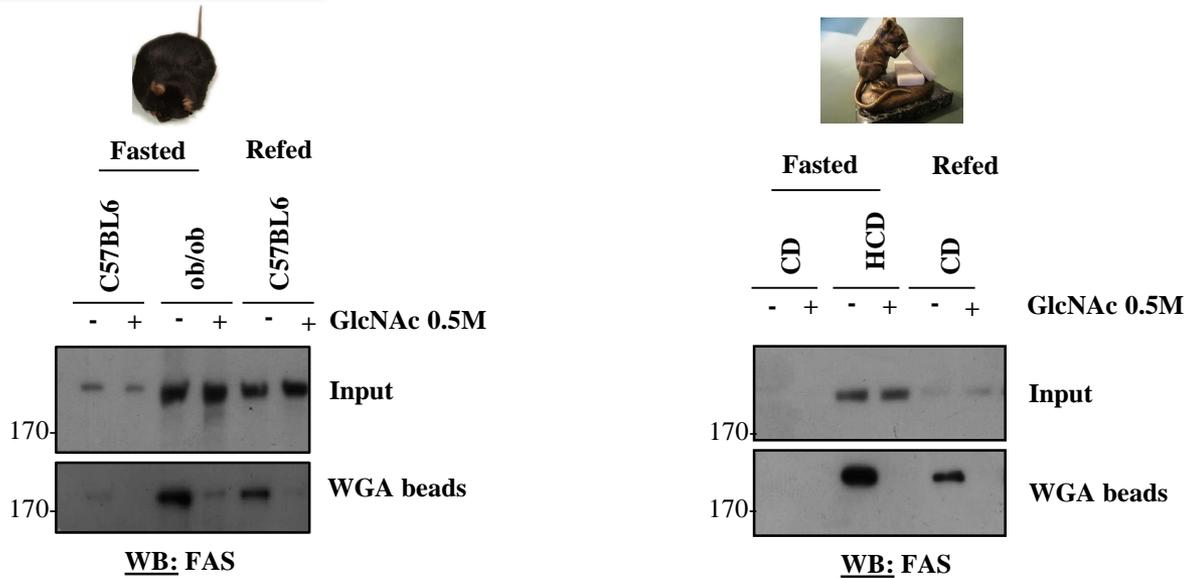
Liver mice

Treatment with β -N-acetylglucosaminidase

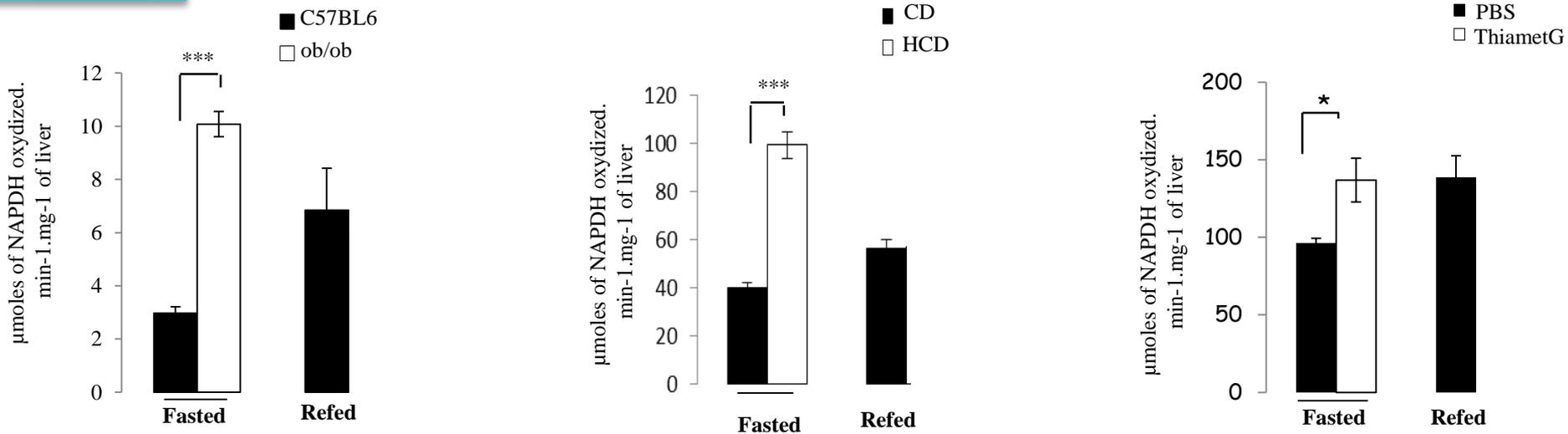


Role of O-GlcNAcylation on FAS activity

FAS O-GlcNAcylation

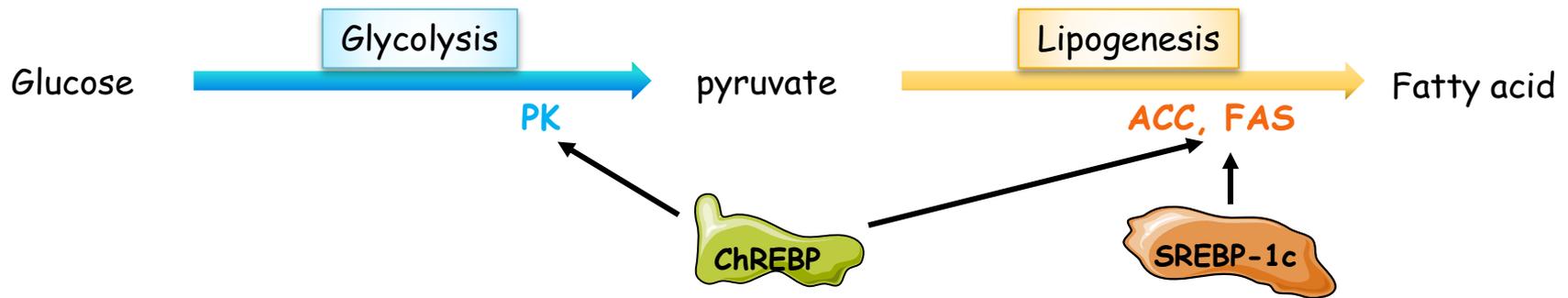


FAS activity



FAS regulation :

❑ by transcription factors



❑ by O-GlcNAcylation

➤ FAS interacts with OGT and it's O-GlcNAcyated

➤ Roles of the O-GlcNAcylation :

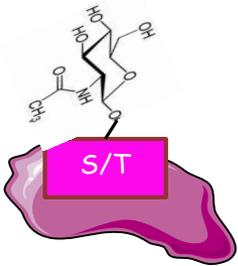
❑ Increase of global O-GlcNAcylation levels is correlated with an increase of FAS expression through a reduction of its degradation.

❑ FAS activity is in correlation with its O-GlcNAcylation



Relation *O*-GlcNAcylation/ubiquitylation of FAS

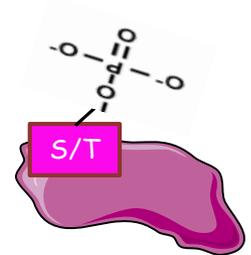
Hypothesis



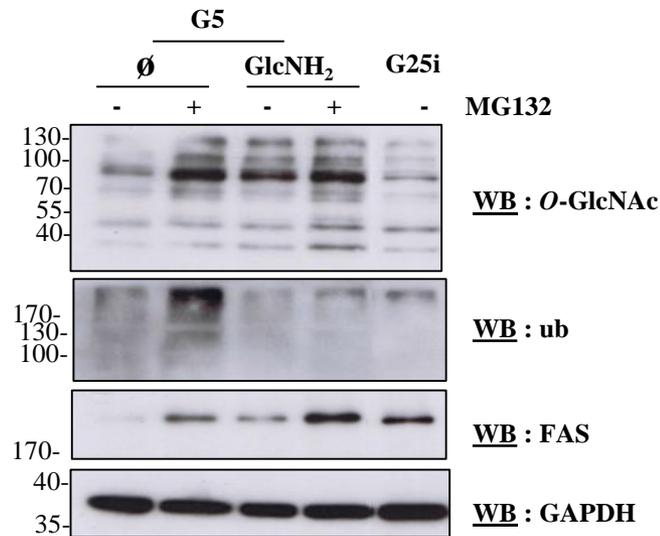
O-GlcNAcylation
Stabilisation



Phosphorylation
Ubiquitylation
Degradation



Preliminary results



Methodology

- Perform siRNA to silent OGT
- Evaluate FAS ubiquitylation in function of *O*-GlcNAcylation levels

➔ The MG132 can restore FAS expression at G5 similar to the condition G25. **FAS** seems to be degraded by the proteasome in the liver

Team of Pr Tony Lefebvre

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Dr Ikram El Yazidi
Dr Anne Sophie Vercoutter Edouart
Dr Agata Steenackers
Dr Nao Yamakawa
Moyira Aquino-gil
Maité Leturcq
Anne-Marie Mir
Marlène Mortuaire
Jeanne Vermuse

Collaborations



Céline Guinez (Unité Environnement Périnatal et santé, V d'Ascq)
Catherine Postic (Institut de Cochin, Paris)
Isabelle Hainault (Centre des Cordeliers, Paris)
David Vocadlo (Simon Fraiser University, Burnaby)