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About OMICS International Conferences

OMICS International is a pioneer and leading science event organizer, which publishes around 500 open access journals and conducts over 500 Medical, Clinical, Engineering, Life Sciences, Pharma scientific conferences all over the globe annually with the support of more than 1000 scientific associations and 30,000 editorial board members and 3.5 million followers to its credit.

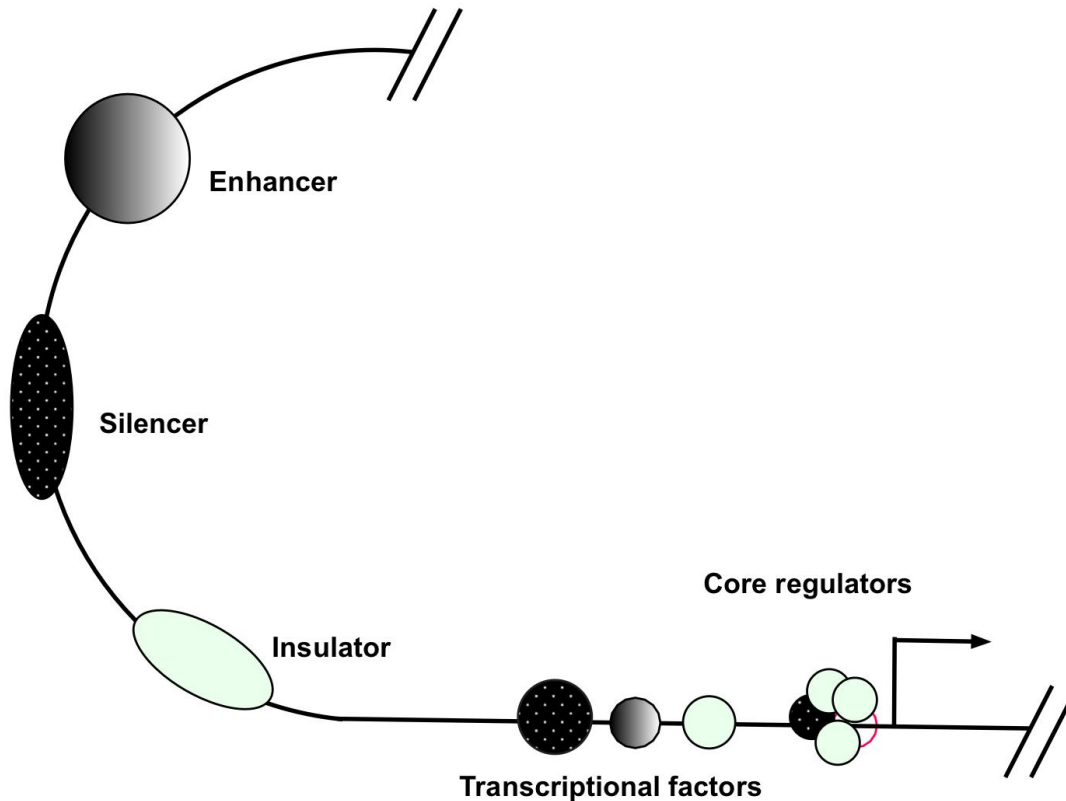
OMICS Group has organized 500 conferences, workshops and national symposiums across the major cities including San Francisco, Las Vegas, San Antonio, Omaha, Orlando, Raleigh, Santa Clara, Chicago, Philadelphia, Baltimore, United Kingdom, Valencia, Dubai, Beijing, Hyderabad, Bengaluru and Mumbai.

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Genome-scale Promoter analysis using Exome Sequences (GPES)

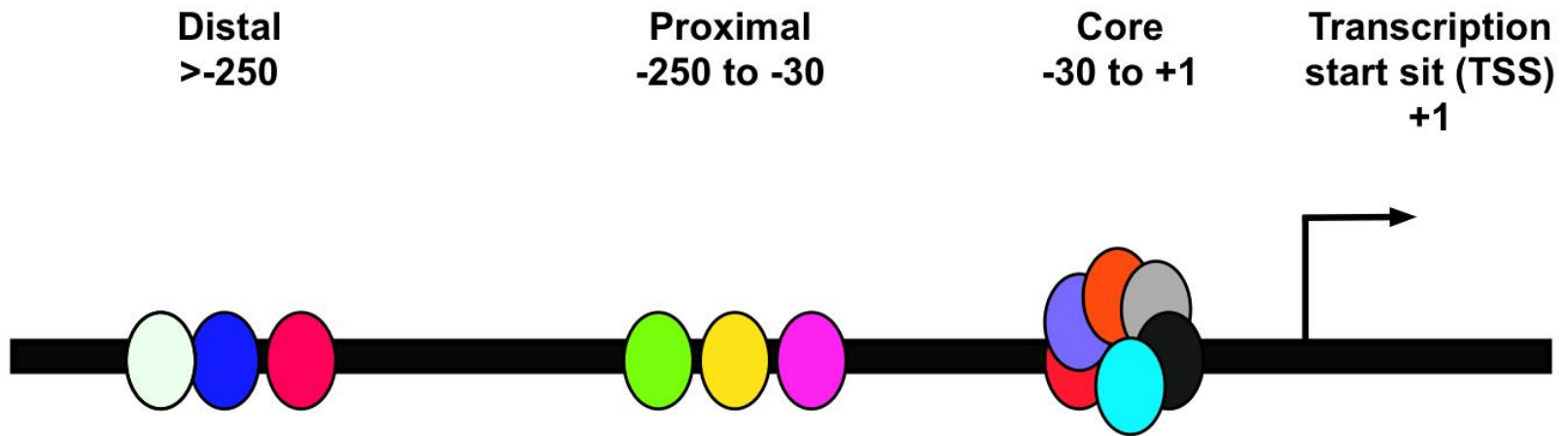
San Ming Wang

Regulation of gene expression



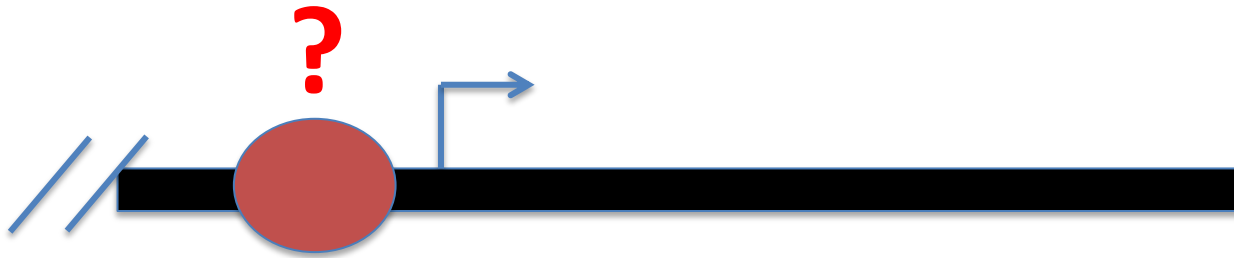
- **Distal elements**
 - enhancer, silencer, insulator
- **Promoter elements**
 - transcriptional factors, basal transcriptional machinery

Promoter structure



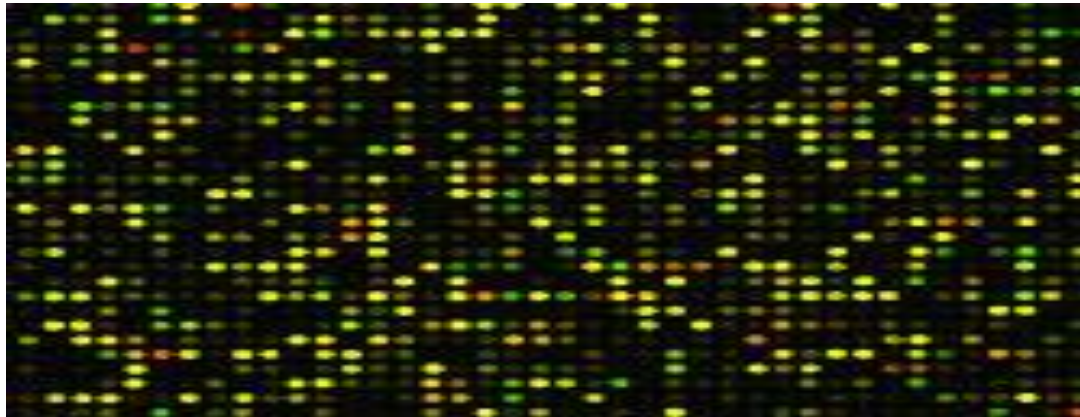
Promoter remains as a less-explored territory in disease study

- Promoter directly controls gene expression
- Mutation in promoter can have profound impact on biology and diseases
- Lack of systematic tools for *de novo* promoter analysis



Existing method - Microarray

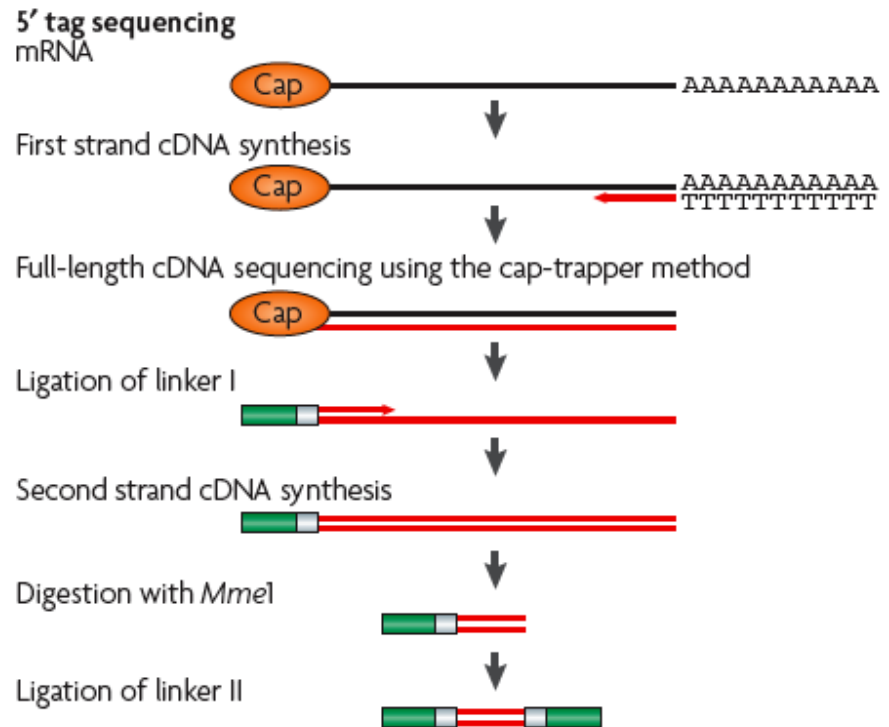
- Promoter probes are based on human genome reference sequences
- Human genome reference sequences were from “normal” individual genomes, not from disease genomes
- Array is lack of the power for *de novo* promoter analysis



Existing method

- Cap Analysis Gene Expression (CAGE)

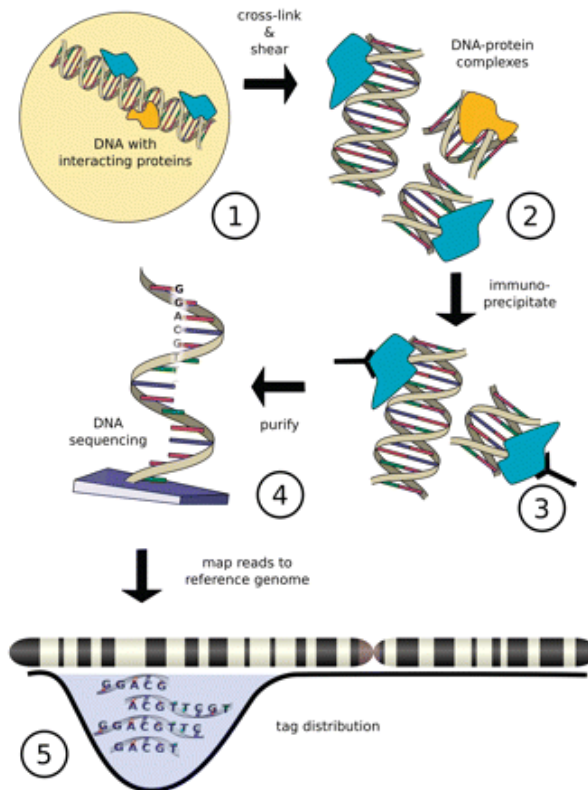
- Determine transcriptional start site
- Do not analyze promoter



(*Nature genetics* **38**, 626–35, 2006)

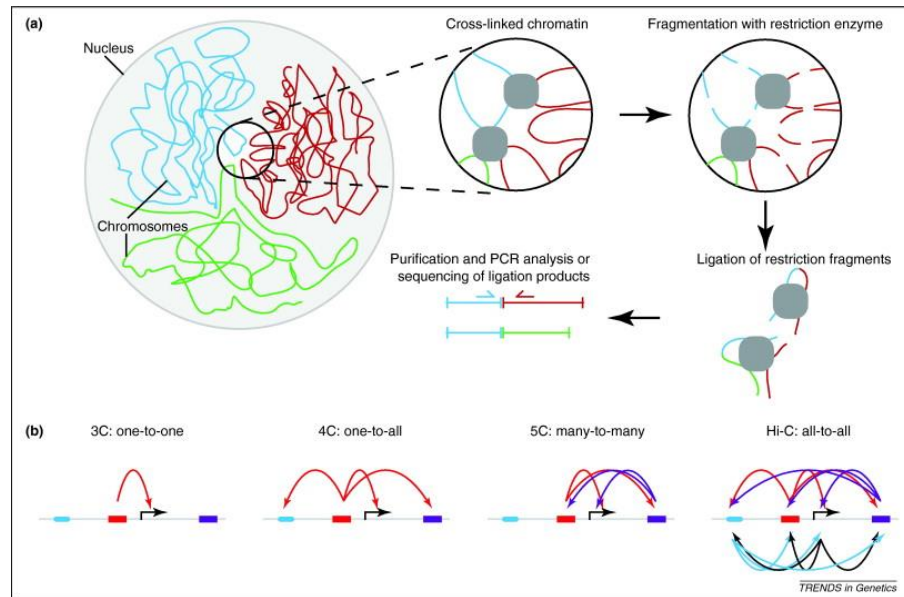
Existing method - ChIP-seq

- Only analyze a given type of transcriptional factor-binding sites by immuno-precipitation



Existing method – Hi-C

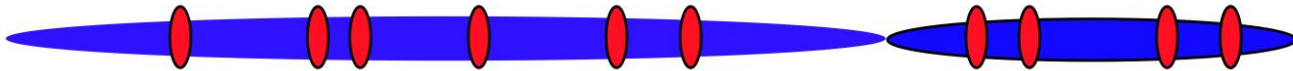
- Detect interaction between distal and promoter
- Not focus on *de novo* promoter mapping



Existing method

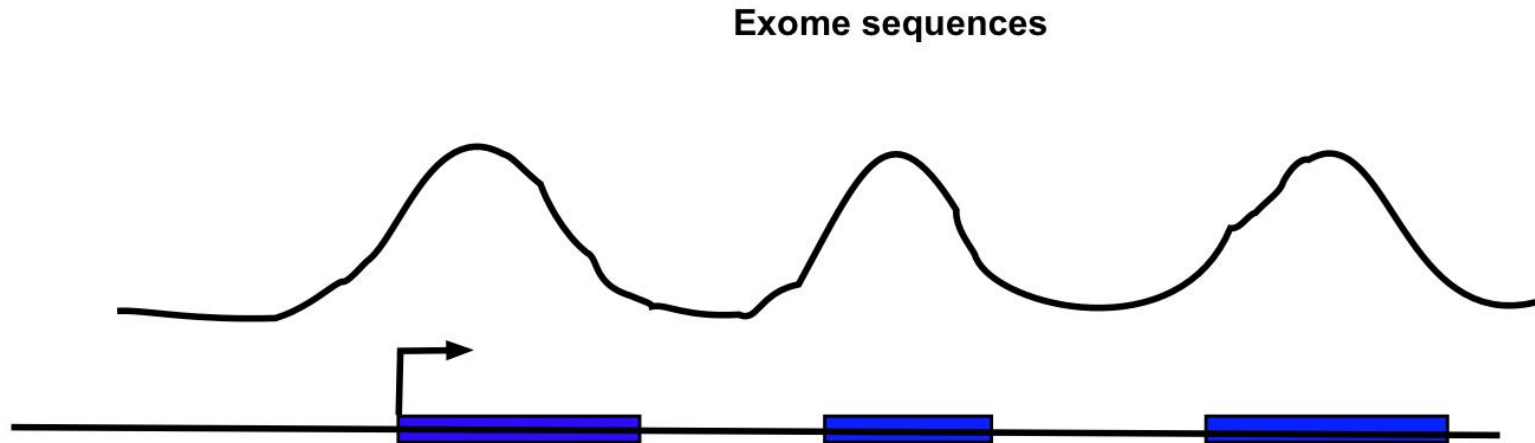
– Whole-genome sequencing

- It is not cost-effective to analyze promoters by sequencing the entire genome, as promoters account for only a small portion of the genome - **using a canon to hit ants**



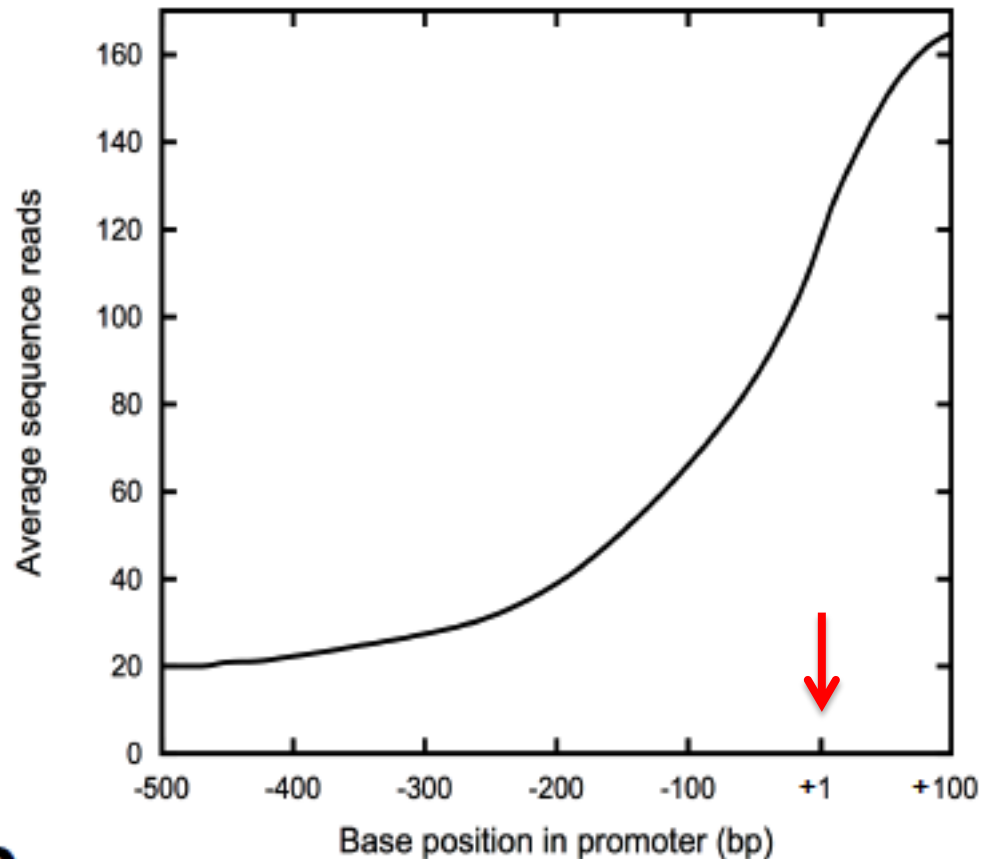
Can exome sequences be used for
genome-scale, *de novo* promoter analysis?

Distribution of actual exome sequences

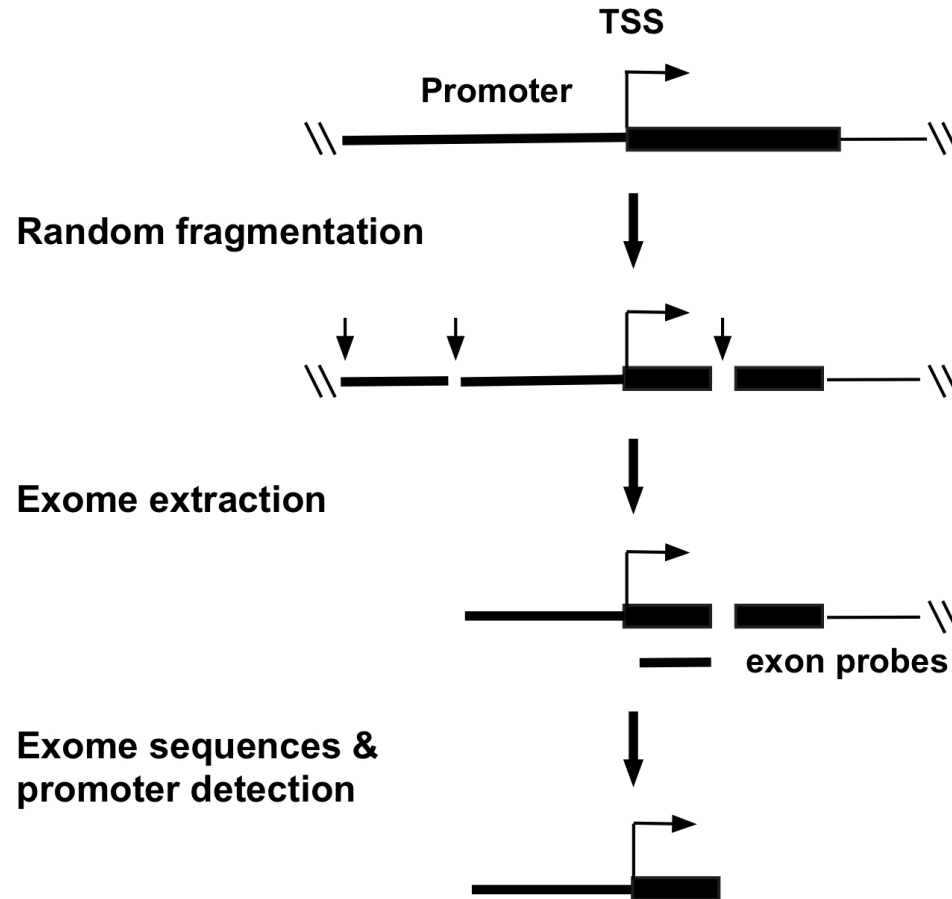


(BMC Genomics. 13:194, 2012)

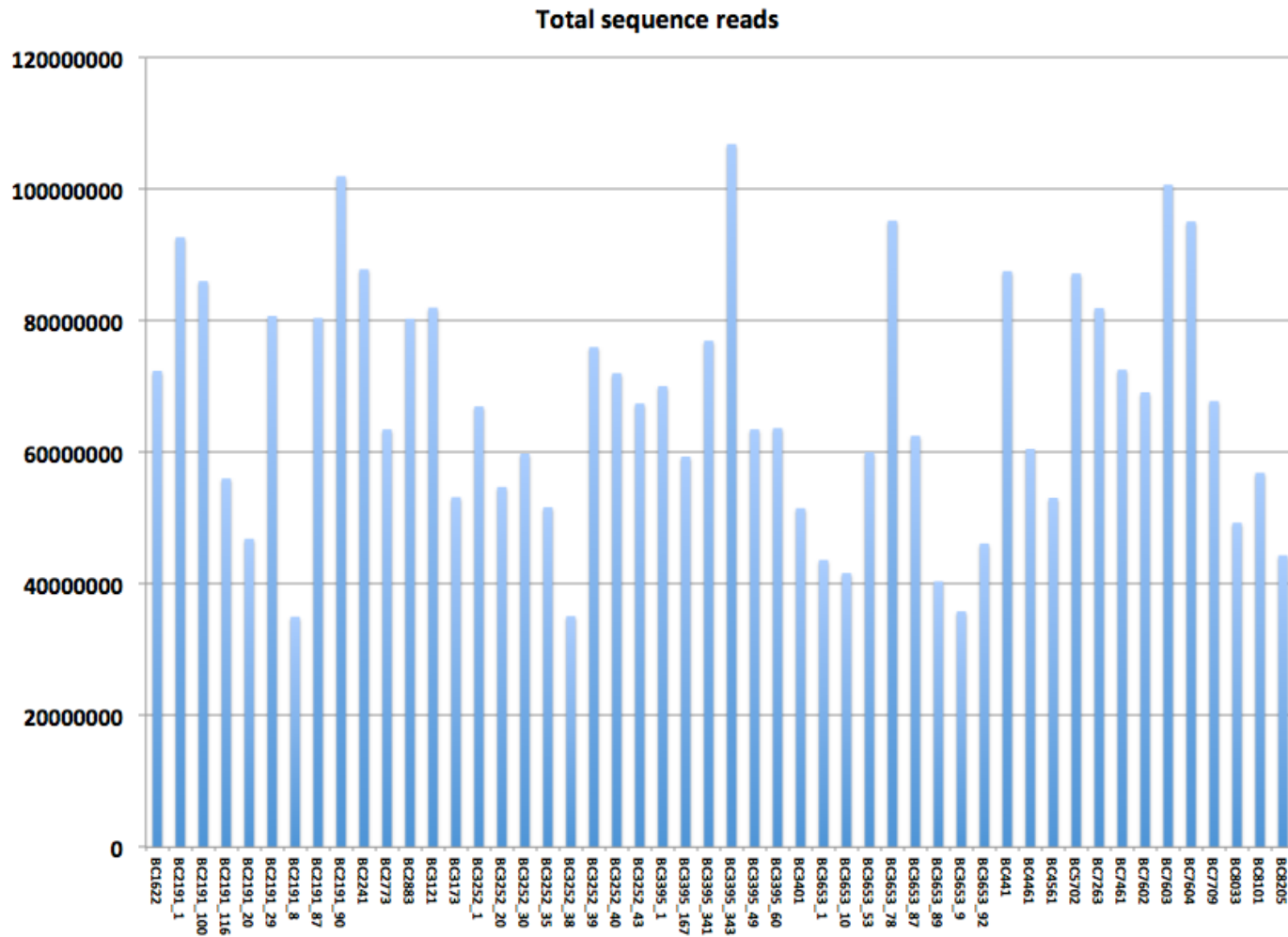
Distribution of exome data in promoter region (-500 to +100) from 50 exome data sets



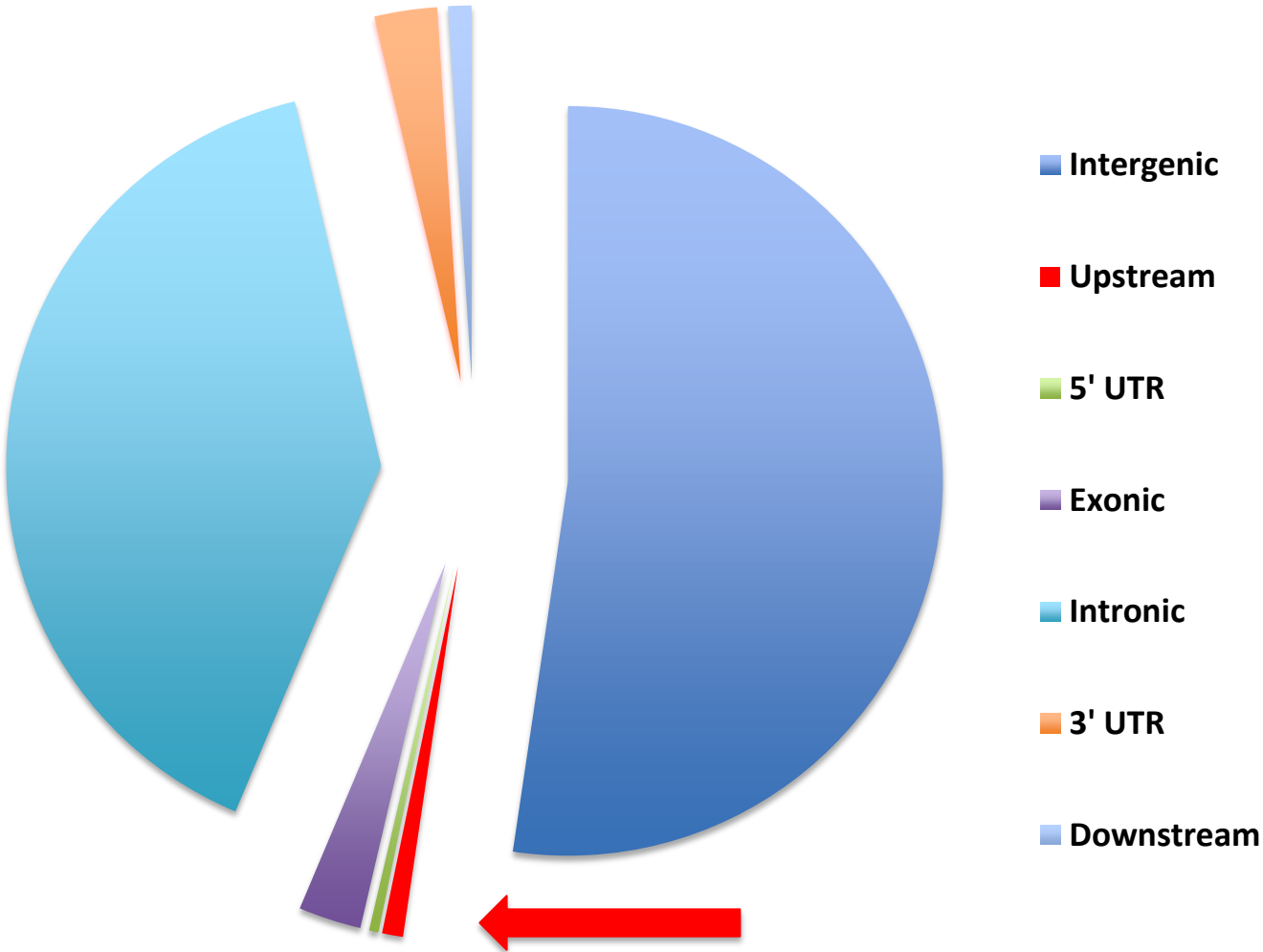
How does exome collect promoter sequences



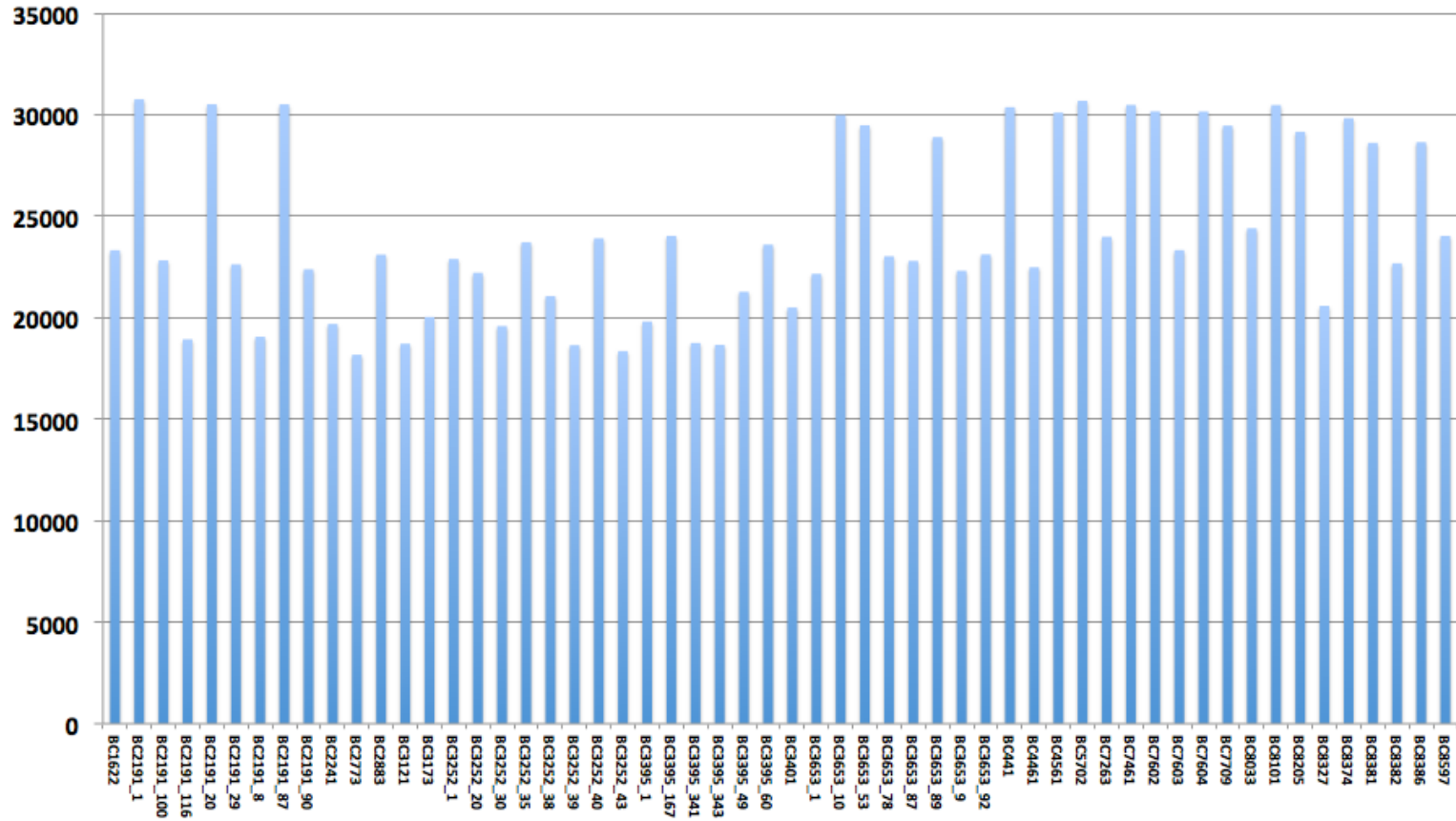
50 exome data from breast cancer



Distribution of variants from the 50 exomes



Variants detected in promoters



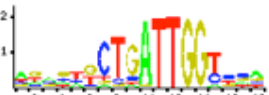
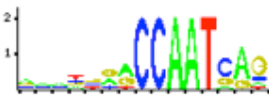



Types of promoter variants identified

Class	Promoter	SNV	Indel	Total
Known motif	601	521	57	578
TFBS motif	238	251	37	288
TATA box	179	195	30	225
Total	1,018	967	124	1,091

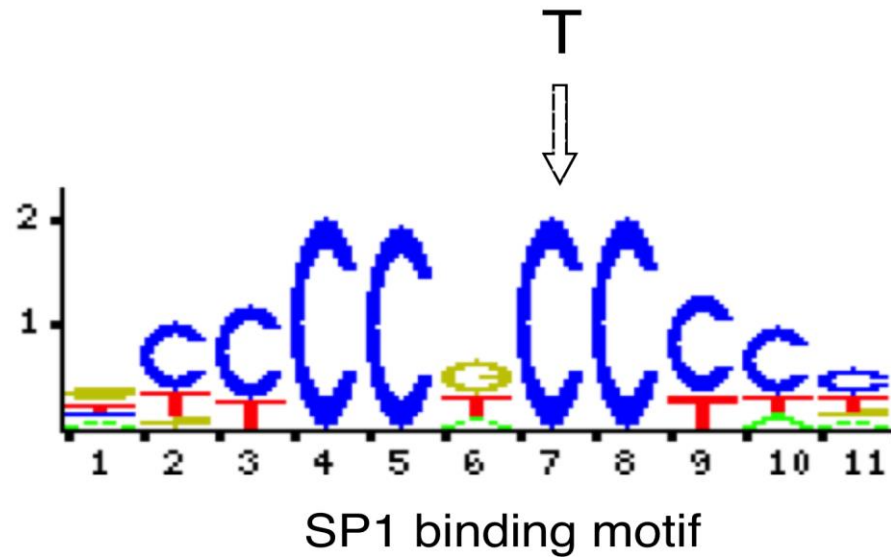
Examples - variants in TATA-boxes

Gene	Chromosome	Position	Reference	Variant	Strand	TATA-box	Changed
SUCLG1	chr2	84686610	A	G	-	TTATAAT	TTACAATT
UGT2B10	chr4	69886144	-	TC	-	TATATAA	TGAATATAA
CMA1	chr14	24977501	A	T	-	TTATAAA	ATATAAA
UFM1	chr13	38923863	T	C	+	TAATTTA	TAACTTA
IL22RA1	chr1	24469814	T	C	-	TACTATTG	TGCTATTG

Examples - variants in TFBS motifs

Gene	Chromosome	Position	Reference	Variant	dbSNP	Strand	Motif start	Motif end	Sequence logo	Name	Class	Family	ID
CHECK2	chr22	29137870	C	T	rs2236141	-	29137869	29137887		NFYA	Other Alpha-Helix	NFY CCAAT-binding	MA0060.2
GID4	chr17	17942708	G	A	Novel	+	17942694	17942709		NFYB	Other Alpha-Helix	NF-Y CCAAT-binding	MA0502.1
MED2B	chr4	17616214	G	T	rs2286773	+	17616212	17616223		USF2	Zipper-Type	Helix-Loop-Helix	MA0526.1
PALB2	chr16	23652769	C	G	Novel	-	23652765	23652776		ELK4	Winged Helix-Turn-Helix	Ets	MA0076.2
SMARCB1	chr22	24129129	G	T	rs11704810	+	24129126	24129137		NRF1	Other	NRF	MA0506.1

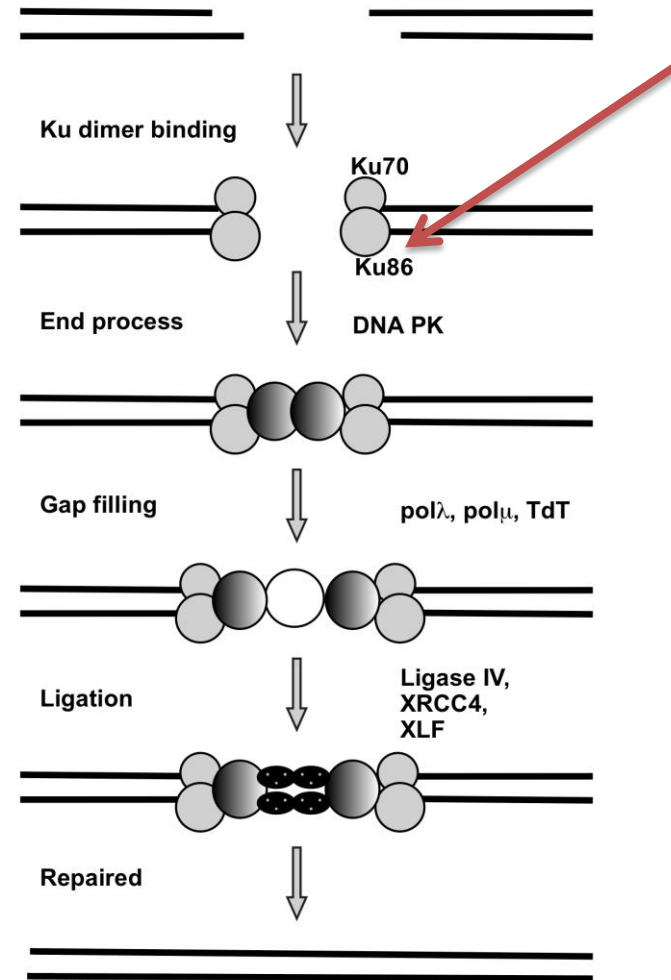
Example - a variant in Sp1 binding site



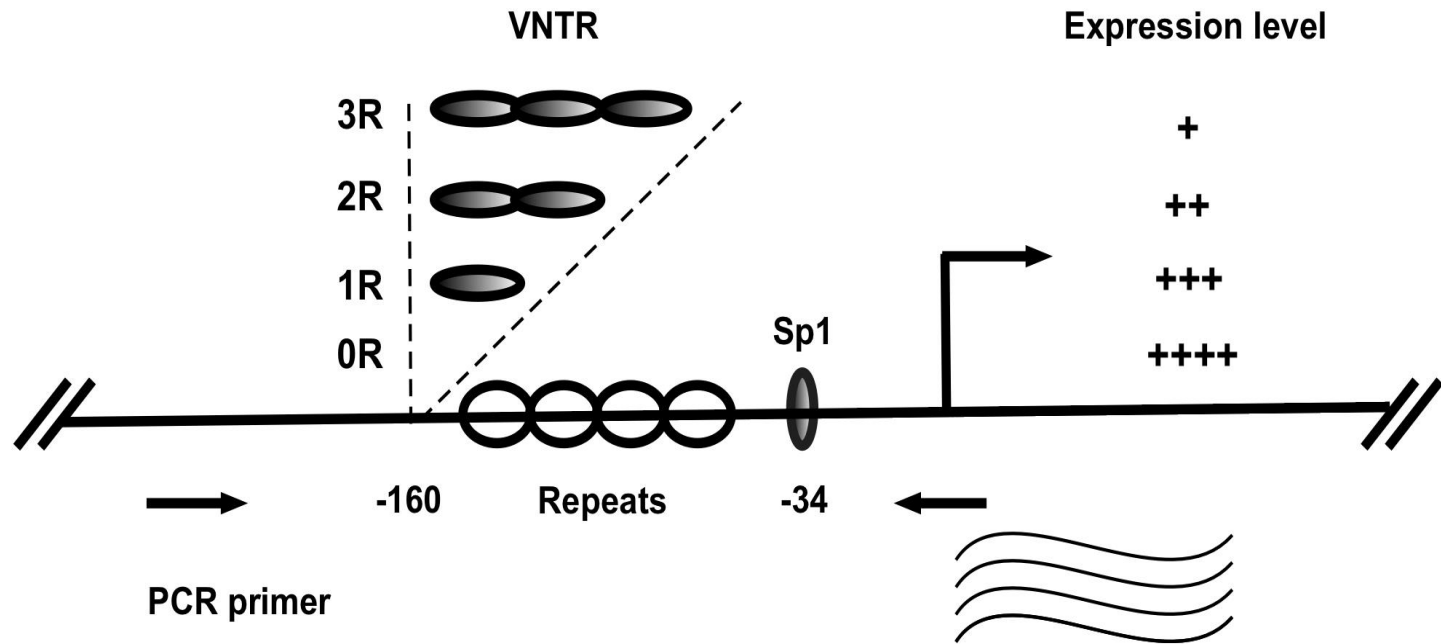
Example - Mapping variants in *XRCC5* promoter

XRCC5:

- Coding for Ku80
- Involve in non-homologous, double-strand break repair (NHEJ) pathway



XRCC5 promoter structure, variable number tandem repeat (VNTR) polymorphism, and expression regulation.

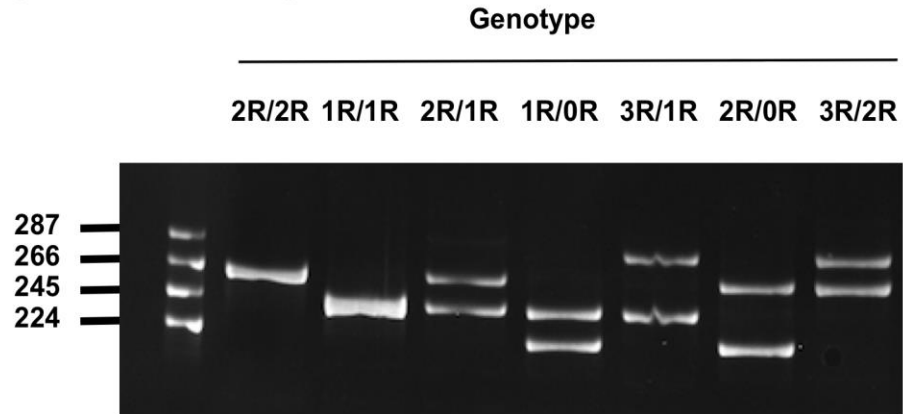


XRCC5 promoter VNTR in breast cancer

Genotype	Case number (%) [*]	
	Normal	BRCA1+
2R/2R	4 (7)	12 (28)
1R/2R	42 (72)	25 (58)
1R/1R	11 (19)	5 (12)
0R/2R	1 (2)	1 (2)
0R/1R	0 (0)	0 (0)
Total	58 (100)	43 (100)

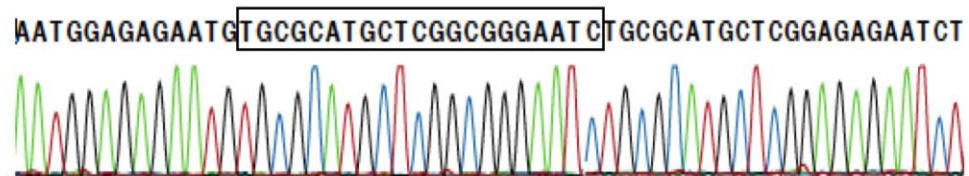
^{*} Fisher test: $p < 0.03$

A. VNTR genotype shown in PAGE gel

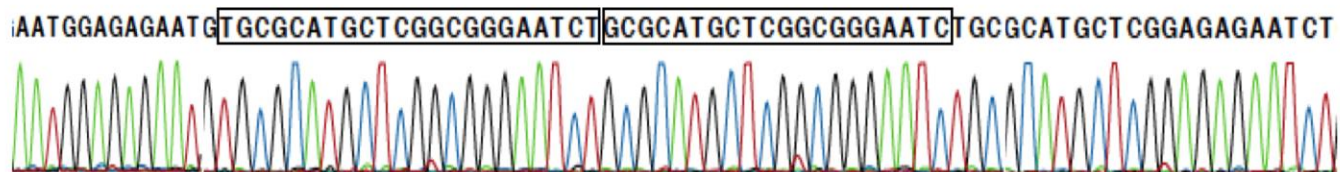


B. VNTR shown by Sanger sequencing

1R



2R



Abundant exome data publically available

- Exome Aggregation Consortium (ExAC): 60,706
- NHLBI GO Exome Sequencing Project (EVS6500): >6500
- 1000 genomes: 2,551
- TCGA breast cancer: 1,081
- GEO: 130
- dbGaP: ?
- ...?
- ...?

Summary

- GPES method provides a powerful tool for *de novo* promoter analysis, especially in the core-proximal region
- Explore existing exome data using GPES should provide extensive knowledge for promoter architecture in biology and medicine
- Genetic variation in promoter is likely far more complicated than currently consideration

Contribution

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Fengxia Xiao
Peixian Chen
Jiangtao Luo
San Ming Wang

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