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OMICS Group International is a pioneer and leading science event organizer, which publishes around 400 open access journals and conducts over 300 Medical, Clinical, Engineering, Life Sciences, Phrama scientific conferences all over the globe annually with the support of more than 1000 scientific associations and 30,000 editorial board members and 3.5 million followers to its credit.

OMICS Group has organized 500 conferences, workshops and national symposiums across the major cities including San Francisco, Las Vegas, San Antonio, Omaha, Orlando, Raleigh, Santa Clara, Chicago, Philadelphia, Baltimore, United Kingdom, Valencia, Dubai, Beijing, Hyderabad, Bengaluru and Mumbai. Crosstalk between acidic phospholipids present in bacterial membranes and DnaA, the initiator of Escherichia coli chromosomal replication

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> Both in prokaryotes and eukaryotes mother cell passes the exact same genetic information to its daughter cell.

> Moreover chromosomal origin of replication initiates DNA synthesis, once and only once per cell cycle.

> Interestingly a striking similarity exists between prokaryotic and eukaryotic initiator protein, which are considered to be the structural homologue.





Erzberger et al EMBO J 21, 4763-4773



Functional domains of DnaA protein



(R. Saxena et al., Int J Mol Sci. 2013: 14: 8517-37)

Architecture of E. coli chromosomal origin



Architecture of E. coli chromosomal origin



(R. Saxena, OA Biochemistry 2013: 1(2):13)

Acidic lipids in a fluid bilayer promote release of DnaA bound nucleotide



Sekimizu and Korenberg JBC 263:7131-35

Cell fractionation and microscopic studies reveal that DnaA protein resides on the membrane



K. Boeneman et al., Mol. Microbiol. 72:645-57.

A distinct region of DnaA protein is preferentially photolabeled



Garner J et al., JBC 273:5167-73

Functional Fragments of DnaA Protein Reveal the Membrane-binding Domain



Garner and Crooke, EMBO J. 15, 3477-85.

Screen for Mutations in *dnaA* That Can Suppress the Growth Arrest Phenotype of Acidic Phospholipid-deficient Cells



Growth arrested phenotype of phospholipid deficient cells can be restored by over-producing DnaA mutant present in membrane binding amphipathic region

HD L1001	N umb	er of Colo	nie s1
trans fo rm ed wit h	+IPTG	-IP	<u>IG</u>
plasmid s ex pressing	+ Gl ucose	+Glucose	+Arabi nose
D na A (L363R, L366E L367E, L369K)	E, 1000	0	506
D na A (L363K, L373F	R)1000	0	594
D na A (L363E)	1000	0	0
D na A (L363K)	1000	0	0
D na A (L366E)	1000	0	0
D na A (L366K)	1000	0	615
D na A (L367E)	1000	0	0
D na A (L367K)	1000	0	0
D na A (L373R)	1000	0	0

¹Values a renormalized to 1000 colonies for growth in the presence of I PTG and are an average from two or three independent transformations.

W. Zheng et al., *EMBO J* 20, 1164-72.

DnaA(L366K) is feeble in binding to low affinity recognitionn sequences but when augmented with low amount of DnaA can convert ORC to pre-RC



Outer space Mature Ipp Outer bacterial membrane PG processing Inner bacterial membrane Diacyl glycerol Cys21G vs21 **Pro-Lpp** Ribosome **Pro-Lpp** Cytoplasm Lpp-polypetide

Accumulation of immature Murein lipoprotein on inner membrane of E. coli Cells causes mucoidy





DnaA(L366K) can restore the altered morphology and cell viability of E. coli overproducing Murein lipoprotein



Future prospective:

- Whether the immature pro-lipoprotein accumulated on the inner membrane is causing toxicity for conversion of replication inefficient ORC to replication efficient pre-RC
- How DnaA-L366K bypasses this toxic affect
- A better understanding of how membrane domains of acidic phospholipids influence nucleotide exchange
- Studying the mutant could be an approach to know that if there is any cross talk between the acidic phospholipids present in bacterial membranes and initiation of chromosomal replication.

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Biochemistry of DnaA

- DnaA, the initiator protein initiates chromosomal replication by binding to high affinity and low affinity recognition elements present within *oriC*.
- Acidic lipids in a fluid bilayer promote release of bound nucleotide. Electrostatic interactions between anionic lipid head- groups and DnaA may stabilizes DnaA-membrane association.
- A discrete domain of DnaA protein that inserts into the hydrophobic region of acidic bilayers is essential for functional membrane interaction.
- Expression of DnaA protein with mutations in its membrane-binding domain suppresses the arrested growth of acidic phospholipid- deficient cells.
 - ADP-DnaA binds to only high affinity recognition sequences to produce replication inefficient origin recognition complex.
- ATP-DnaA binds to both high and low affinity recognition sequences and convert ORC to replication efficient pre-Replication Complex.

Architecture of E. coli chromosomal origin



ADP-DnaA forms an origin recognition complex by binding to *oriC* sites R1, R2, and R4.



ATP-DnaA can bind within both low affinity sites (such as I1, I2, I3, R1, R5M) and high affinity DnaA boxes (R1, R2, and R4) to form a pre-RC

Neither ATP- or ADP-DnaA(L366K) can unwind oriC



DnaA(L366K) cannot bind low affinity sites but when bound to a strong site can assist loading of wild-type DnaA to low affinity site



ATP-DnaA(L366K) and ADP-DnaA(L366K) confer the same DNase I protection to *dnaA* promoter as ATP-DnaA



DnaA(L366K) in either nucleotide form represses *in vitro* transcription from *dnaA* promoter similar to ATP-DnaA



	W	/T _{ATP}	W	T _{ADP}	L3	66K _{ATP}	L36	56K _{ADP}
		(% ex	pression)			(% exp	ression)	
nM protein	50	300	50	300	50	300	50	300
dnaAp1	60±5.8	10±3.2	98±2.8	52±13.0	75±7.6	21±3.2	73±4.7	20±2.1
dnaAp2	40±9.0	6±2.0	90±7.0	40±11.0	68±5.0	12±2.0	71±3.6	14±3.6

Each value is calculated as relative percentage of intensity obtained with no protein control (0 nM)





Hydrolysis of ATP bound to DnaA, is a slow process by itself



Colony Formation of HDL1001 Cells Transformed With Plasmids Harboring Deletion Mutant *dnaA* Genes

	Number of Colonies			
HDL1001 transformed	+IPTG	I	PTG	
with plasmids expressin	g+Glucose	+Glucose	+Arabinose	
Wild-type DnaA	1000	0	9	
DnaA∆317-322	1000	0	9	
DnaA∆322-329	1000	0	6	
DnaA∆329-333	1000	0	0	
DnaA∆332-337	1000	0	16	
DnaA∆336-340	1000	0	2	
DnaA∆340-345	1000	0	0	
DnaA∆346-356	1000	0	161*	
DnaA∆357-368	1000	0	517	
DnaA∆363-367	1000	0	700	
DnaA∆369-376	1000	0	442	
DnaA∆373-381	1000	0	577	
DnaA∆377-386	1000	0	407	
DnaA∆387-396	1000	0	764	
DnaA∆397-404	1000	0	227*	
DnaA∆405-417	1000	0	9	
DnaA∆418-422	1000	0	407	
DnaA∆432-427	1000	0	262	
DnaA∆428-435	1000	0	362*	
DnaA∆436-445	1000	0	667	
DnaA∆446-455	1000	0	714*	
DnaA∆456-464	1000	0	577	
(Vector)	1000	0	0	

Zheng et al, EMBO J <u>20</u>, 1164-1172.

Acidic Phospholipids Are Involved in Initiation of Replication at *oriC In Vivo*





DnaA(L366K) is competent at repressing the dnaA

<i>E. coli</i> strain	protein expressed	β-galatosidase activity (in Miller units)
EH2827/pBAD	none	250
EH2827 pZL606	DnaA	130
EH2827pZL606 (L366K)	DnaA(L366K)	125
M 182/p B A D	none	207
M 182/pZL606	DnaA	100
M182/pZL606 (L336K)	DnaA(L366K)	80

TABLE IDnaA(L 366K) is competent at repressing the dnaA promoter in vivo

Regulation of DnaA Protein Activity by Acidic Membranes



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