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3rd International Conference and Exhibition on Pathology

Molecular impacts of virus infections and genetic variants on the course of HBV-related liver diseases in Vietnam

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HBV infection

□ Worldwide

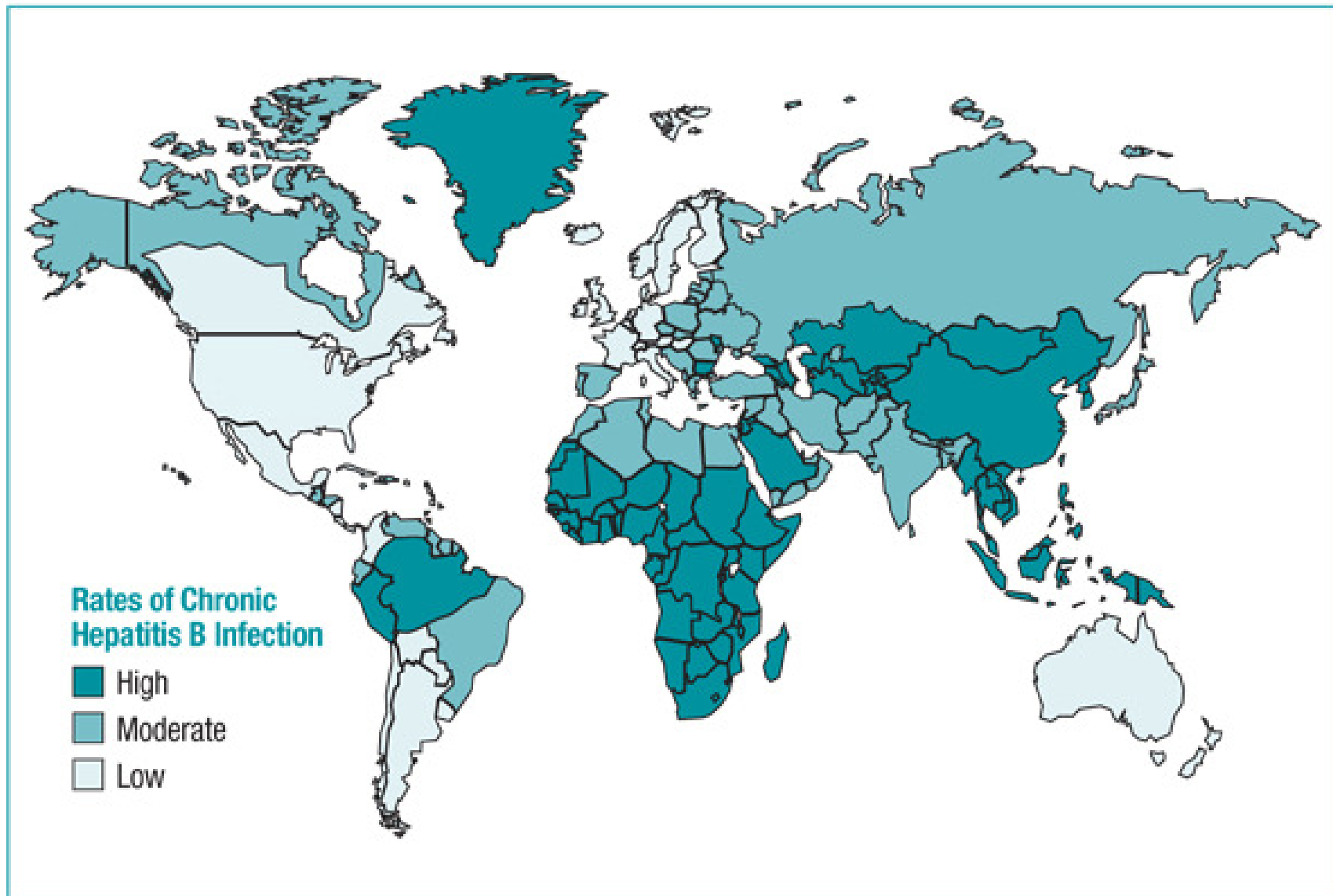
- HBV is leading cause of HBV-related liver diseases (AHB, CHB, LC and HCC)
- Approximately 240 million people are living with chronic hepatitis B
- 600,000 people die each year from HBV-related chronic liver diseases
- 500,000 new cases of HCC are diagnosed each year

□ Vietnam

- Prevalence of current HBV infection (HBsAg+), >10% in the population
- 10 million Vietnamese people with chronic HBV resulting in 23,300 deaths (2005)



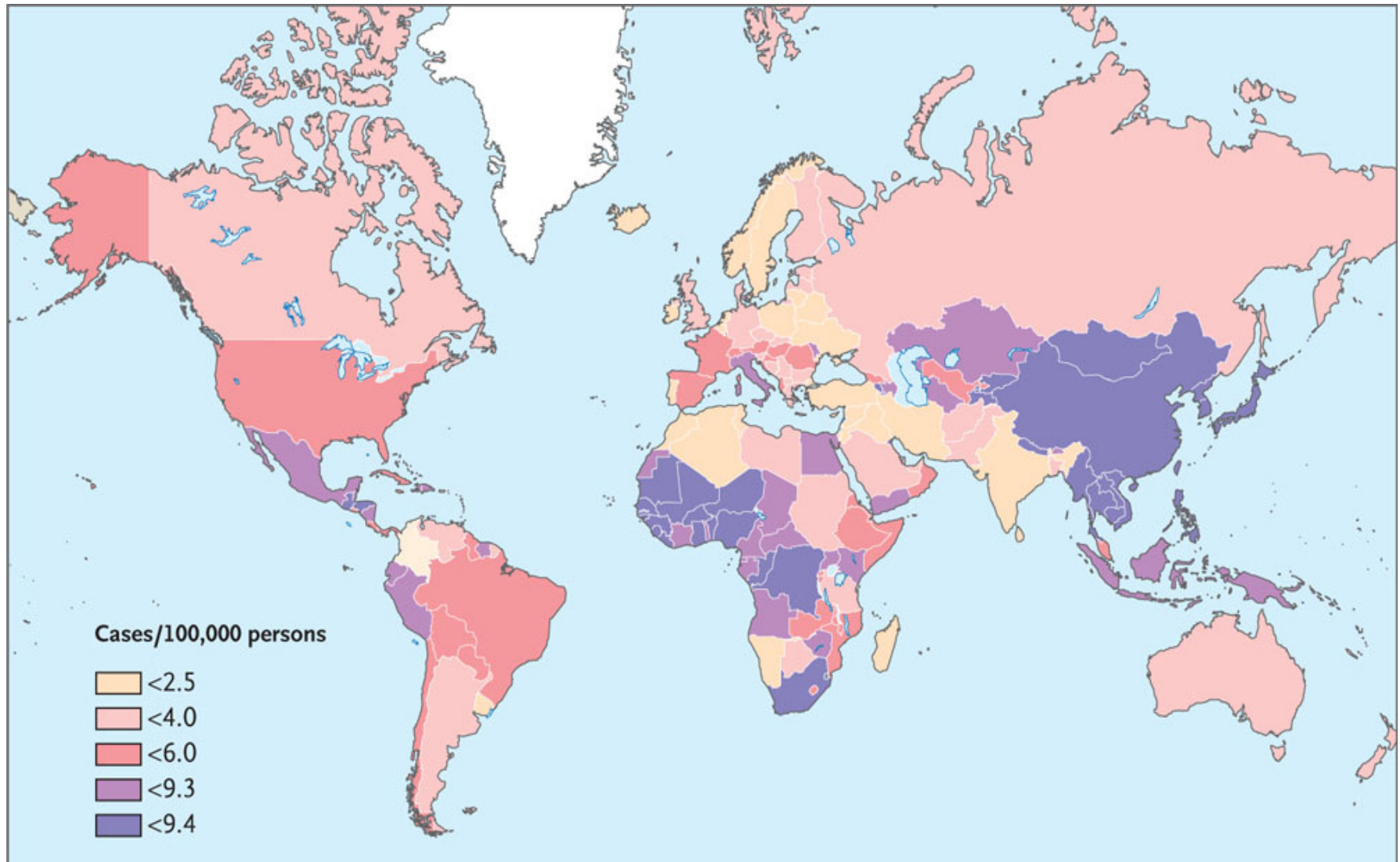
Chronic hepatitis B prevalence



Source: <http://www.cdc.gov>



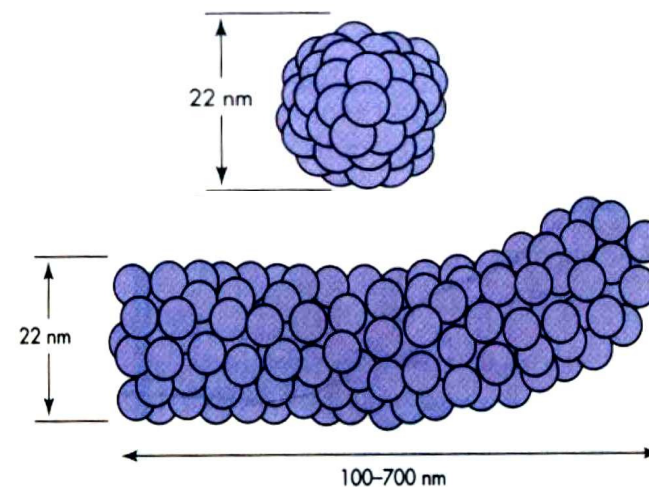
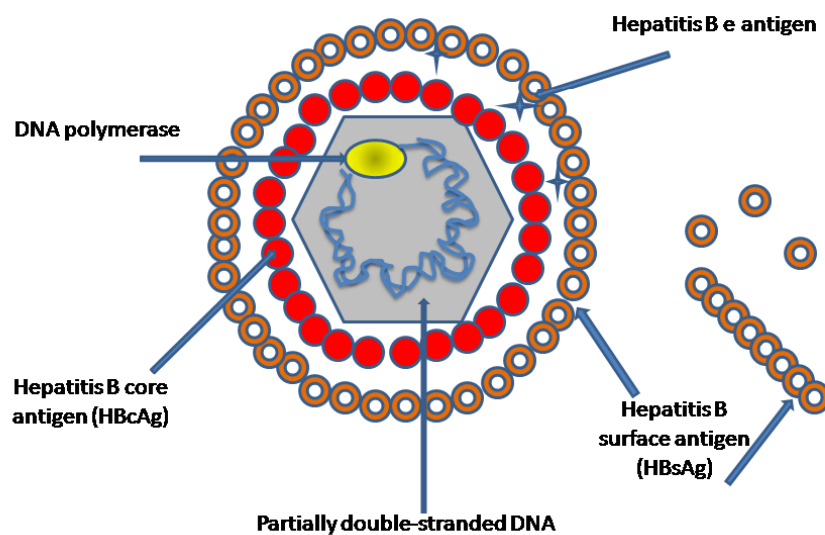
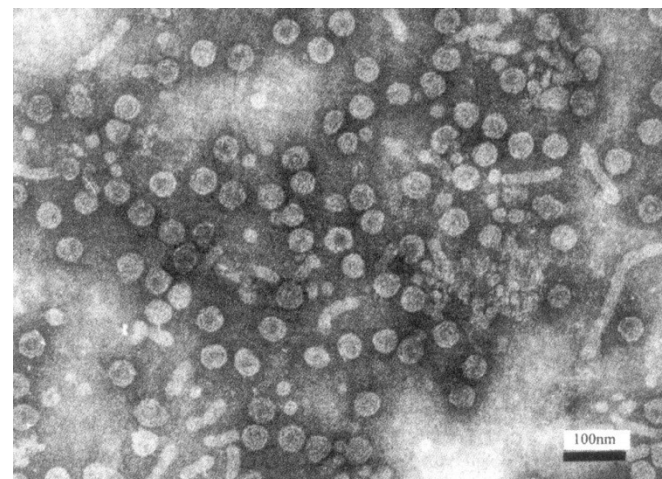
Estimated incidence of liver cancer





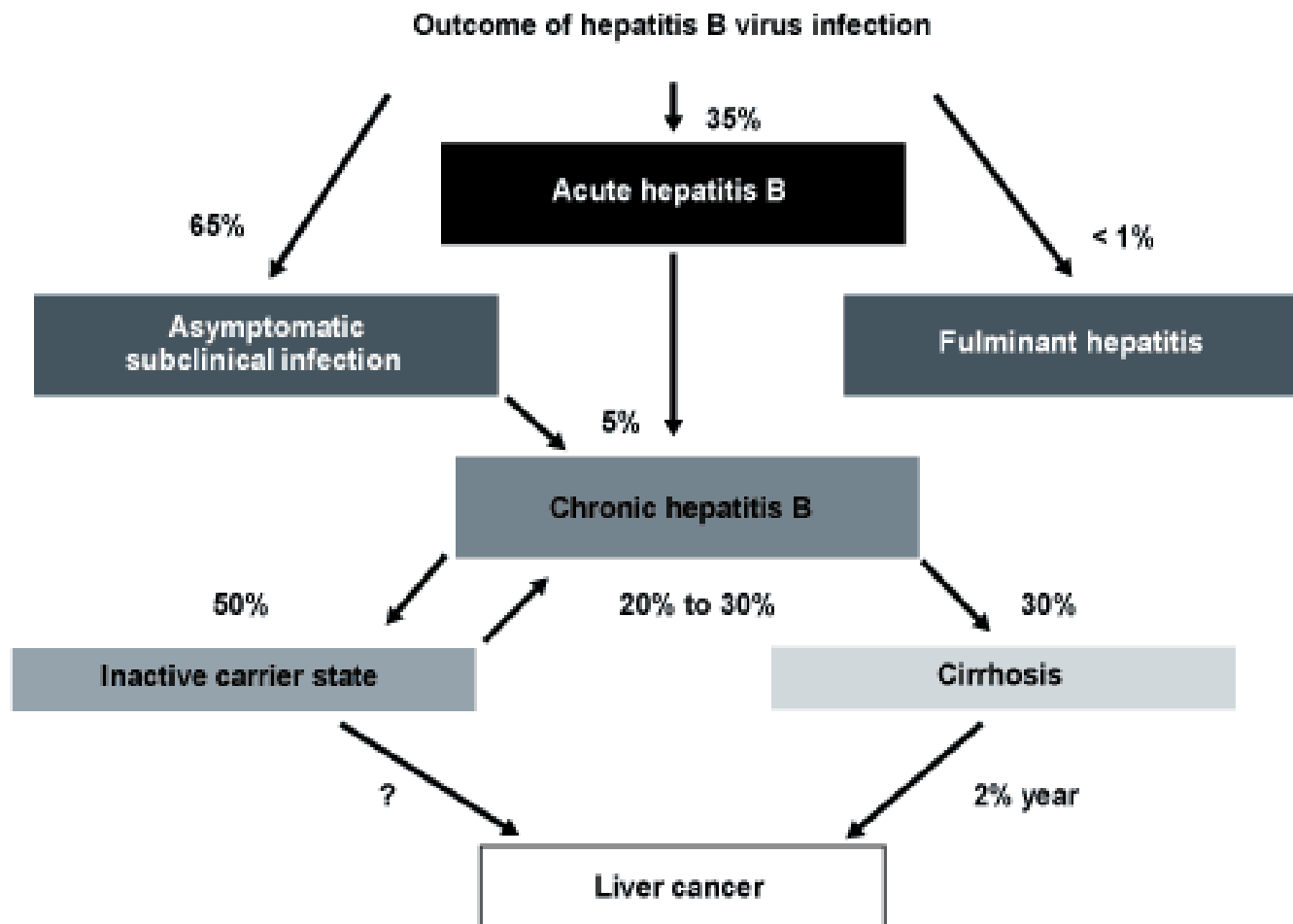
Hepatitis B virus

- ❖ Circular partially double-stranded DNA virus
- ❖ Genus: *Orthohepadnavirus*
- ❖ Family: *Hepadnaviridae*





Natural history of hepatitis B infection





HBV genotype and liver diseases

Table 4. Distribution of HBV Genotypes in Vietnamese Patients With HBV Infection

Genotype	ASYM n = 86 (%)	AHB n = 43 (%)	CHB n = 70 (%)	LC n = 92 (%)	HCC n = 84 (%)	Total n = 375 (%)
A	29 (33.7)	6 (13.9)	15 (21.4)	10 (10.8)	8 (9.5)	68 (18.13)
B	4 (4.6)	8 (18.6)	7 (10.0)	8 (8.7)	10 (11.9)	37 (9.86)
C	29 (33.7)	10 (23.3)	13 (18.6)	13 (14.1)	29 (34.5)	94 (25.06)
D	5 (5.8)	6 (13.9)	8 (11.4)	41 (44.6)	16 (19.0)	76 (20.26)
E	2 (2.3)	3 (6.9)	2 (2.8)	2 (2.2)	4 (4.8)	13 (3.47)
F	2 (2.3)	2 (4.6)	0	3 (3.3)	2 (2.4)	9 (2.40)
G	0	4 (9.3)	5 (7.1)	3 (3.3)	7 (8.3)	19 (5.06)
A/C	2 (2.3)	2 (4.6)	9 (12.8)	7 (7.6)	5 (6.0)	25 (6.66)
A/D	1 (1.2)	2 (4.6)	0	3 (3.3)	0	6 (1.6)
C/D	4 (4.6)	0	3 (4.3)	1 (1.1)	2 (2.4)	10 (2.66)
Other remaining genotype mixtures	8 (9.3)	0	8 (11.4)	1 (1.1)	1 (1.2)	18 (4.80)

Table 5. Distribution of Genotype Mixtures in HBV-Infected Vietnamese Patients

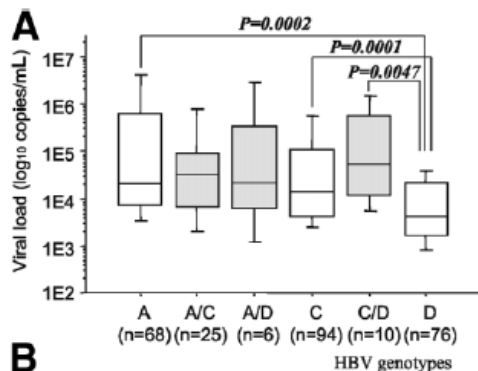
Genotype	ASYM n = 86 (%)	AHB n = 43 (%)	CHB n = 70 (%)	LC n = 92 (%)	HCC n = 84 (%)
Mixtures	15 (17.4)	4 (9.3)	20 (28.6)*	12 (13)	8 (9.5)
Single	71 (82.6)	39 (90.7)	50 (71.4)	80 (87)	76 (90.5)

* $P < .02$ in comparison to AHB, LC, and HCC groups.

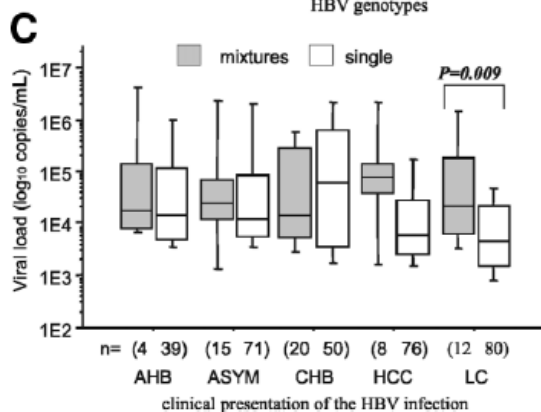
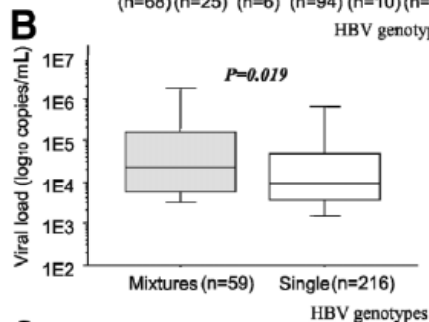
HBV-genotype mixtures were more frequent in Vietnam (59/375; ~16%)



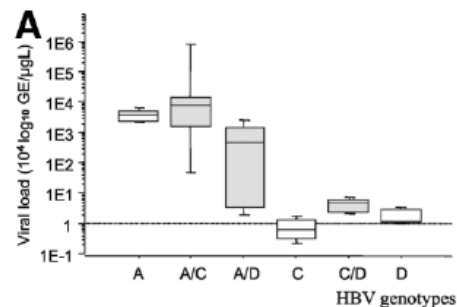
HBV genotype and liver diseases



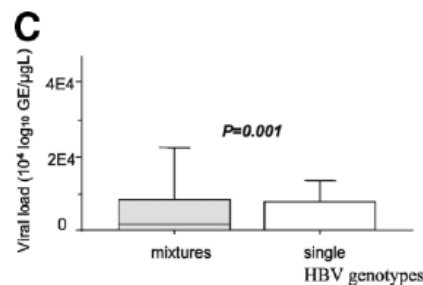
Genotypes	P value
A vs C	0.0159
A vs D	0.0002*
A/C vs D	0.0012
A/D vs D	0.0079
C vs D	0.0001*
C/D vs D	0.0047*



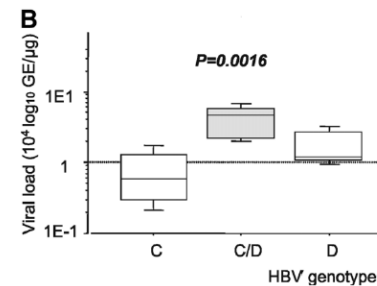
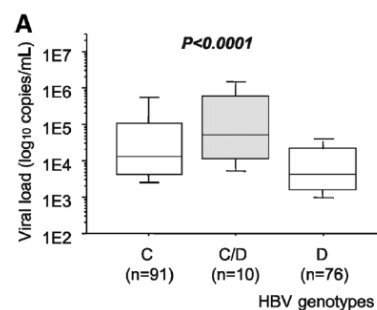
In patients



Genotypes	P value
A vs A/D	0.0001
A vs C	0.0002
A vs C/D	0.0009
A vs D	0.0009
A/C vs A/D	0.0015
A/C vs C	<0.0001
A/C vs C/D	0.0002
A/C vs D	0.0002
A/D vs C	0.0003
A/D vs D	0.0067
C vs C/D	0.0015
C vs D	0.0451
C/D vs D	0.0280



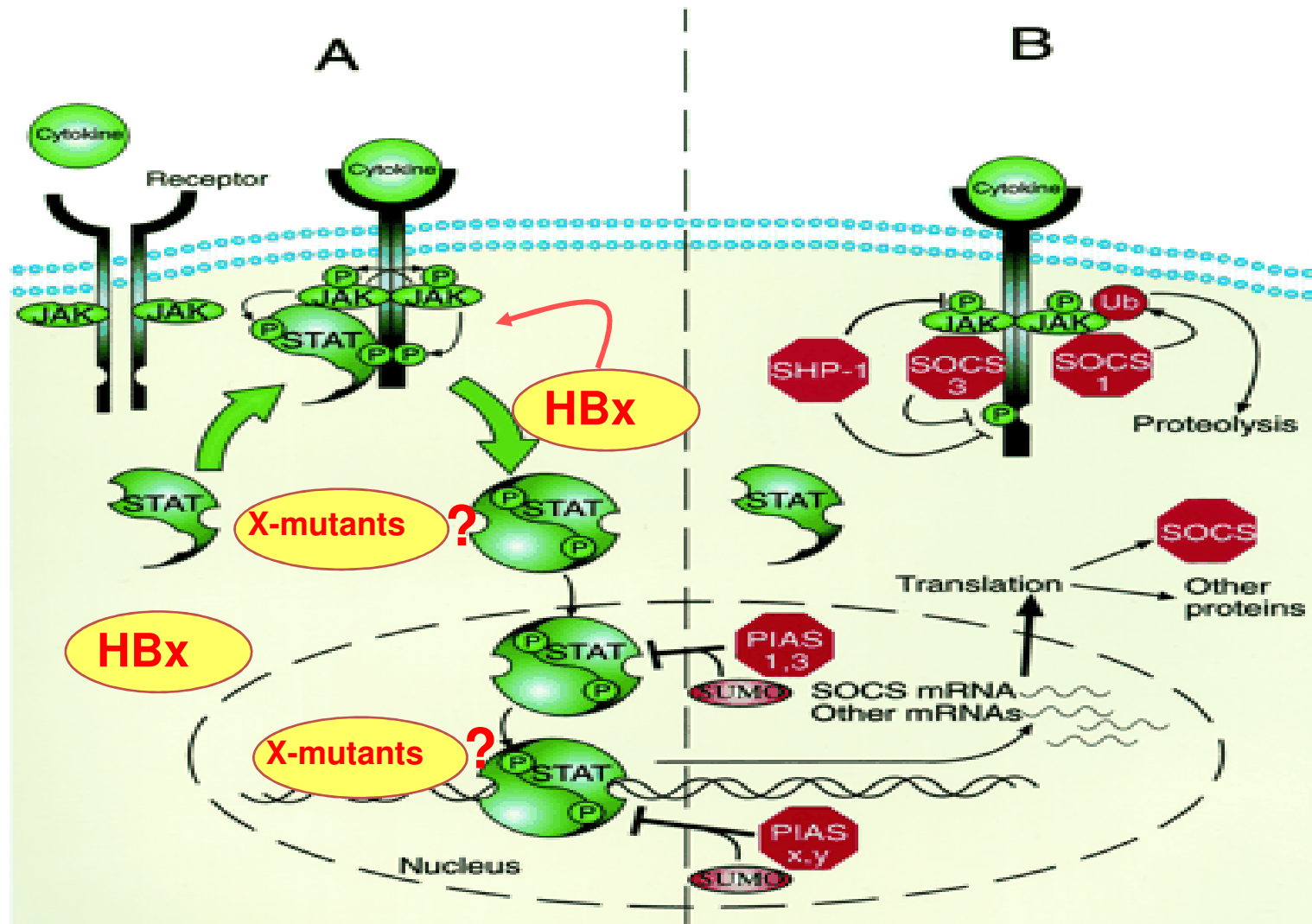
In vitro experiments



Conclusion: Mixed HBV-genotype infections are associated with altered pathogenesis and clinical outcome of liver disease

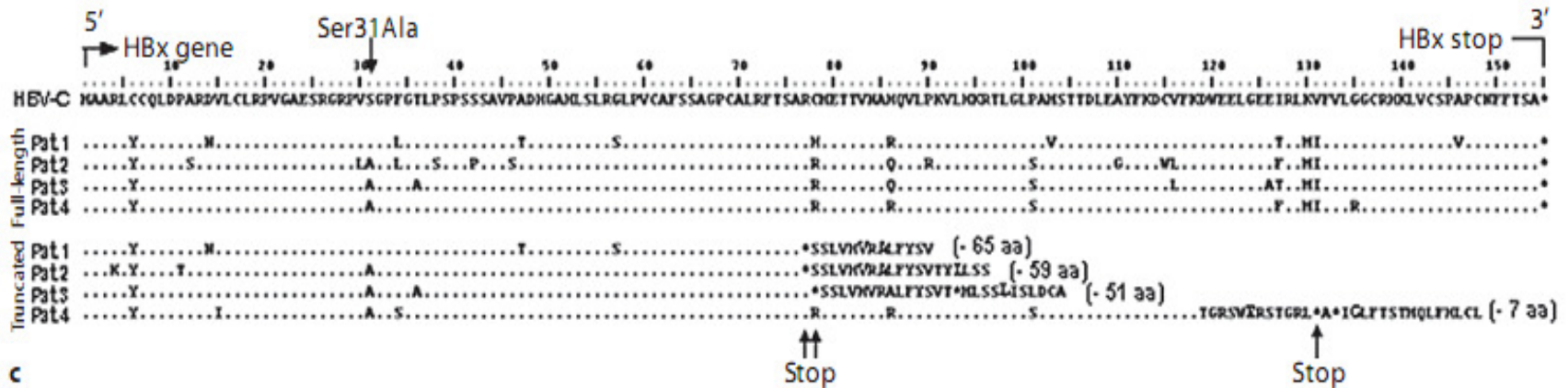


JAK/STAT/SOCS signaling

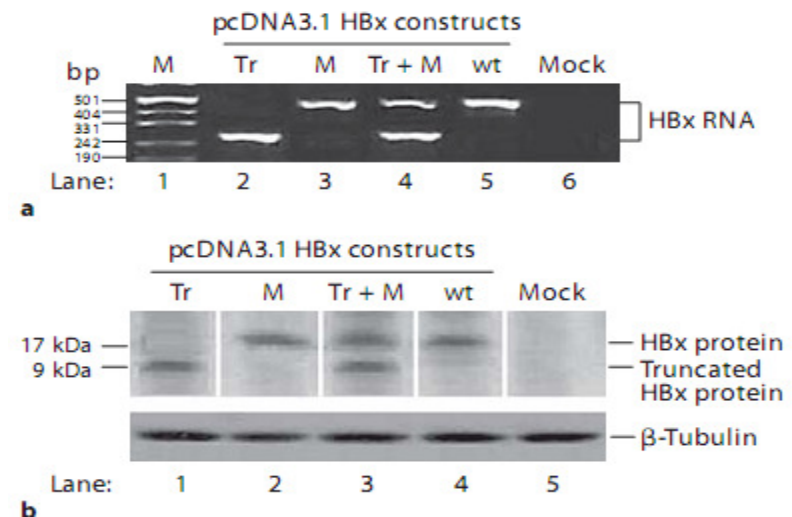




HBx Mutants Dysregulate STAT/SOCS Signaling

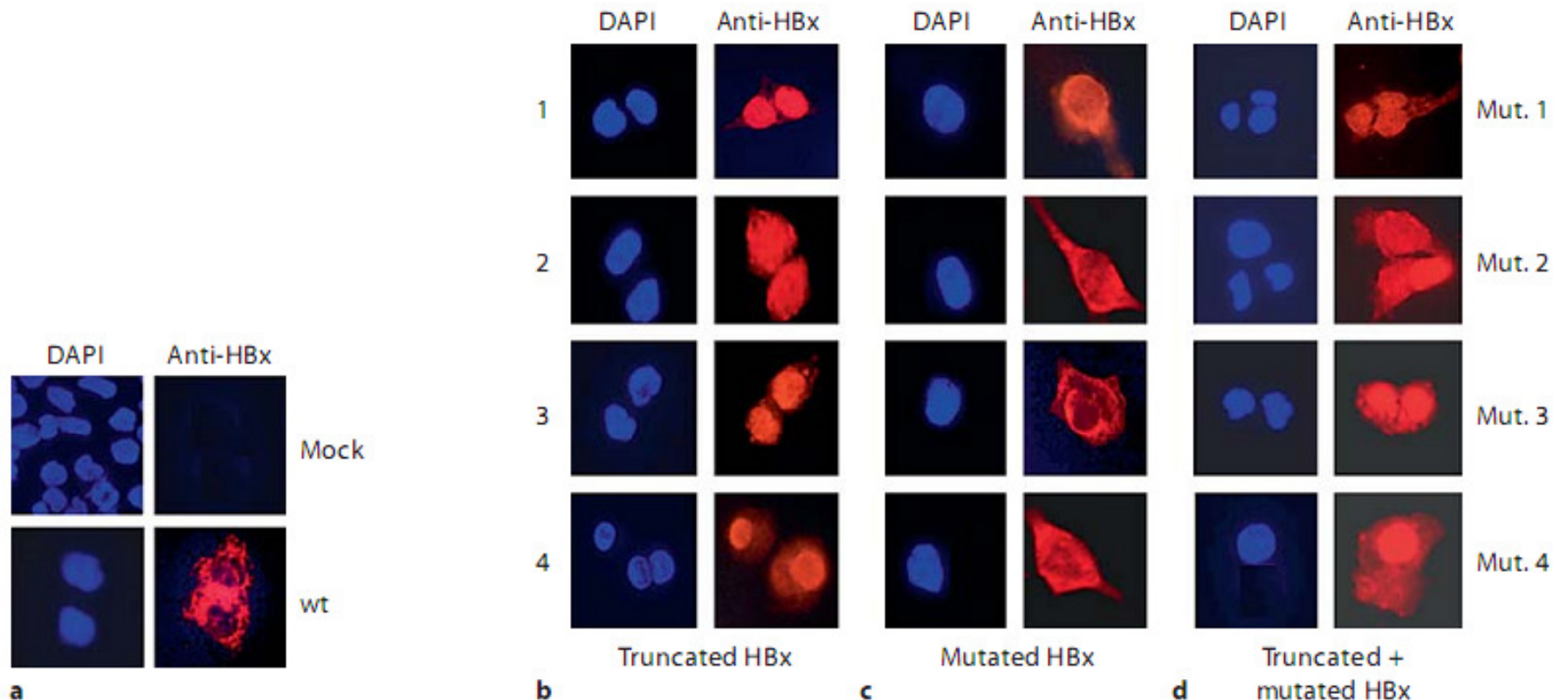


In HCC patients shown random mutations throughout HBx-gene, and 4/48 HCC patients were combination to stop codon insertions at codons 77, 78 and 131 together with HBx gene deletions.





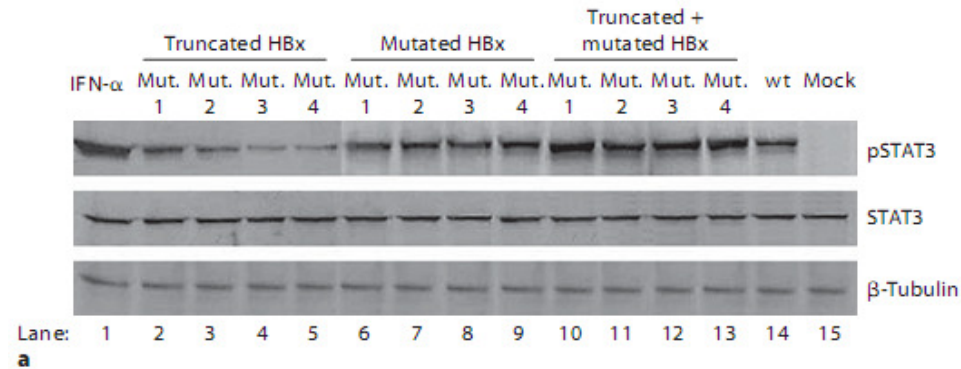
HBx Mutants Dysregulate STAT/SOCS Signaling



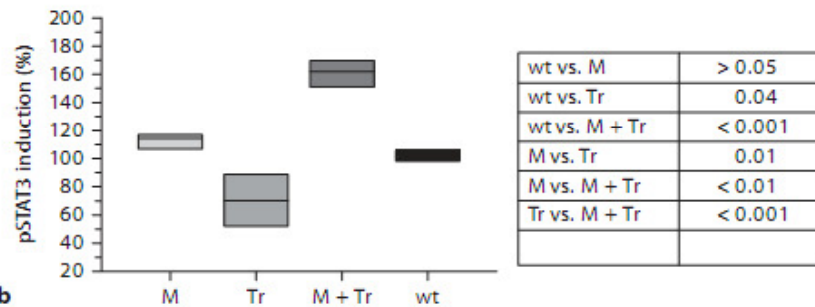
A different distribution of HBx mutant proteins in transfected cells, wt-HBx protein were predominant distributed in cytoplasm. The truncated HBx-protein is mainly distributed in the nuclei, randomly mutated HBx protein had a predominant perinuclear localization and the combination of truncated and randomly mutated HBx is distributed into both the nuclei and cytoplasm.



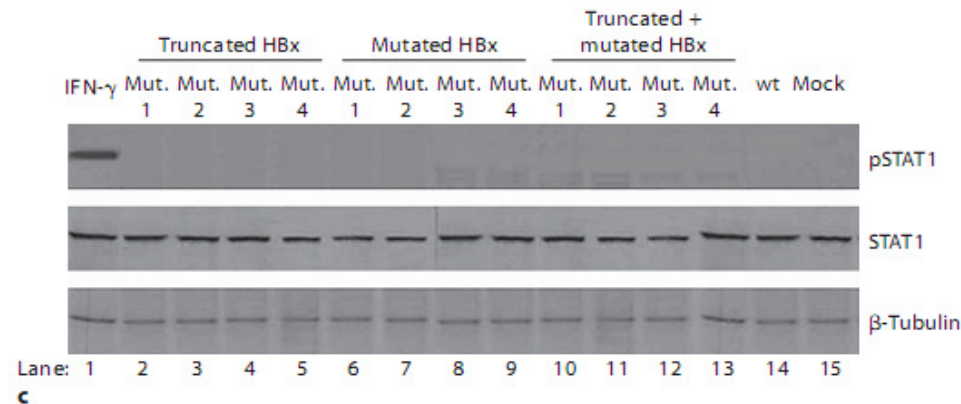
HBx Mutants Dysregulate STAT/SOCS Signaling



a



b

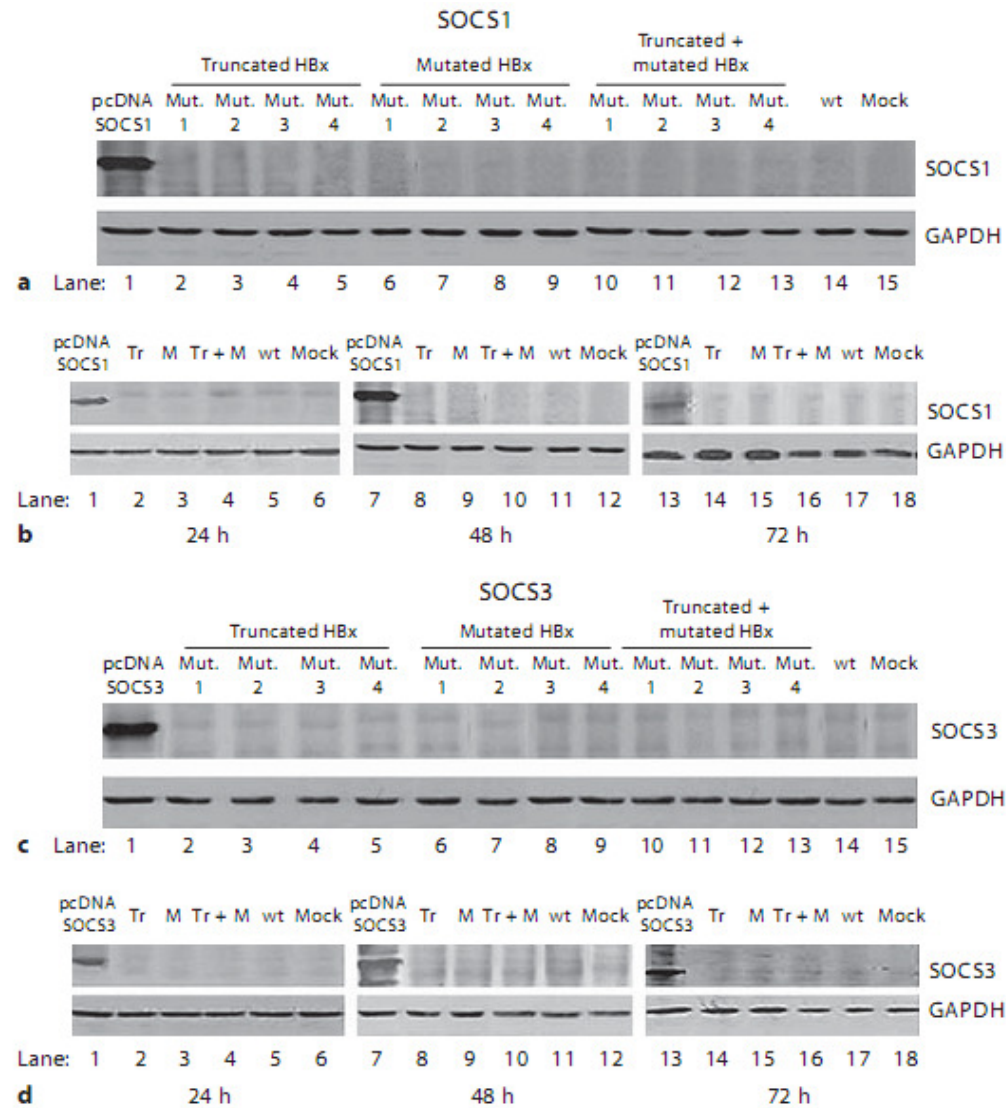


c

Conclusion: Mutated HBx-protein activates STAT-3 but not STAT-1.



HBx Mutants Dysregulate STAT/SOCS Signaling



There were no activation SOCS3 and SOCS1 proteins in Wt or mutated HBx-proteins.



Interferon alpha 2 and HBV infection

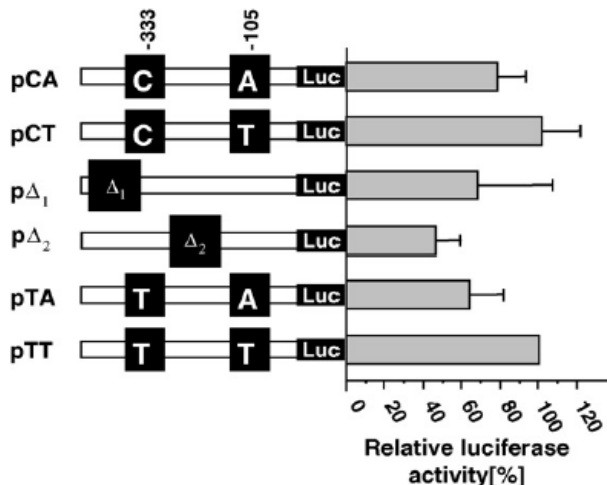
- IFN α is a critical mediator of immunity to hepatitis B virus (HBV) infection
- IFN has been used in the treatment of viral hepatitis. In this study we investigated the mutated IFN α gene in 344 HBV-infected patients and 293 HC control

Allelic distribution of variants in *INF- α* promoter region in patients and controls

	Vietnamese			
	HBV-		HBV+	
	Wild type	Variant	Wild type	Variant
C-105T	1	0	1	0
T-333A	1	0	0.99	0.01
Δ_1	1	0	1	0
Δ_2	0.89	0.11	0.81	0.19

Allelic frequency and OR of the deletion in *INF- α* promoter region in patients and controls

Diagnosis (n)	Δ_2 n (frequency)	OR (CI)	P
AHB (n = 82)	17 (0.21)	2.2 (1.1–4.1)	0.020
CHB (n = 124)	29 (0.23)	2.6 (1.5–4.3)	<0.001
LC (n = 180)	28 (0.16)	1.6 (0.9–2.6)	n.s.
HCC (n = 230)	44 (0.19)	2.0 (1.3–3.1)	0.002
Asym (n = 72)	8 (0.11)	1.1 (0.4–2.3)	n.s.
Total HBV-infected (n = 688)	126 (0.18)	1.9 (1.4–2.7)	<0.001
HC (n = 586)	62 (0.11)	n.a.	n.a.



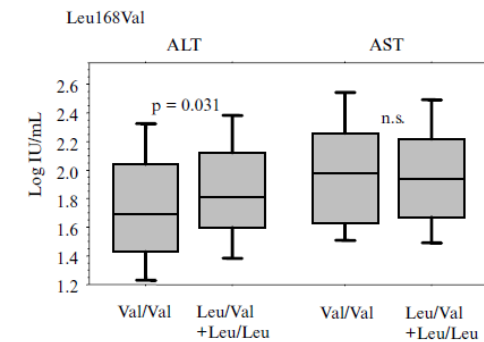
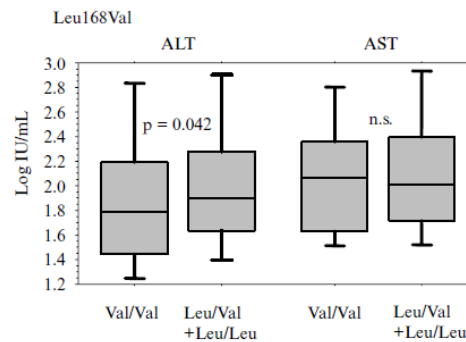
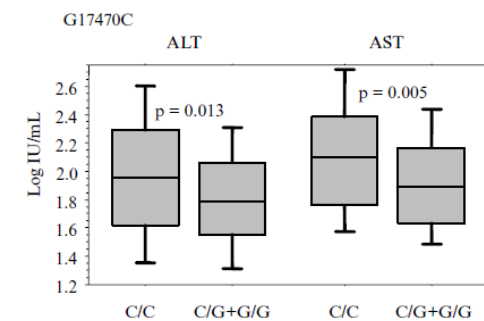
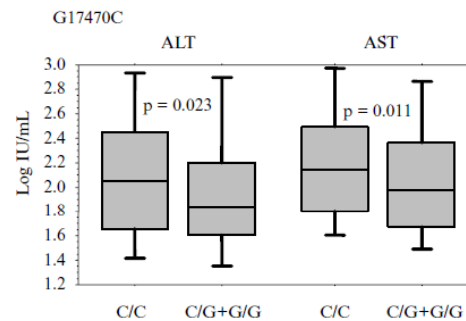
Conclusion: Deletion in the promoter of the *INF- α 2* gene reduces the transcription in vitro and was susceptibility to hepatitis B



Interferon alpha receptor 1 and HBV infection

- 17470C allele (rs1012335) in *IFNAR1* was more frequent in HBV-infected patients (OR: 2.6; $p < 0.001$)
- G allele (rs2257167, cause Val to Leu) was more frequent in the healthy control (OR: 0.54, $p = 0.004$)

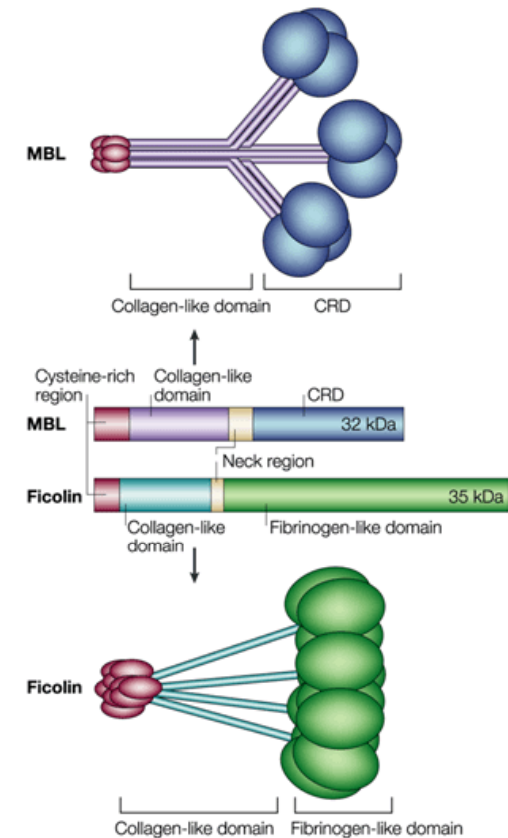
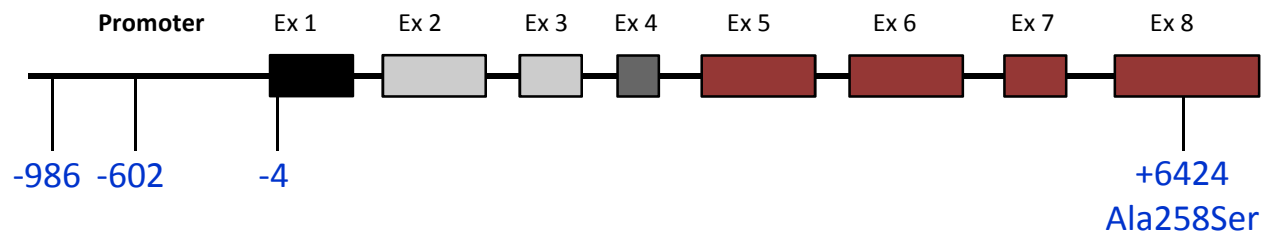
Conclusion: two variants of *IFNAR1* gene are associated with clinical outcomes of HBV infection





Ficolin-2 and *FCN2* polymorphism

- Soluble pattern recognition molecules
- Produced mainly in liver
- Recognize pathogen recognition receptors (PRRs)
- Activate complement system via lectin pathway
- Clearance of apoptotic cells
- SNPs (-986G/A, -602G/A -4A/G and +6424G/T) are associated with ficolin-2 levels and diseases



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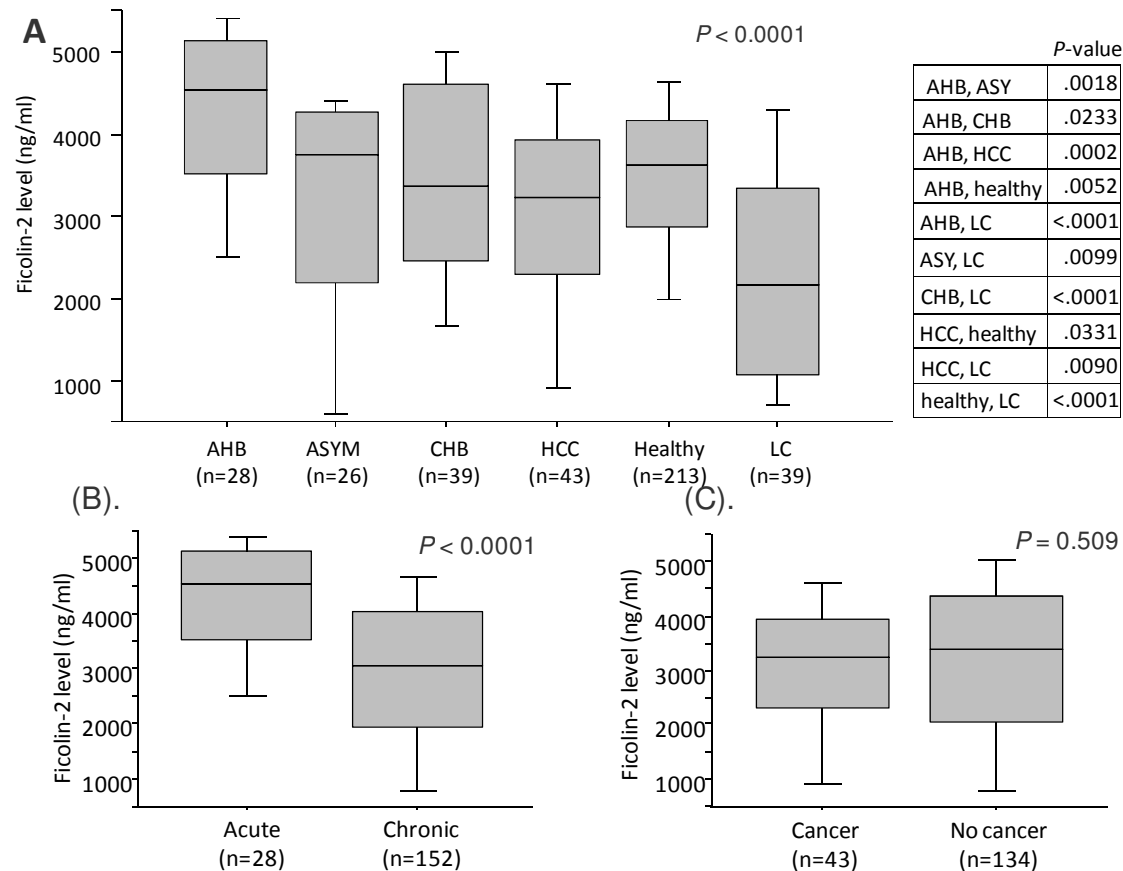


FCN2 polymorphism and HBV infection

Haplotype	Cases (%)	Controls (%)	OR (95%CI)	P
-986/-602/-4/+6424				
	AHB (n=92)	Controls (n=606)		
GGAG	64 (69.6)	411 (67.8)	NA	NS
GGAT	24 (26.1)	122 (20.1)	NA	NS
AAAG	2 (2.2)	15 (2.5)	NA	NS
AGGG	2 (2.2)	58 (9.6)	0.2 (0.02-0.8)	0.02
	HCC (n=224)	Controls (n=606)		
GGAG	159 (71)	411 (67.8)	NA	NS
GGAT	56 (25)	122 (20.1)	NA	NS
AAAG	8 (3.6)	15 (2.5)	NA	NS
AGGG	HBV Patients (n=796) 1 (0.1)	58 (9.6)	0.04 (0.001-0.25)	<0.0001
		Controls (n=606)		
GGAG	561 (70.5)	411 (67.8)	NA	NS
GGAT	172 (21.6)	122 (20.1)	NA	NS
AAAG	27 (3.4)	15 (2.5)	NA	NS
AGGG	36 (4.5)	58 (9.6)	0.4 (0.28-0.7)	0.0002



Ficolin-2 levels in HBV infection



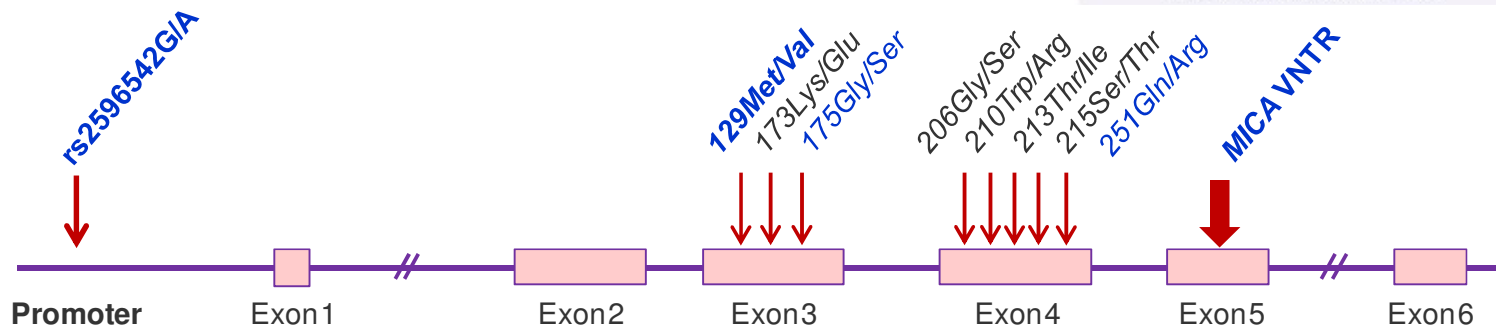
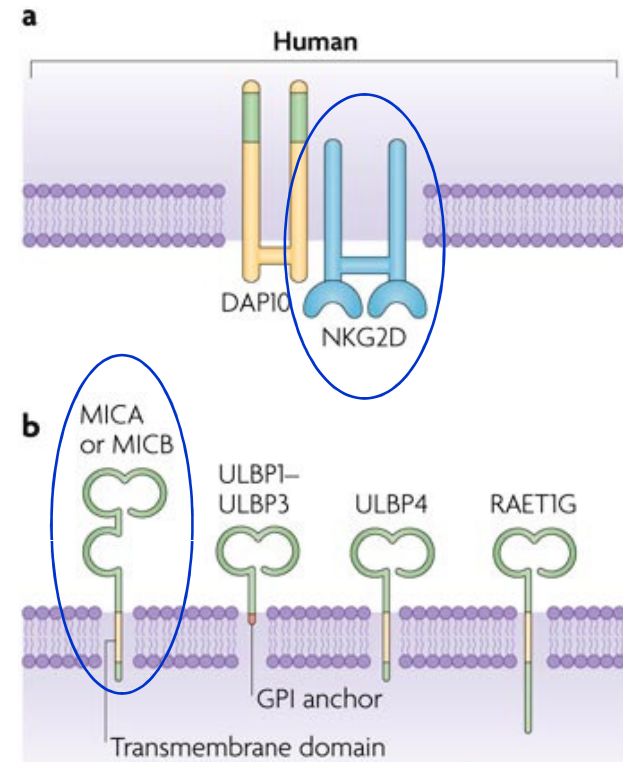
Soluble ficolin-2 levels were significantly increased in acute groups and significantly decreased in CHB, LC and HCC.

sFicolin-2 protein was significantly modulated during infection, and the high level of serum ficolin-2 significantly contributed to immune response against HBV infection.



MICA : gene and protein

- The human major histocompatibility complex class I (MHC) chain-related gene A/B (**MICA/MICB**)
- Ligands for NKG2D receptors of NK, NKT cells
- Members of *MIC* gene family including **MICA**, MICB and *MICC*to *MICG*
- *MICA* polymorphism is associated with many diseases (autoimmune, infectious diseases and cancer)





MICA polymorphisms and HBV-related HCC

HCC versus LC				
	OR	95% CI	P-value	Best fit model
rs2596542G/A	1.6	1.13-2.3	0.006	Allelic
MICA-129 (rs1051792)	1.5	1.04-2.1	0.03	Allelic
MICA-175 (rs1131896)	1.7	0.97-2.9	0.05	Gly/Gly (GG)
MICA-251 (rs1063635)	1.6	1.1-2.3	0.008	allelic
A5/A5.1	0.4	0.2-0.9	0.015	Genotype
A6/A9	5.4	1.2-49.9	0.02	Genotype
A4	1.8	1.02-3.2	0.04	Allele
A5	0.7	0.5-0.99	0.03	Allele
GGAG-A5	0.64	0.45-0.9	0.009	Haplotype
AAGA-A4	2.3	1.04-5.4	0.04	Haplotype
AGAA-A6	3	0.93-12.7	0.05	Haplotype

HCC versus CHB				
	OR	95% CI	P-value	Best fit model
rs2596542G/A	1.5	1.1-2.1	0.01	Allelic
MICA-129 (rs1051792)	2.3	1.1-5	0.02	Recessive
MICA-175 (rs1131896)	0.47	0.3-0.8	0.0015	Heterozygote advantage
A5/A5.1	0.35	0.15-0.8	0.005	Genotype
A5/A6	4.25	1.3-17.7	0.006	Genotype
AAGA-A4	2.1	1-4.6	0.04	Haplotype
GGAA-A5	0.06	0.001-0.4	0.0001	Haplotype

Variants in MICA gene were significantly associated with HCC susceptibility from CHB and LC patients. Some variants contributed to an increased risk of HCC (Odds ratio > 1), Exp. promoter variant rs2596542 contributed to increased risk of HCC with the OR is 1.6.



MICA polymorphisms and HBV persistence

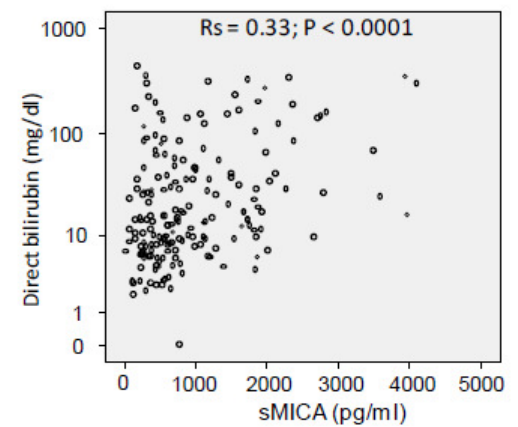
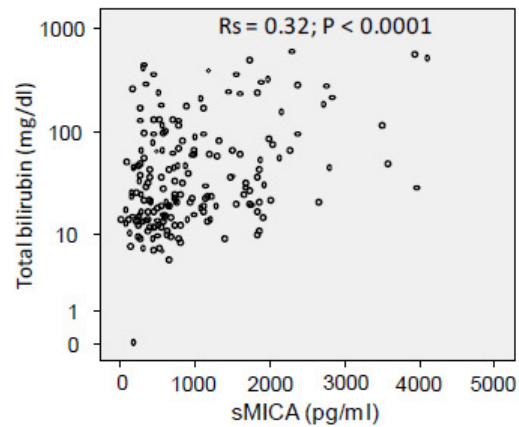
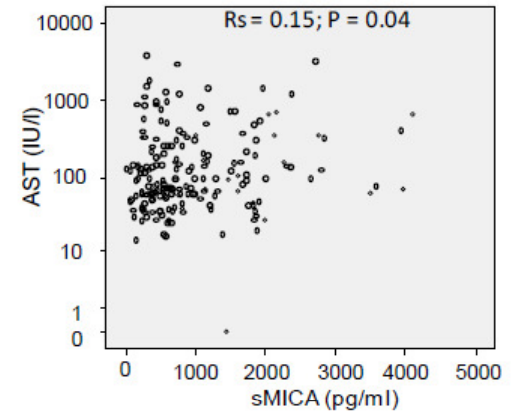
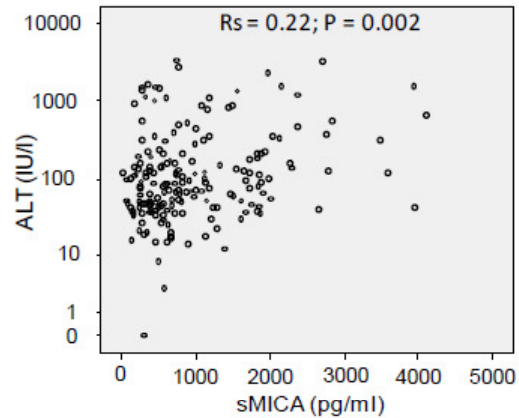
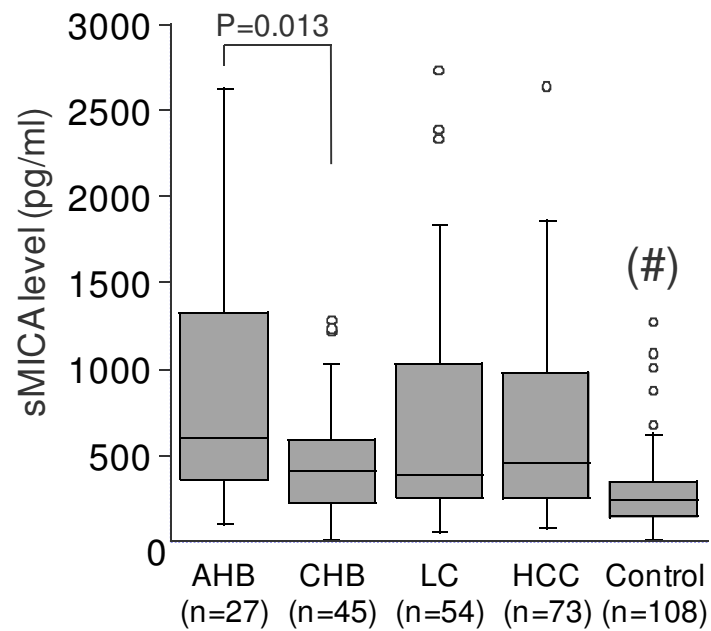
HBV cases vs. healthy control

	OR	95% CI	P-value	Best fit model
MICA-175 (rs1131896)	1.3	1.1-1.5	0.0095	Allelic
A5/A5	1.6	1.14-2.34	0.005	Genotype
A9/A9	0.47	0.24-0.9	0.02	Genotype
A5	1.24	1.02-1.5	0.02	Allele
A6	1.5	1.02-2.2	0.04	Allele
A9	0.8	0.6-0.97	0.02	Allele
GGGG-A5	2.4	1-6.7	0.051	Haplotype

To comparison between groups of patients infected with HBV and healthy controls, the polymorphisms in MICA were significantly associated with HBV infection and HBV persistence.



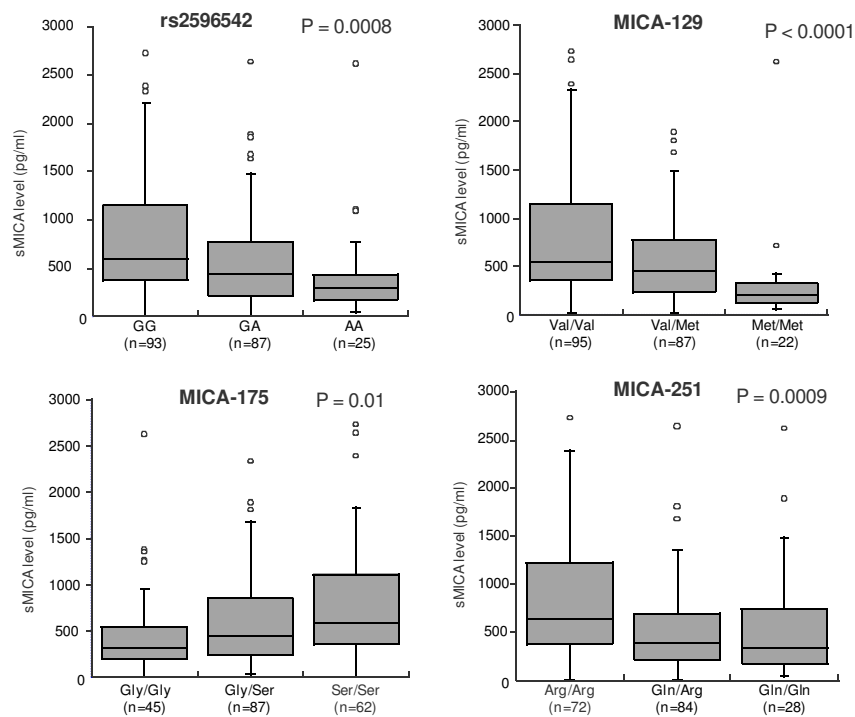
sMICA levels and HBV outcomes



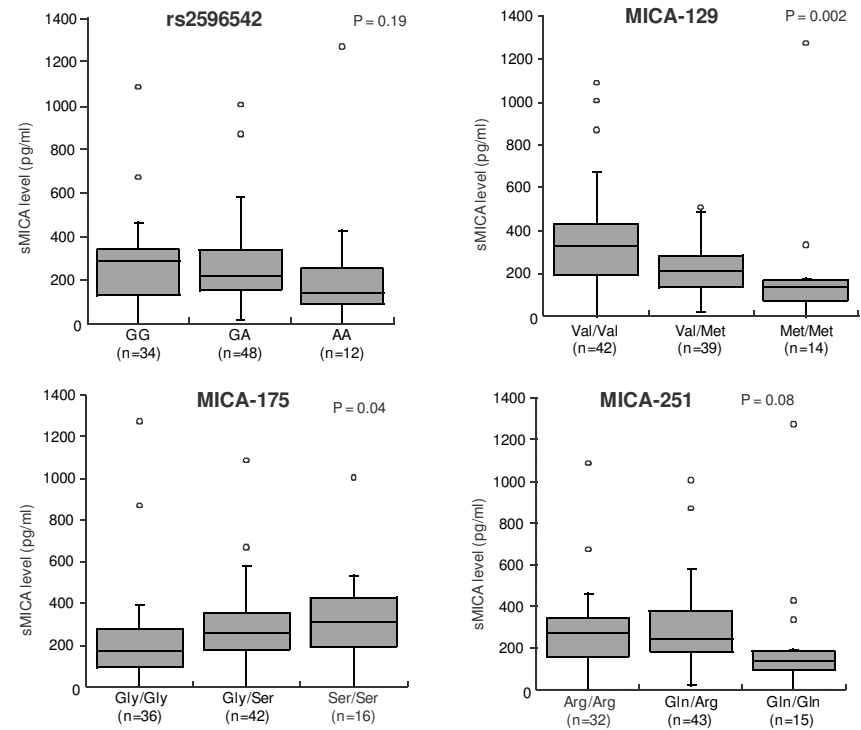


sMICA levels according to *MICA* SNPs

in patients



in healthy controls





Main message: MICA

- ❖ Polymorphisms in *MICA* gene are associated with HBV persistence, HBV-induced HCC
- ❖ *MICA* polymorphisms modulate MICA protein serum levels
- ❖ MICA protein modulate HBV infection, liver disease progression and tumor surveillance



CISH and HBV infection

- Cytokine-inducible SRC homology 2 (SH2) domain containing protein (*CISH*)
- SOCS family of proteins (suppressor of cytokine signaling)
- *CISH* controls the signaling of a variety of cytokines, in particular **IL-2**
- *CISH* polymorphism (e.g. **rs414171**) associated with the susceptibility to different infectious diseases



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CISH and HBV infection

-292A>T (#rs414171)	HBV Patients (n=473)	Healthy controls (n=416)	OR (95% CI)	P value
Genotype				0.029
AA	177 (37.4)	169 (40.6)	Reference	
AT	214 (45.3)	201 (48.3)	1.02 (0.76-1.37)	0.91
TT	82 (17.3)	46 (11.1)	1.7 (1.1-2.65)	0.012
Allele			1.22 (1-1.49)	0.04
Allele A	568 (60.04)	539 (64.8)		
Allele T	378 (39.96)	293 (35.2)		
Dominant			1.14 (0.87-1.51)	0.33
Wild type	177 (37.4)	169 (40.6)		
AT+TT	296 (62.6)	247 (59.4)		
Recessive			1.69 (1.23-2.54)	0.0078
AA+AT	391 (82.7)	370 (88.9)		
Mutant	82 (17.3)	46 (11.1)		

No significant association of *CISH* variant rs414171 with clinical parameters



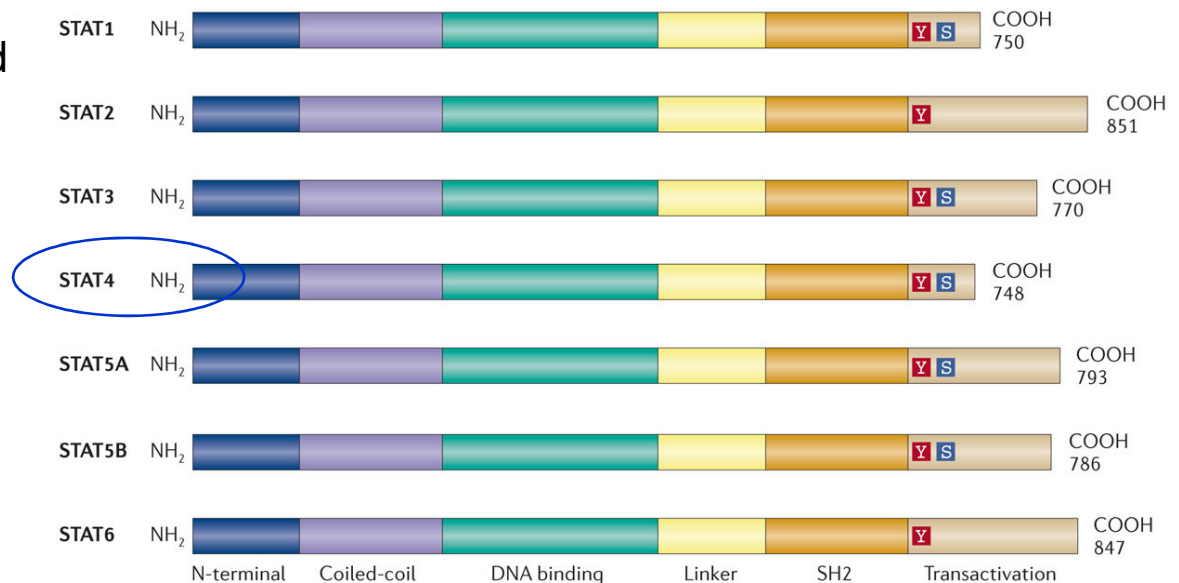
STAT4 and HBV-induced HCC

STAT protein family (signal transducer and activator of transcription)

Transduce signals from cytokine-receptor complexes and regulate transcription of specific genes

Response mainly to IL-12, IL-23, interferon type I and regulates the transcription of various genes including IFN- γ via the JAK/STAT pathway

Variants in *STAT4* (e.g. **rs7574865**) are associated with several autoimmune diseases





STAT4 and HBV-induced HCC

STAT4 rs7574865G/T	CHB n(%)	LC n(%)	HCC n(%)	HCC vs. CHB	
				OR(95% CI)	P value
GG	86 (41.7)	112 (50.5)	117 (49)	reference	
GT	92 (44.7)	87 (39.2)	102 (42.7)	0.9 (0.7-1.15)	0.41
TT	28 (13.6)	23 (10.3)	20 (8.3)	0.7 (0.45-0.98)	0.039
Allele					
G	264 (64.1)	311 (70)	336 (70.3)	reference	
T	148 (35.9)	133 (30)	142 (29.7)	0.84 (0.7-0.99)	0.048
Dominant					
GG	86 (41.7)	112 (50.5)	117 (49)	reference	
GT+TT	120 (58.3)	110 (49.5)	122 (51)	0.85 (0.7-1.1)	0.16
Recessive					
GG+GT	178 (86.4)	199 (89.6)	219 (91.6)	reference	
TT	28 (13.6)	23 (10.4)	20 (8.4)	0.7 (0.5-0.99)	0.047

Not significant

❖ HCC vs. LC

❖ HCC vs. LC+CHB

❖ LC vs. CHB

❖ No significant association of *STAT4* variant rs7574865 with clinical parameters

❖ Variant rs9275319 in *HLA-DQ* is not in Hardy-Weinberg disequilibrium in Vietnamese population



Summary and Conclusion

- ❖ **HBV genotype and genotype mixtures are associated with altered pathogenesis and clinical outcome of HBV infection.**
 - ❖ **Atypical nuclear/perinuclear localization of HBx mutants might be responsible for an enhanced activation of STAT3, inhibition of STAT1 and silencing of SOCS1/SOCS3 expression.**
 - ❖ **Polymorphisms in host immune genes (*INF- α* , *INF- α* receptor, *FCN2*, *MICA*, *CISH*, *STAT4*) are associated with clinical outcomes of HBV infection in Vietnamese population.**
 - ❖ **Ficolin-2, MICA play an important role in immune modulation of HBV infection, liver disease progression and tumor surveillance.**
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Acknowledgments



Thanks for your attention!

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