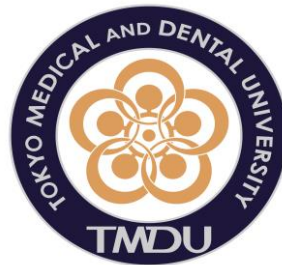


Glycosylation regulates CD38 assembly on the cell surface

GLYCOBIOLOGY 2015,
August 12, 2015, Philadelphia

Miki Hara-Yokoyama

Tokyo Medical and Dental University (TMDU), Japan

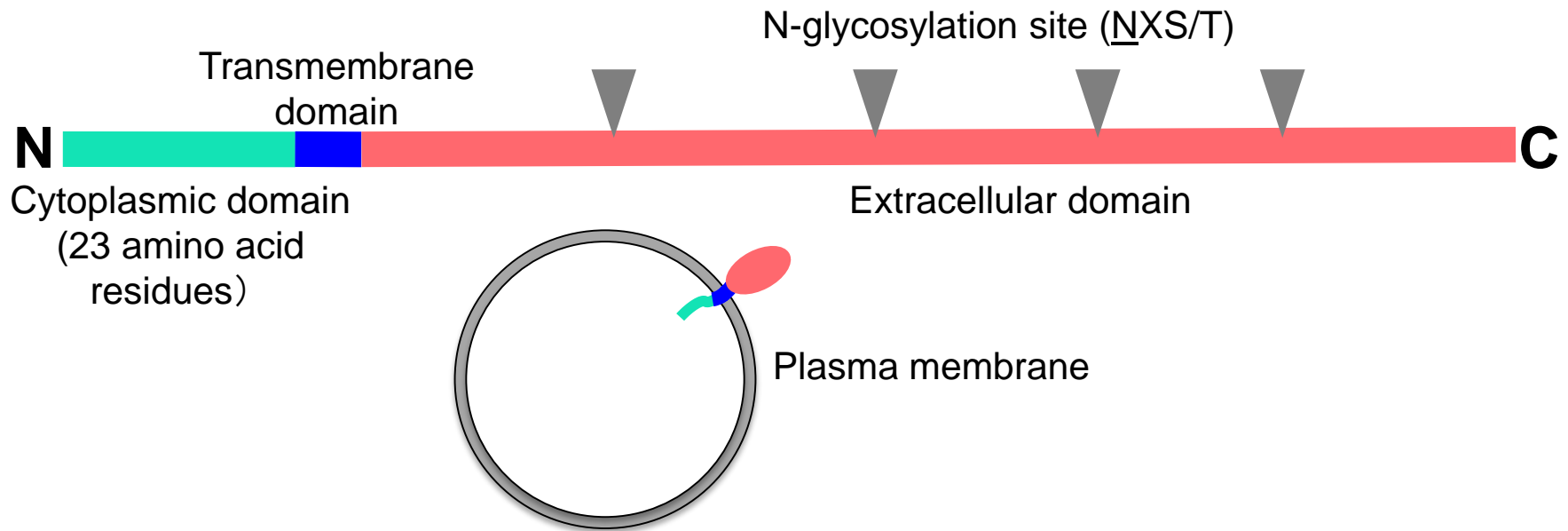


Outline

- Introduction of CD38
- Aim of this study
- The structural analysis of assembly
 - 1) the extracellular domain of CD38 in solution
 - 2) CD38 on the cell surface
- The functional significance of the assembly
- The regulation by glycosylation

Introduction of CD38

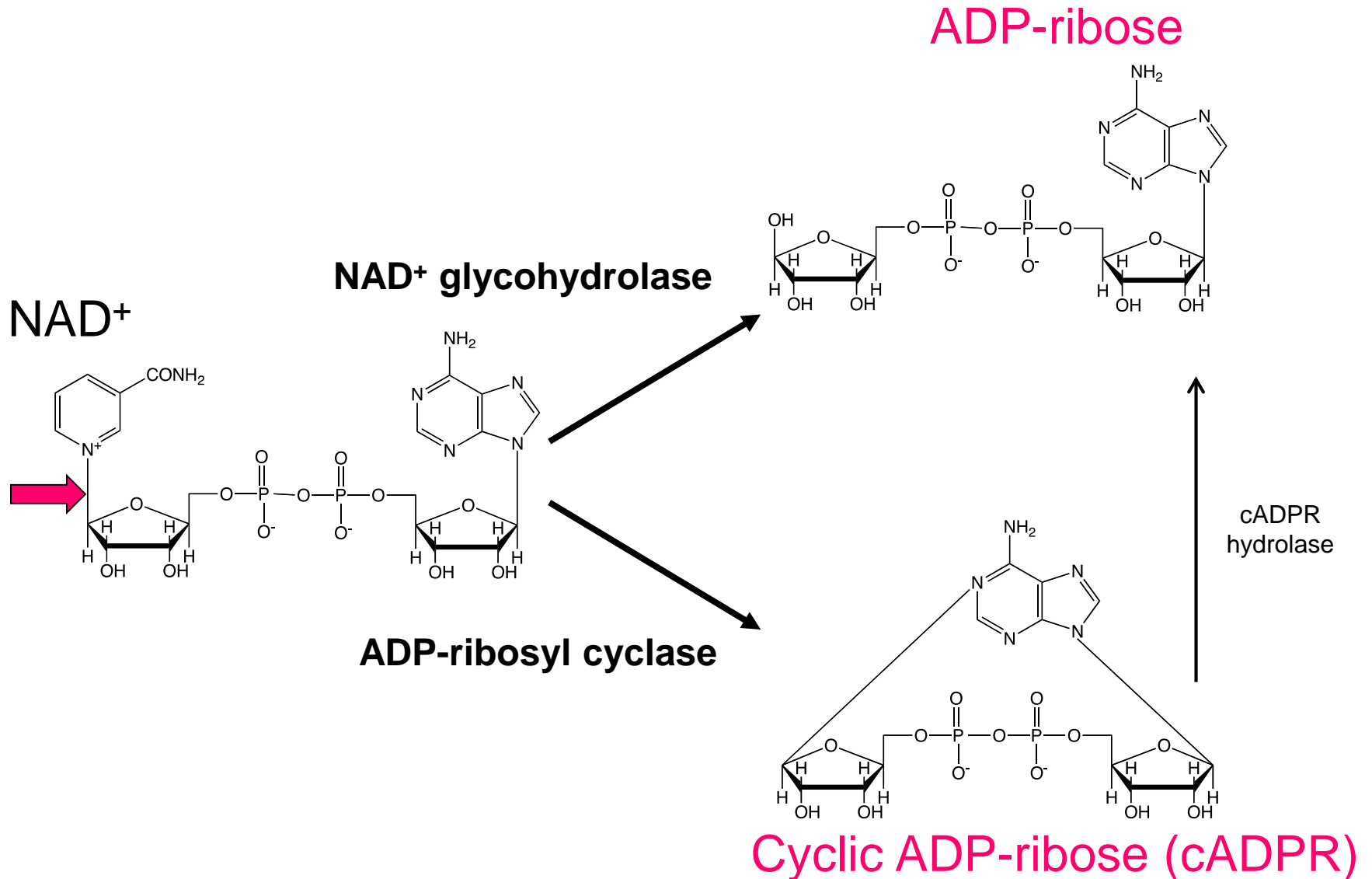
The leukocyte cell-surface antigen CD38 is a multifunctional protein.



Ectoenzyme

Raft-dependent signaling molecule

CD38 is the major NAD⁺ glycohydrolase in mammals.

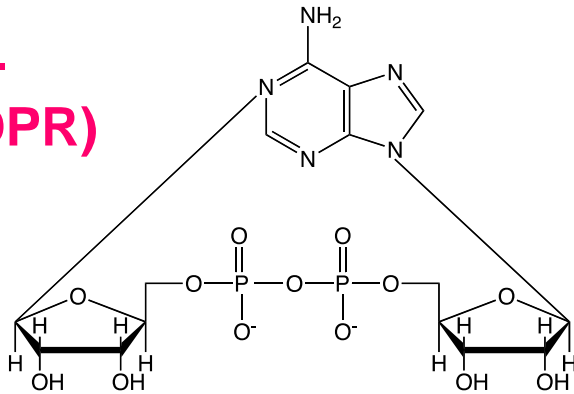


Cyclic ADP-ribose triggers intracellular calcium mobilization in an IP₃-independent manner.

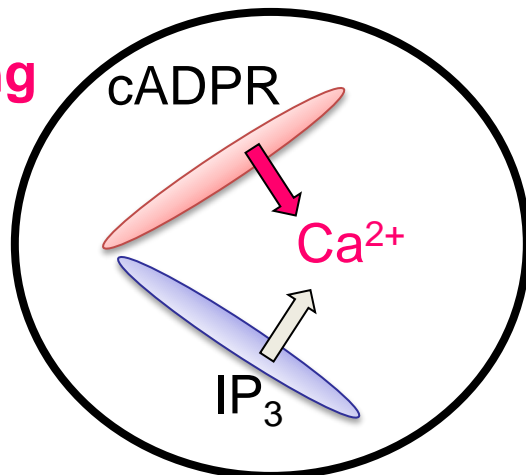
Ectoenzyme

Trafficking of neutrophils and dendritic cells
Secretion of insulin and oxytocin

Cyclic ADP-ribose (cADPR)



Ca²⁺ mobilizing messenger



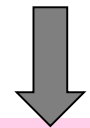
Aplysia



Aplysia ADP-ribosyl cyclase



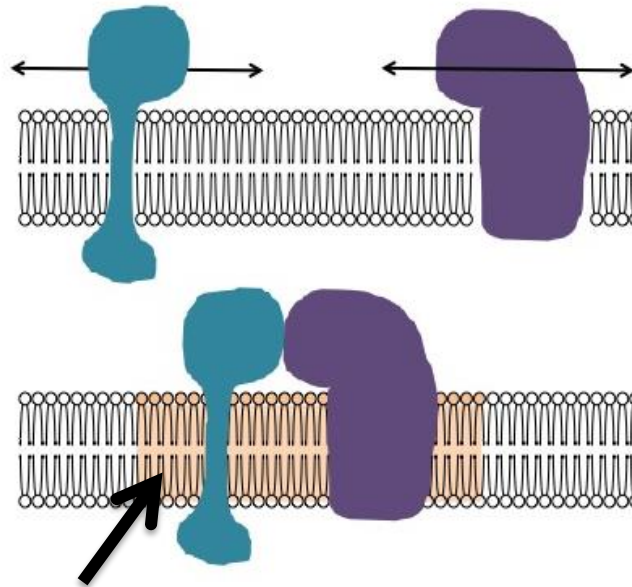
Human



CD38

Similar structure

CD38 associates with various supramolecular complexes within lipid rafts.

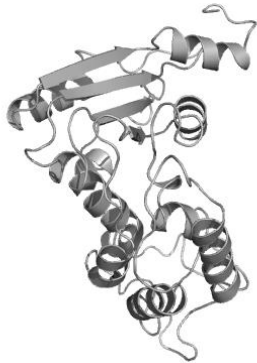


Cholesterol/sphingolipid-enriched membrane domain (lipid raft)

T cells	CD38/CD3/Lck/LAT
B cells	CD38/BCR/CD19/CD81
NK cells	CD38/CD16
Monocytes	CD38/MHC Class II/CD9
Dendritic cells	CD38/CD83/CD11b/CD81

Aim of this study

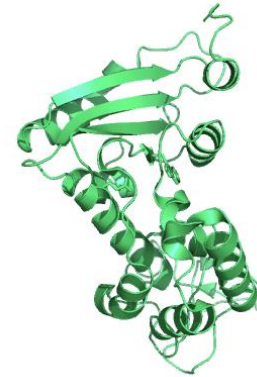
Aplysia ADP-ribosyl
cyclase
(cytosolic protein)
1LBE



BST1/CD157
(GPI-anchored
protein)
1ISF



CD38
(transmembrane
protein)
1YH3



Dimer

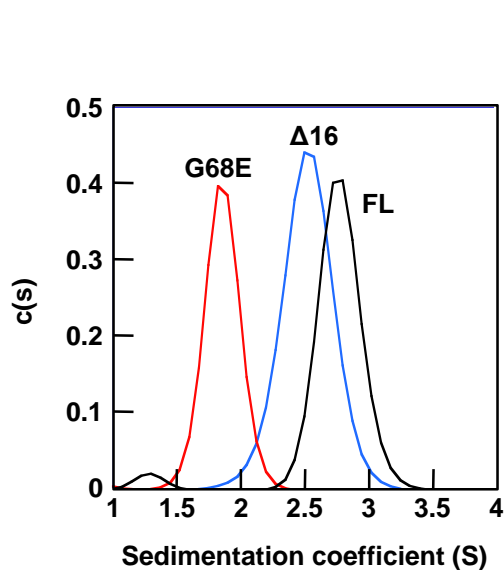
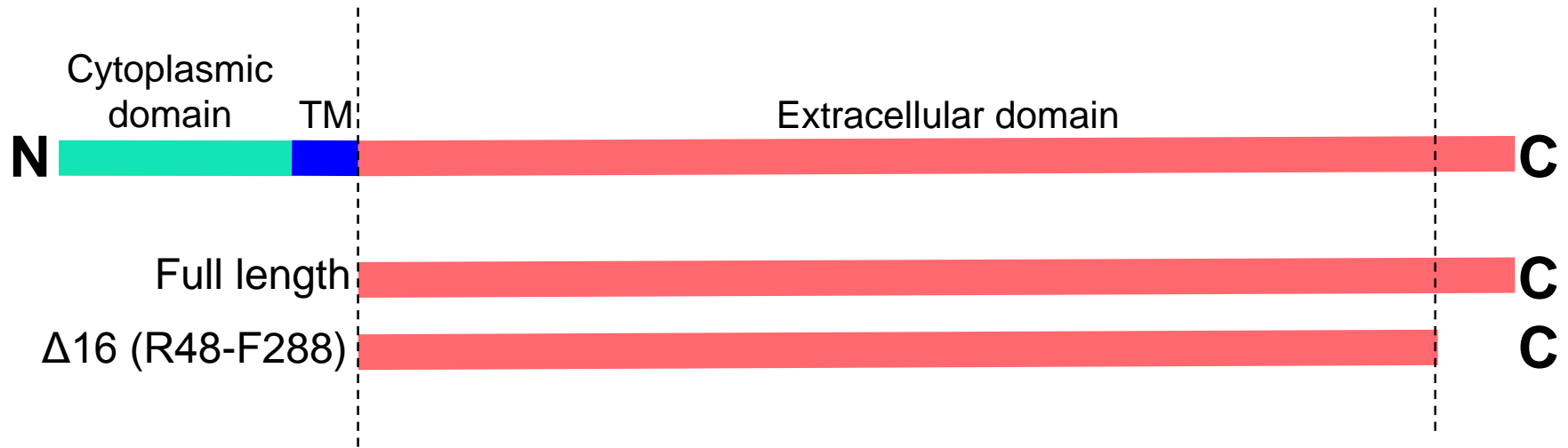
Structural basis &
functional significance

?

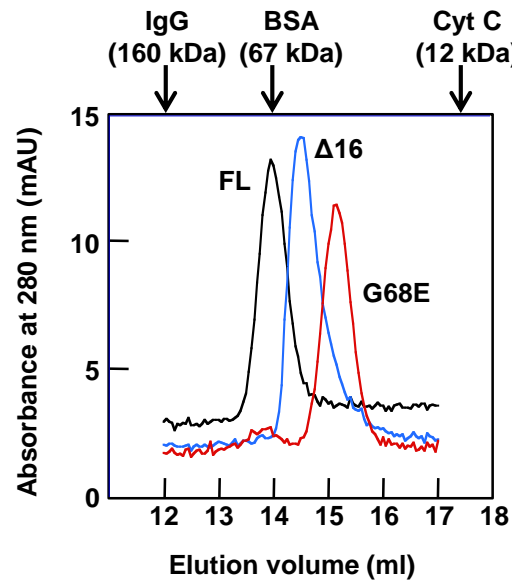
Tetramer

The structural analysis
(the extracellular domain of CD38 in
solution)

The full-length and the C-terminal-truncated extracellular domains of mouse CD38 exist as homodimers in solution.

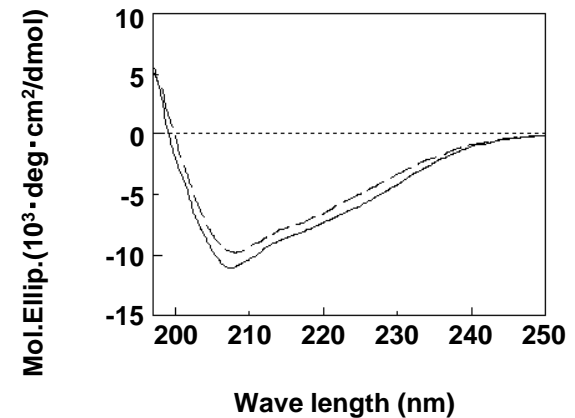


Analytical ultracentrifugation



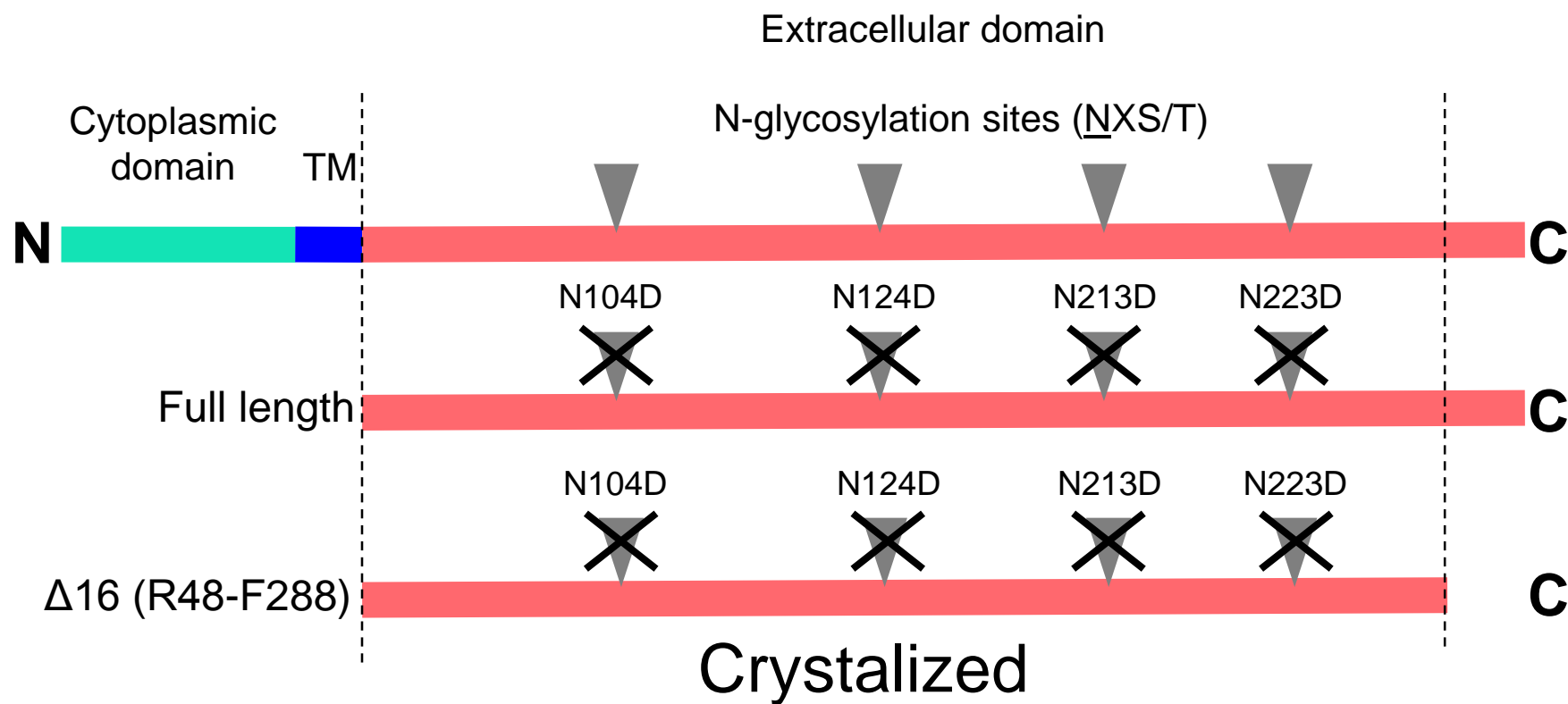
Size exclusion chromatography

The overall structure was not significantly altered by the truncation.



Circular dichroism spectra

Homophilic interfaces were found in the crystal packing of the C-terminal-truncated extracellular domain of CD38 !

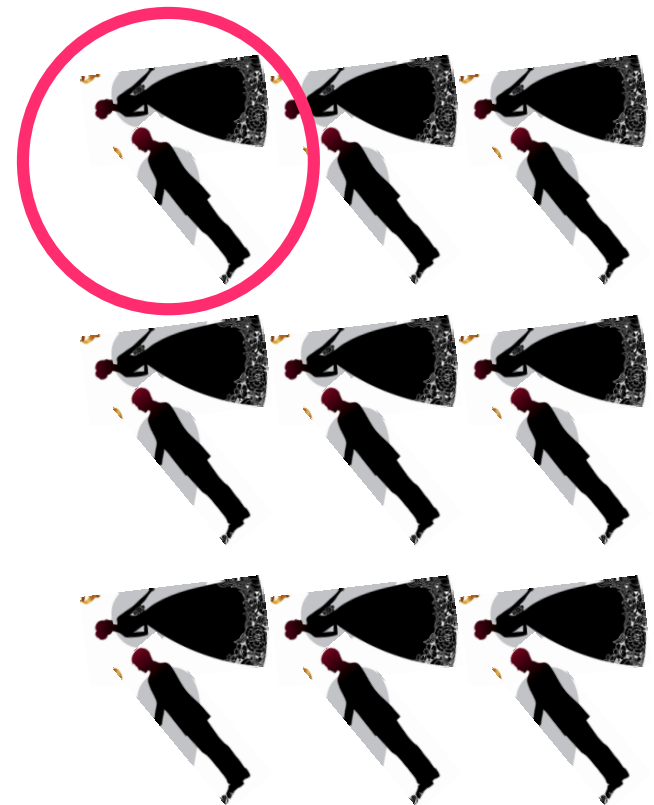


Concept of crystal packing

Information of
interfaces

Suggested

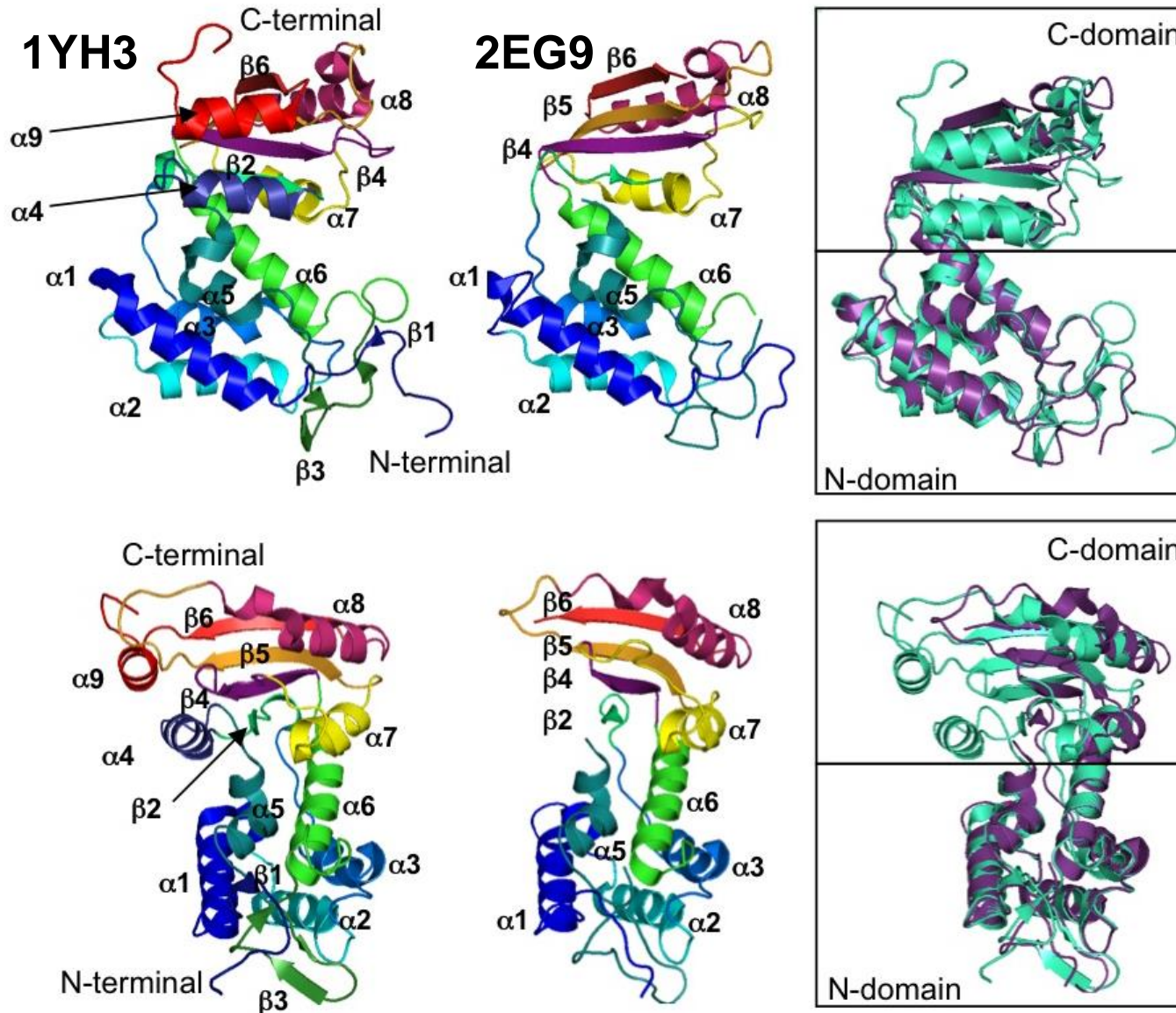
No information



This study (2EG9)

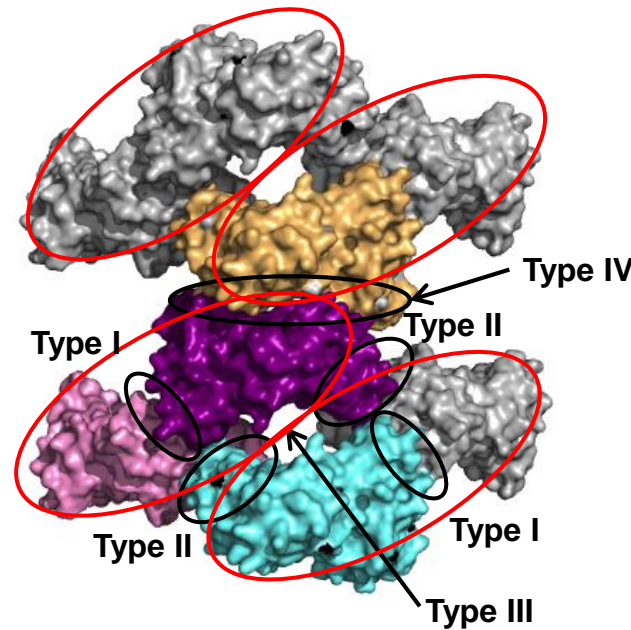
Previous study (1YH3)

The monomeric structure was not significantly altered by the truncation, except the loss of the $\alpha 9$ helix and the fluctuation of the $\alpha 4$ helix.

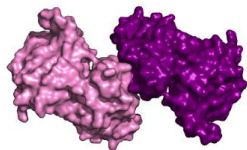
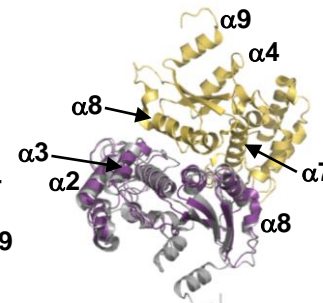
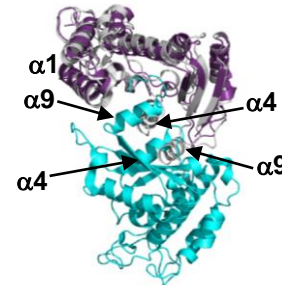
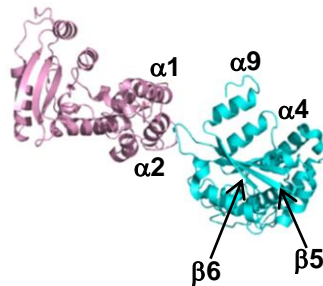
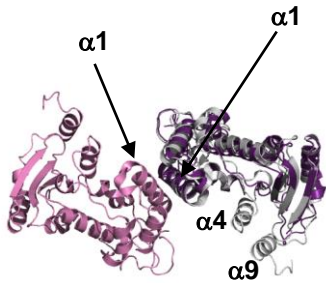


Four types of interfaces (I-IV) were found in the crystal packing of mCD38(R48-F288).

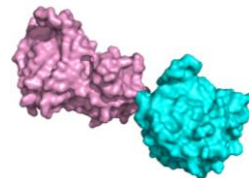
2EG9



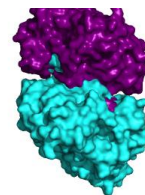
Do the interfaces exist also in solution?



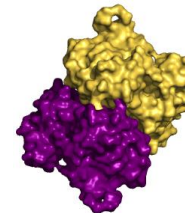
Type I



Type II



Type III

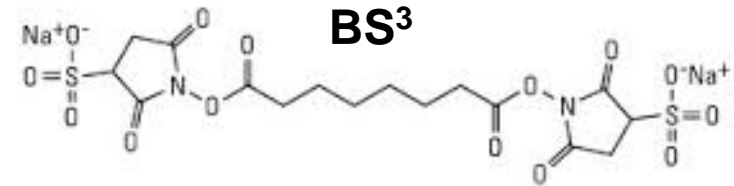
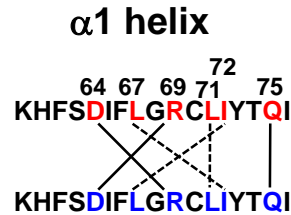
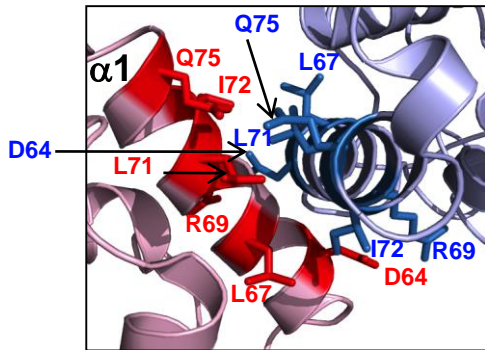


Type IV

Two molecules of the full-length extracellular domain are oriented according to those in 2EG9.

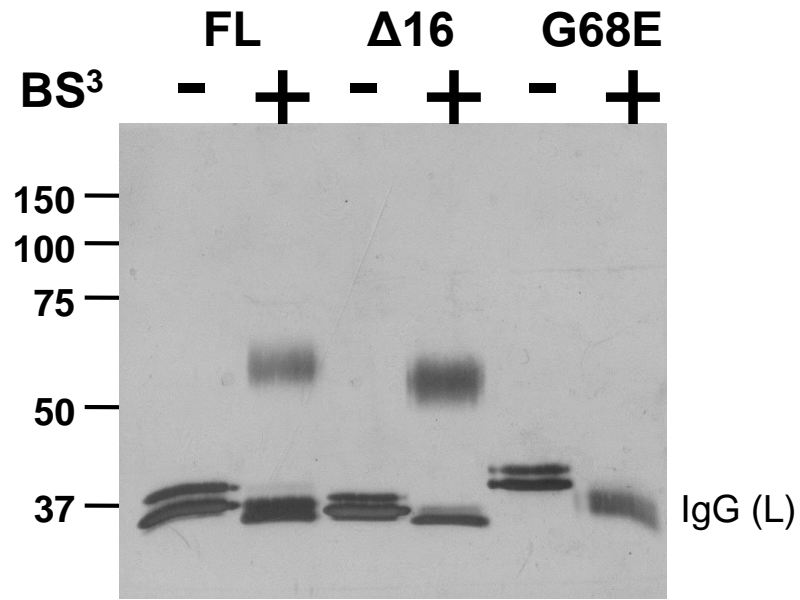
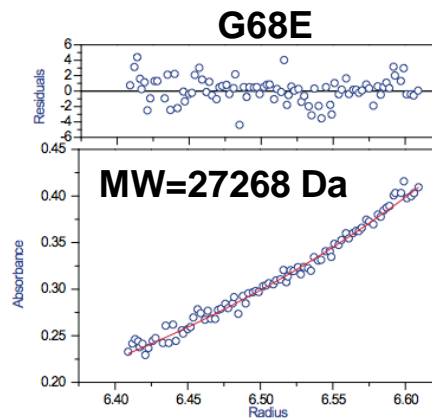
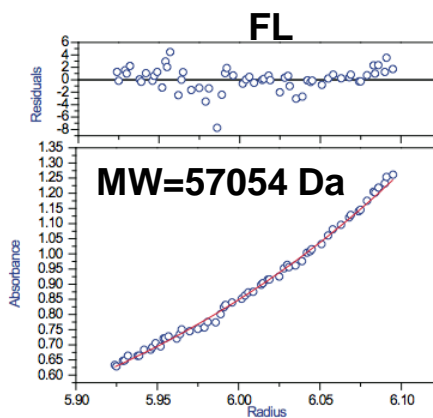
The results of the G68E mutation support the interaction between the $\alpha 1$ helices of the extracellular domain of CD38 in solution.

The G68E mutation should disrupt the type I interface.



BS3
Bis(sulfosuccinimidyl) suberate
MW 572.43
Spacer Arm 11.4 Å

Analytical ultracentrifugation
(sedimentation equilibrium)



The BS³-dependent crosslinking likely occurs via the type III interaction mode.

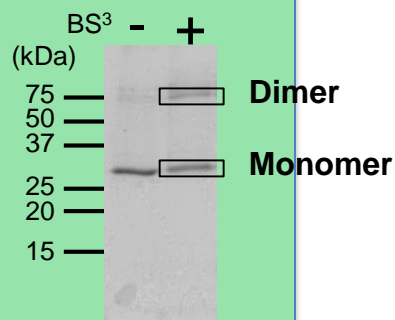
FLAG-CD38



crosslink



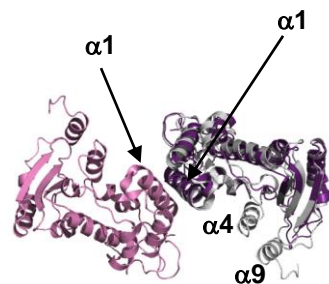
SDS-PAGE



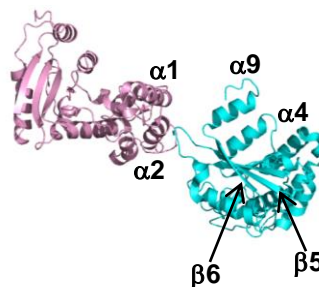
Trypsin digestion



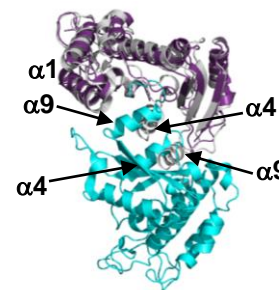
MS analysis



Type I



Type II



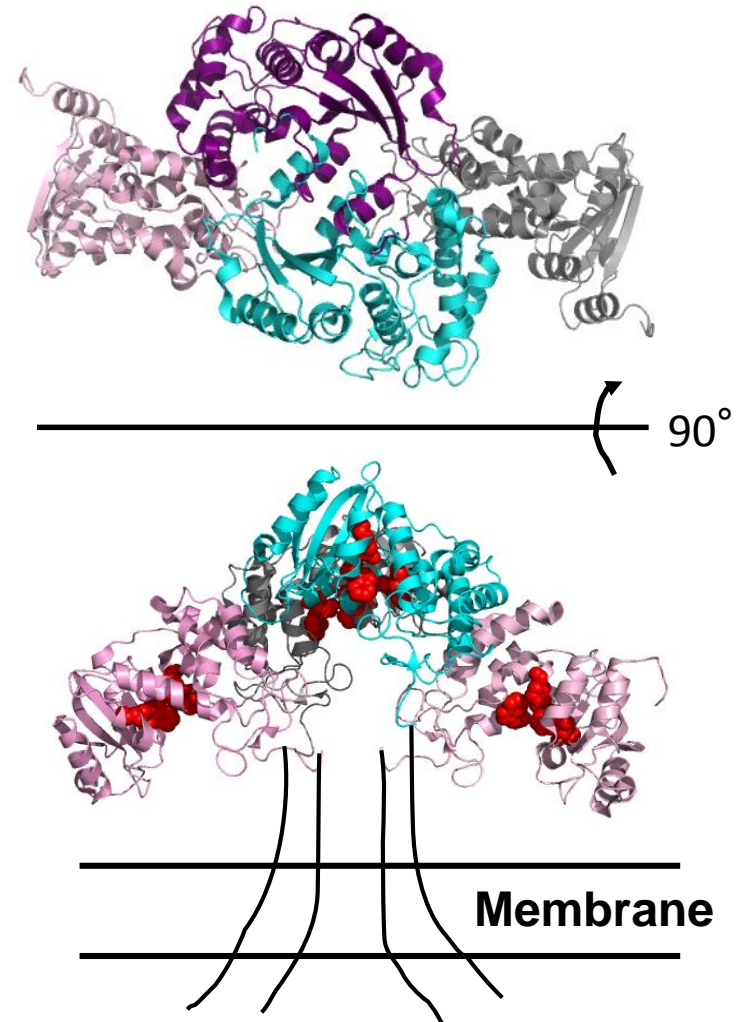
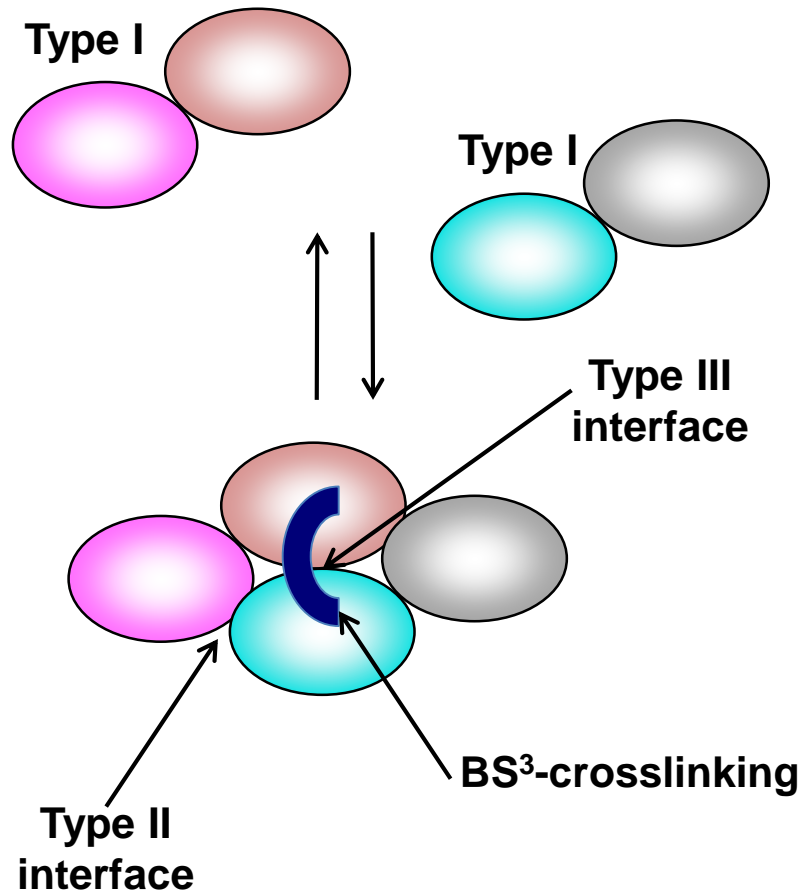
Type III

Lys145
 SKSKHLAHQYTWIQGKIMFTLED
 $m/z = 1481.79$
 (peak 6)

FLAG-tag
 RAMDYKDDDDKLRSLLVWTGEPTTKHFSDIFLGRCLIIYQ
 $m/z = 1331.75$ (peak 1)
 $m/z = 1091.60$ (peak 2)

Monomer	+	+
Dimer	+	-

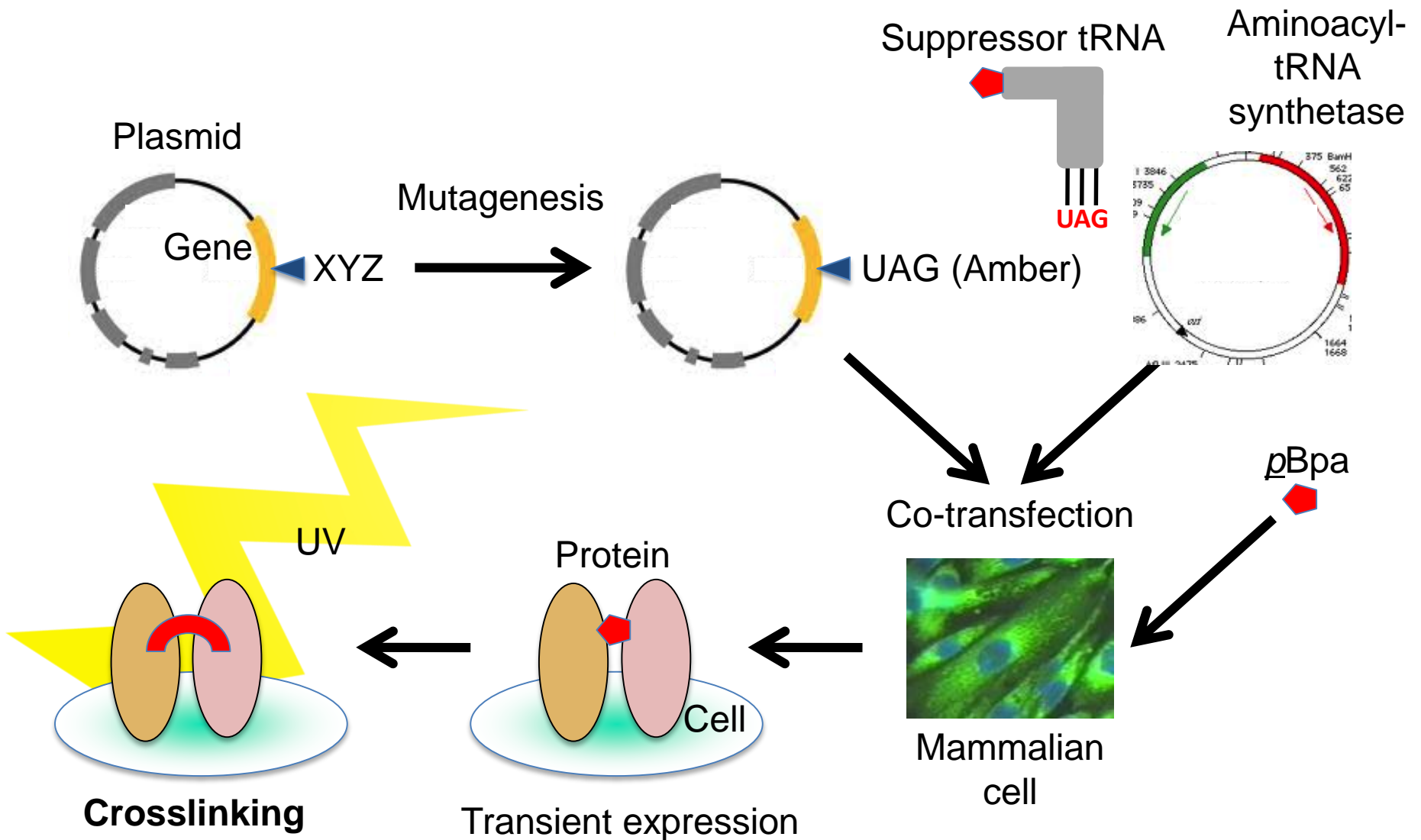
The dimer via the type I interaction mode exist in equilibrium to form a tetramer via the type II/III interaction mode, which is compatible with membrane association.



The structural analysis (CD38 on the cell surface)

Are the interfaces present in CD38 on the cell surface?

Site-specific crosslinking on the cell surface with an expanded genetic code.



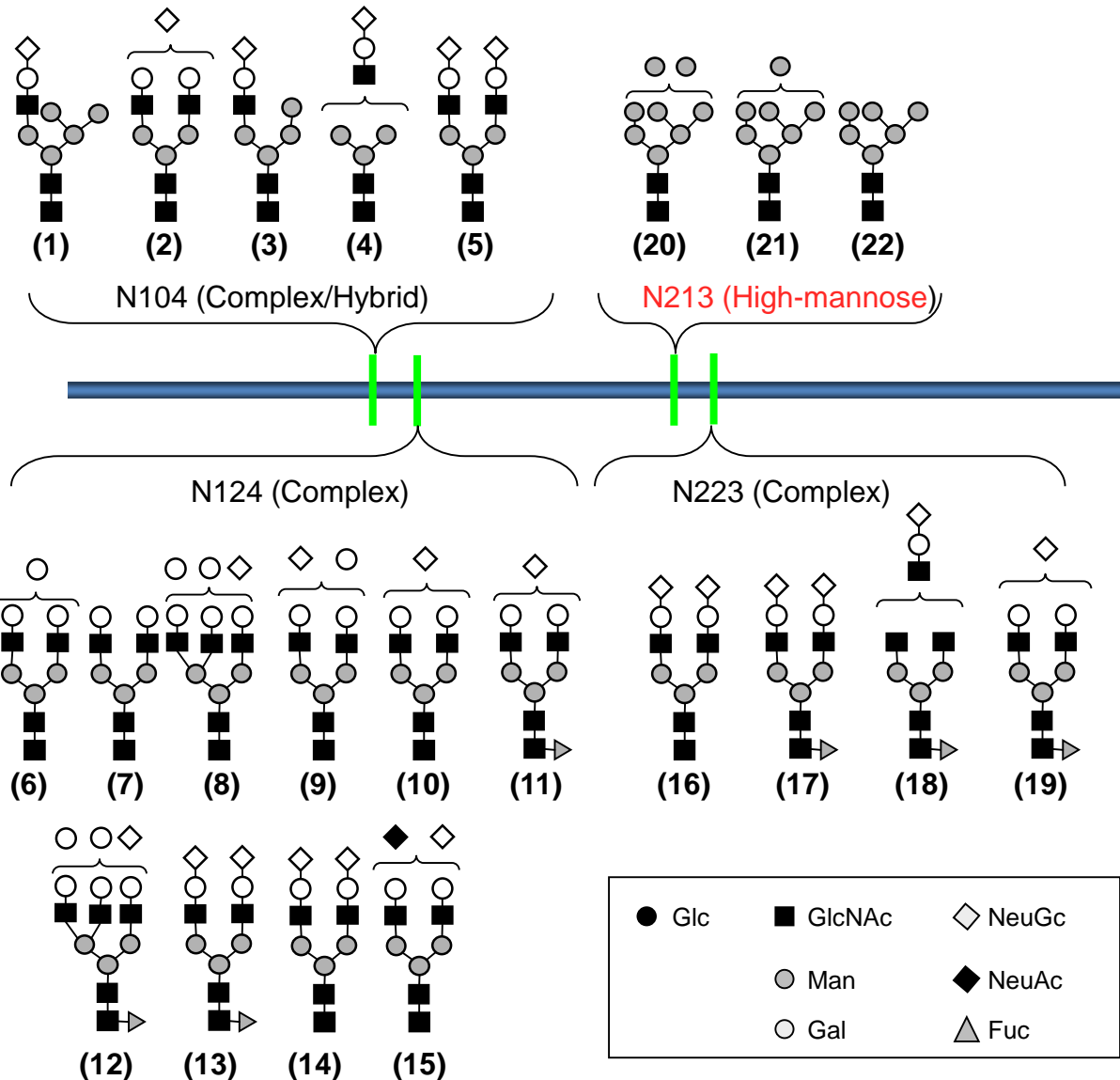
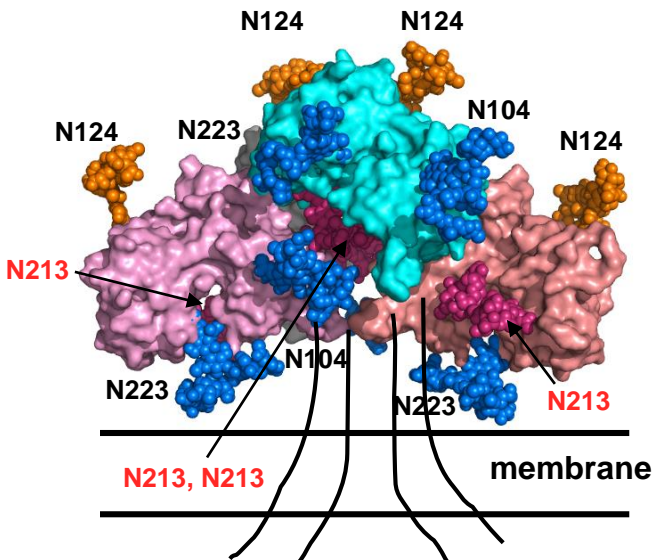
Only the oligosaccharides attached to the N213 residue remained as the high-mannose-type.

A20 cells expressing
FLAGx3-CD38

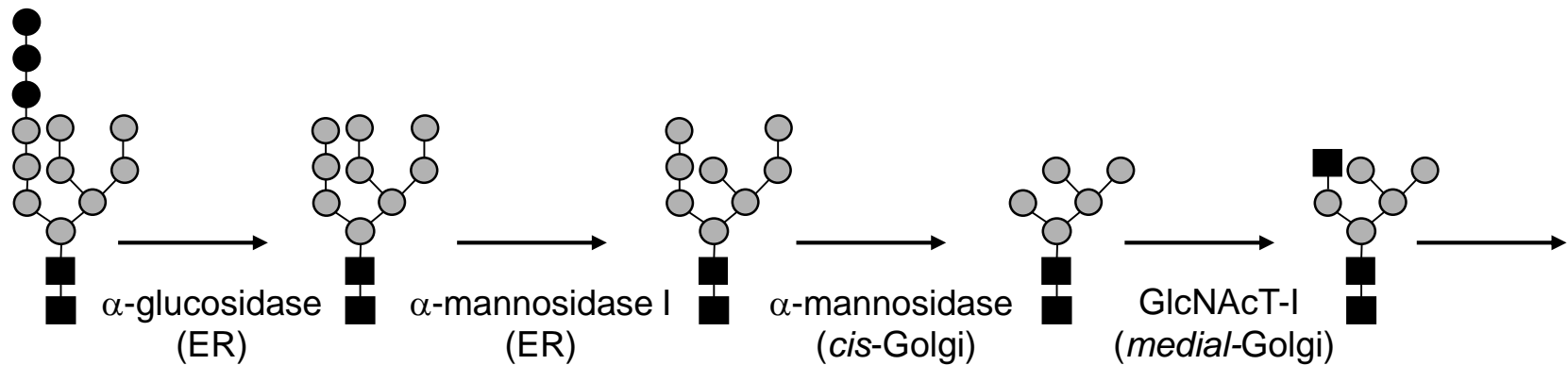
↓
IP: FLAG

↓
Trypsin digestion

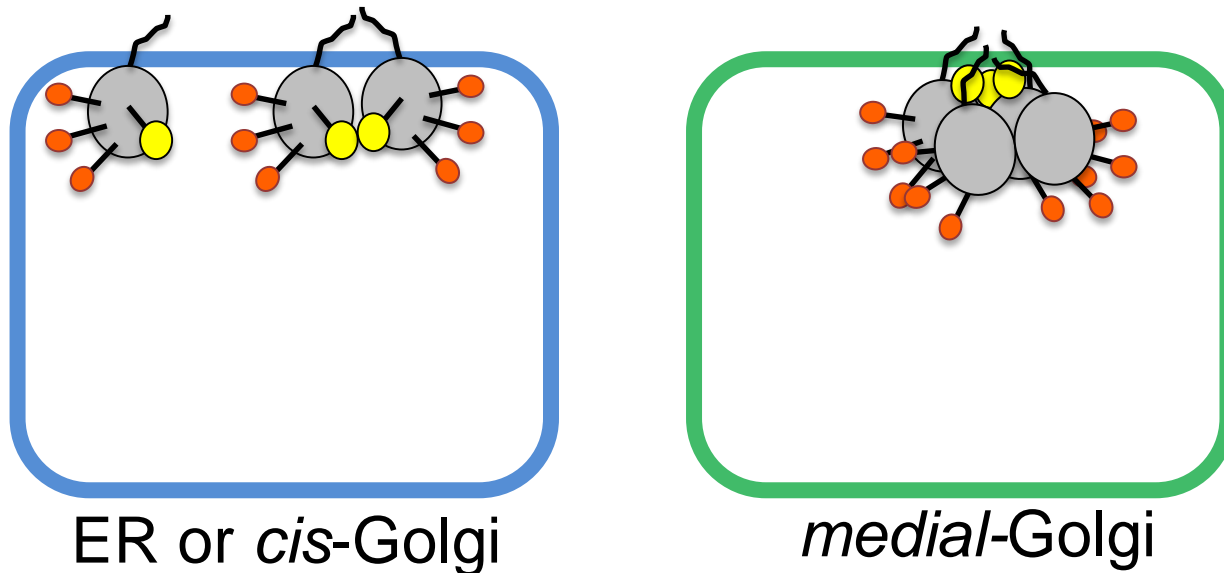
↓
MS analysis



The processing of the N-glycan of CD38 is compatible with tetramerization.



Within the tetramer, the processing enzymes are not accessible to the N213 residues.



The functional significance of tetramerization

Evaluation of the significance of the tetramerization of CD38 on the cell surface.

To impair the type I interface

G68E

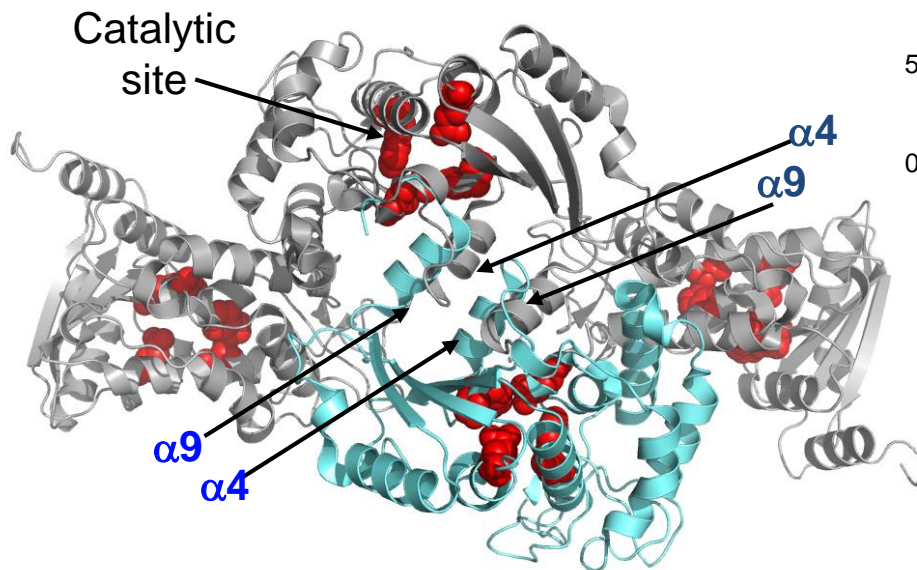
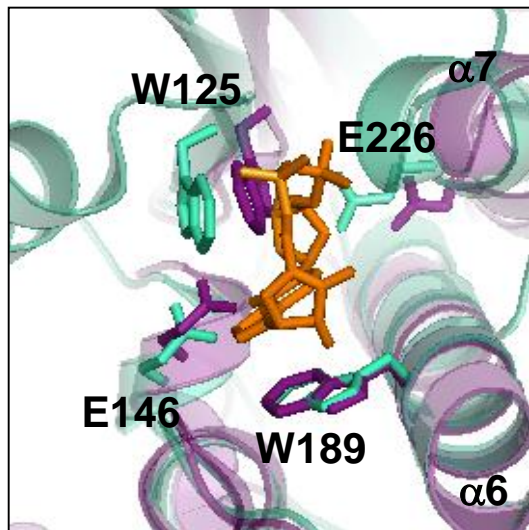
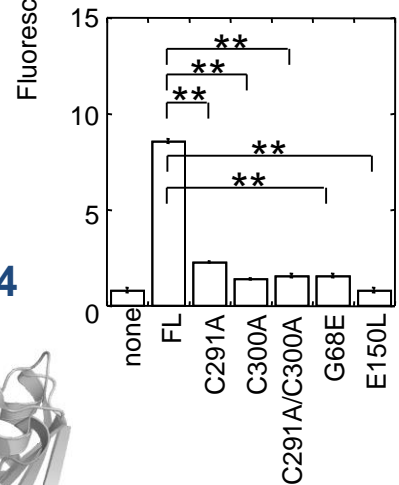
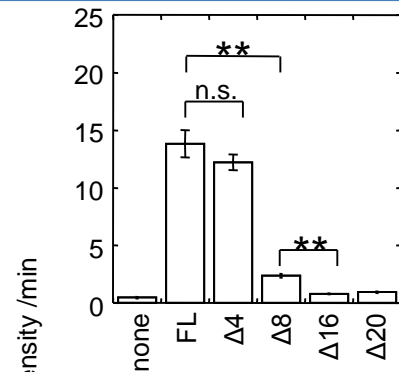
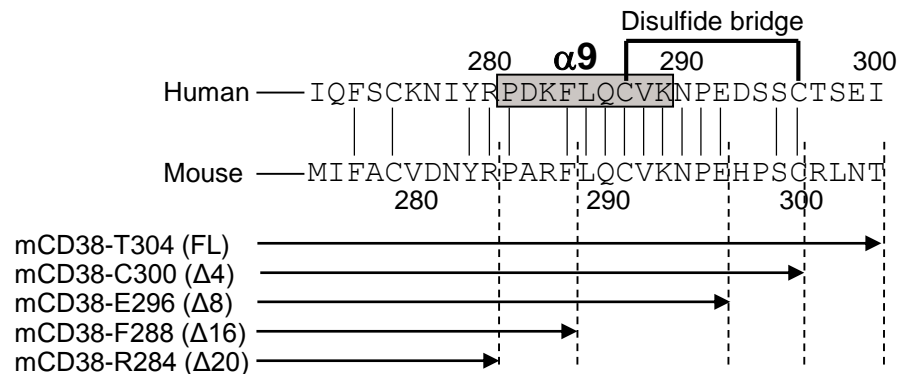
To affect the type III interface

C-terminal deletion

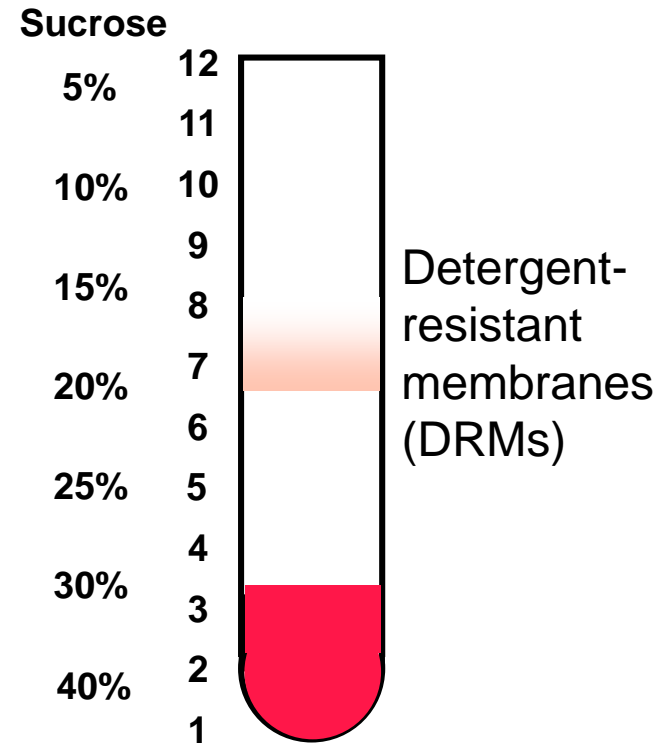
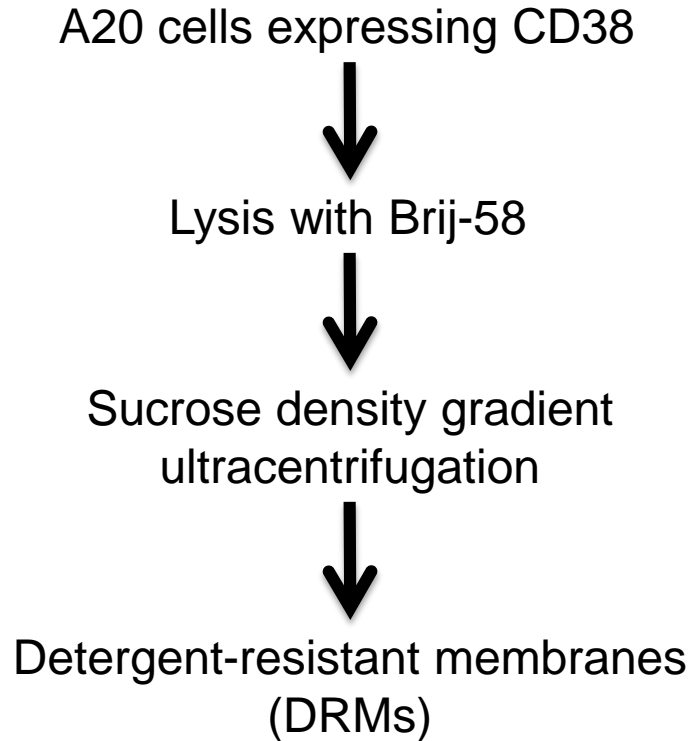
C291A, C300A, C291A/C300A

Both the I and type III interfaces are crucial for the tetramerization on the cell surface.

The tetramer structure (both type I and II/III) is required for the catalytic activity of CD38 in A20 cells.



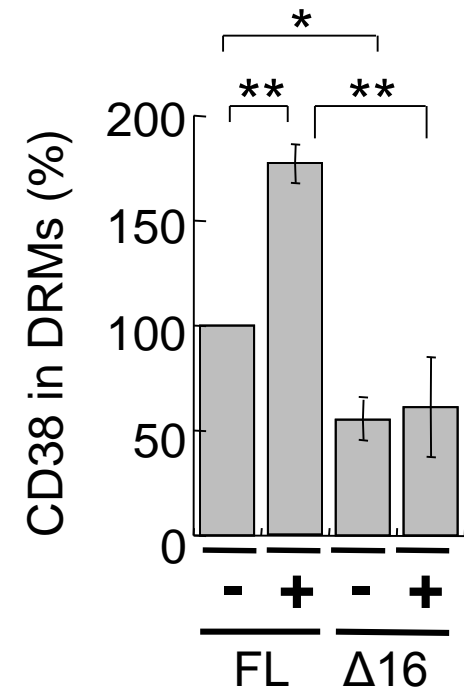
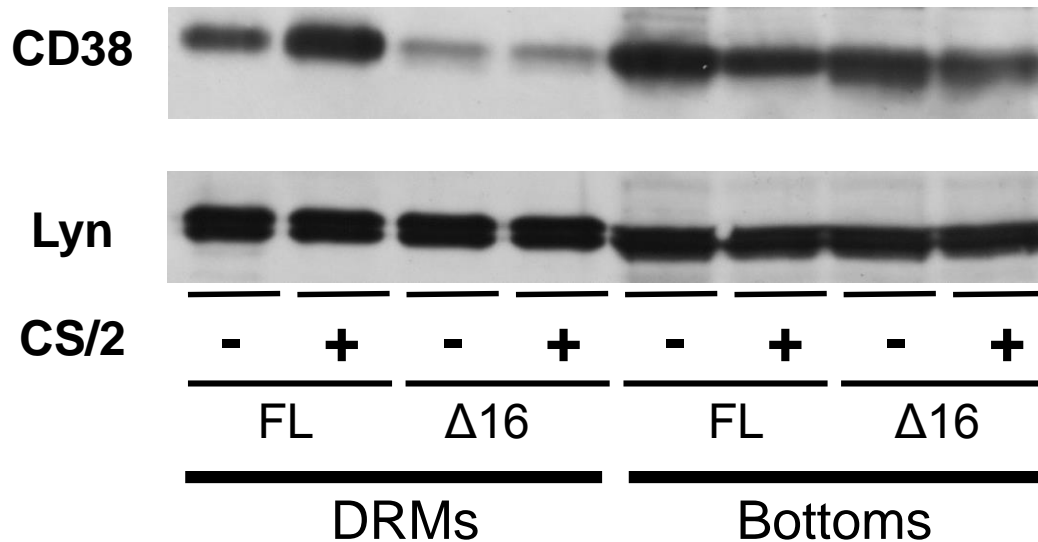
Preparation of detergent-resistant membranes (DRMs)



The tetramer structure is required for the association of CD38 with DRMs in A20 cells.

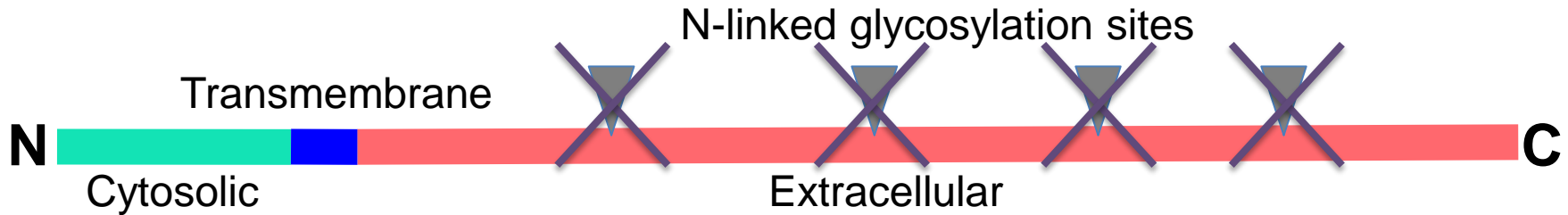
A20 cells expressing full-length CD38

A20 cells expressing truncated CD38



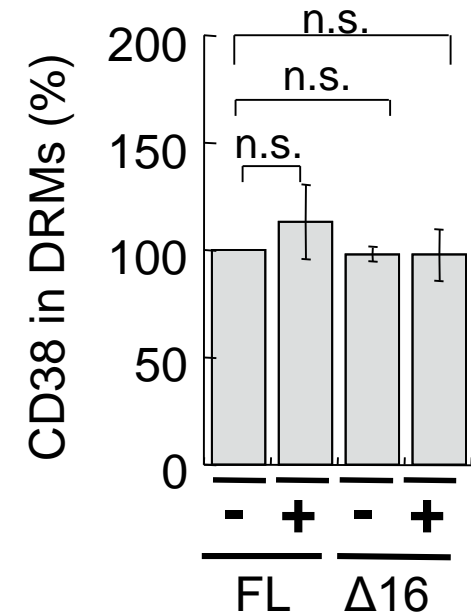
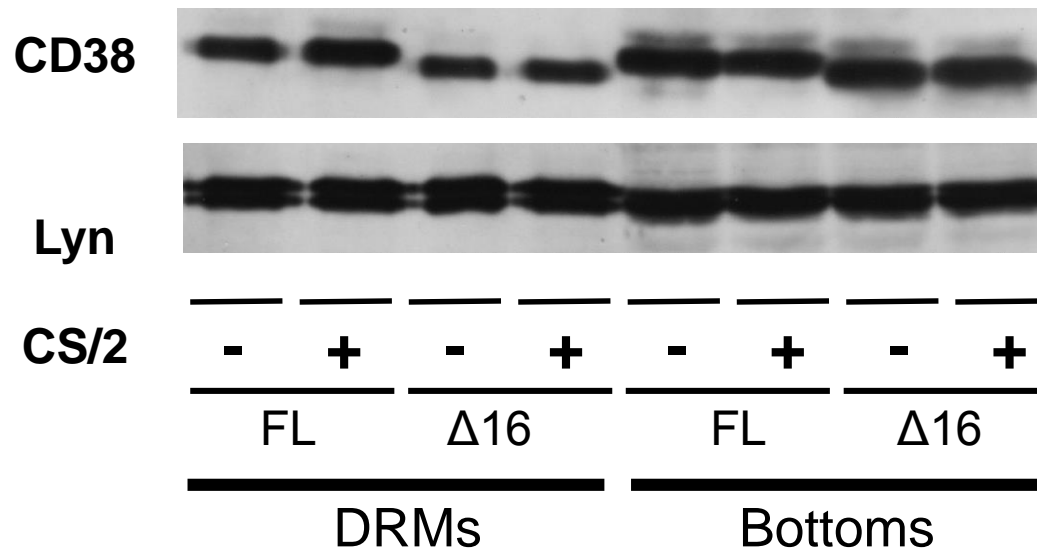
The effect of glycosylation

The C-terminal truncation did not alter the amount of nonglycosylated CD38 in DRMs.

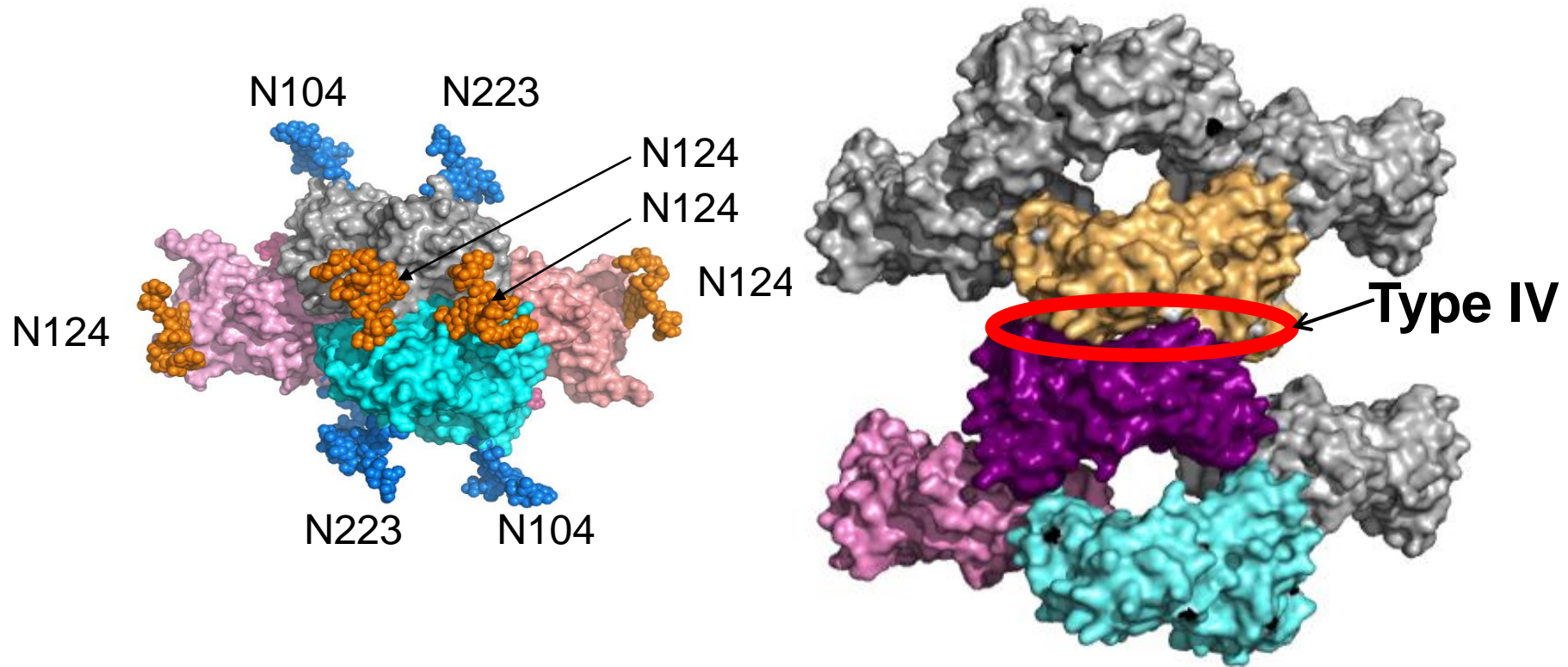


A20 cells expressing full-length deglycosylated CD38

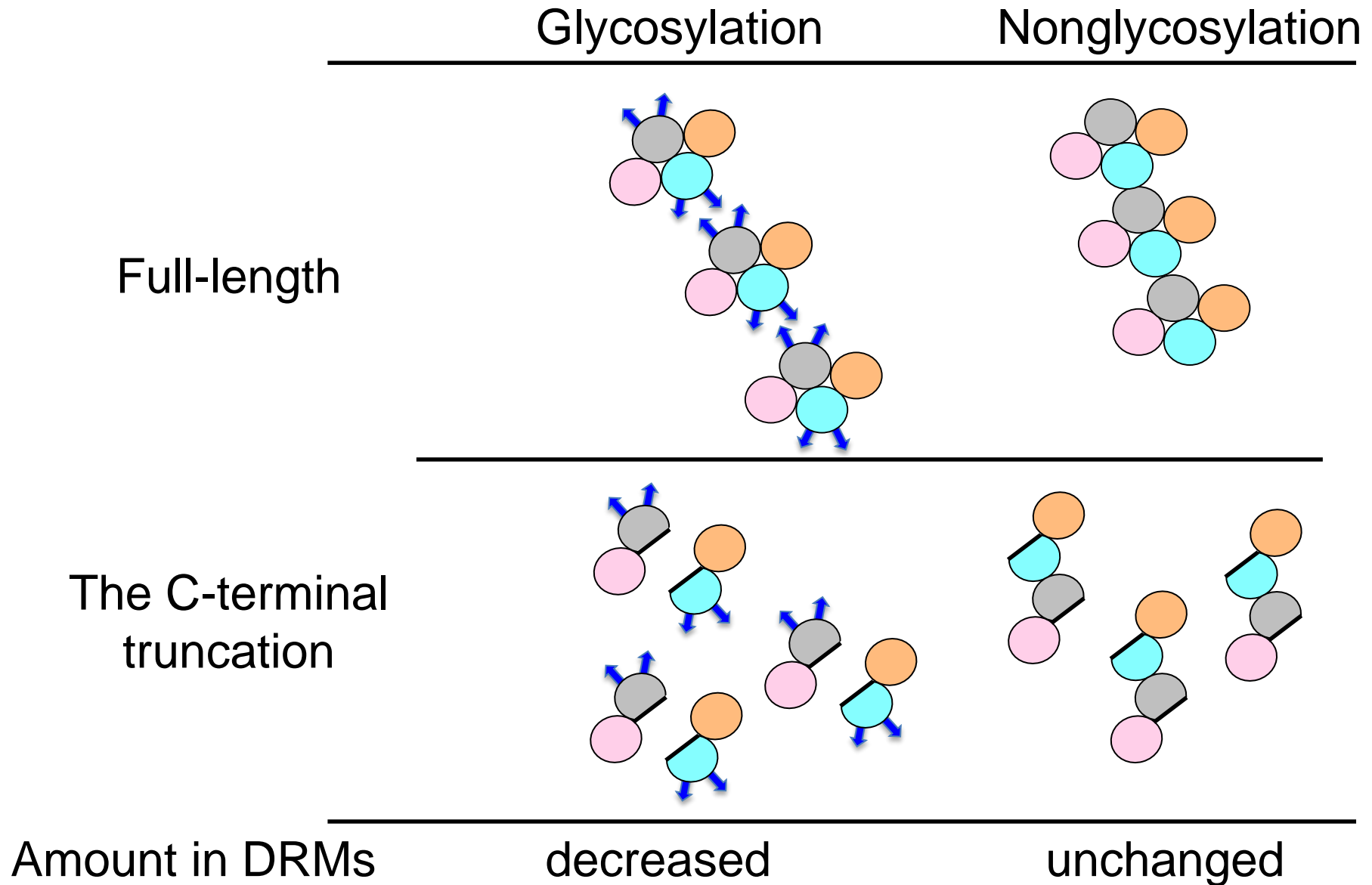
A20 cells expressing truncated deglycosylated CD38



The absence of the N-glycans attached to N104 and N223 enables the formation of the “type IV” interface in the case of cell-surface CD38.

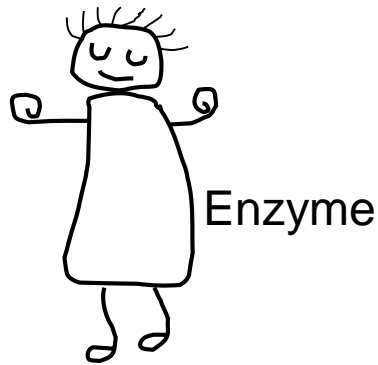


The N-glycans probably regulate the assembly of CD38 on the cell surface by inhibiting the “aggregating” type IV interface.

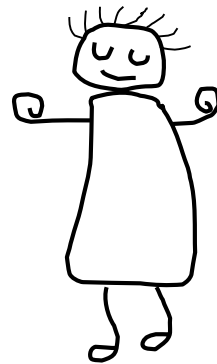


Summary

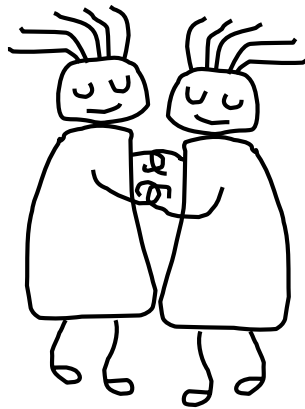
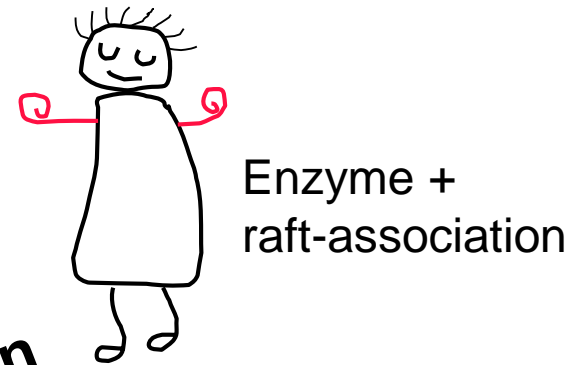
Aplysia ADP-ribosyl
cyclase
(cytosolic protein)
1LBE



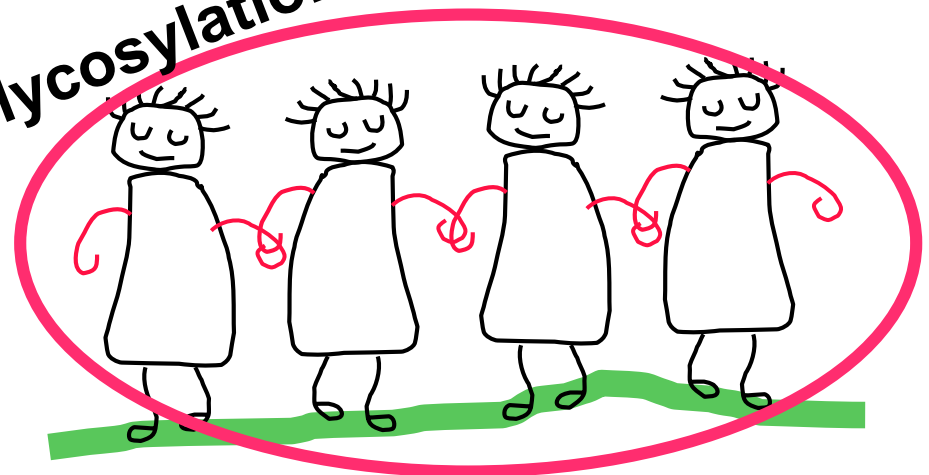
BST1/CD157
(GPI-anchored
protein)
1ISF



CD38
(transmembrane
protein)
1YH3



Glycosylation



Collaborators

Tokyo Medical and Dental University (TMDU)

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Katarzyna A. Podyma-Inoue
Takeshi Kasama
Hiroshi Takayanagi
Masaki Yanagishita

RIKEN (SSBC)

Mutsuko Kukimoto-Niino
Nobumasa Hino
Kensaku Sakamoto
Chiemi Mishima-Tsumagari
Yoko Kaitsu
Tomoko Matsumoto
Motoaki Wakiyama
Mikako Shirouzu
Yoshio Hirabayashi
Shigeyuki Yokoyama

National Institute of Health Sciences

Satsuki Itoh
Noritaka Hashimoto
Yoko Hiruta
Nana Kawasaki

Musashino University

Naoko Ustunomiya-Tate

University of Toyama

Kiyoshi Takatsu

RIKEN (BSI)

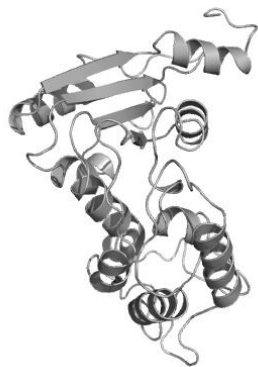
Yoshio Hirabayashi

University of Tokyo

Toshiaki Katada

We identified the interfaces contributing the tetramer formation.

***Aplysia* ADP-ribosyl
cyclase
(cytosolic protein)
1LBE**



**BST1/CD157
(GPI-anchored
protein)
1ISF**



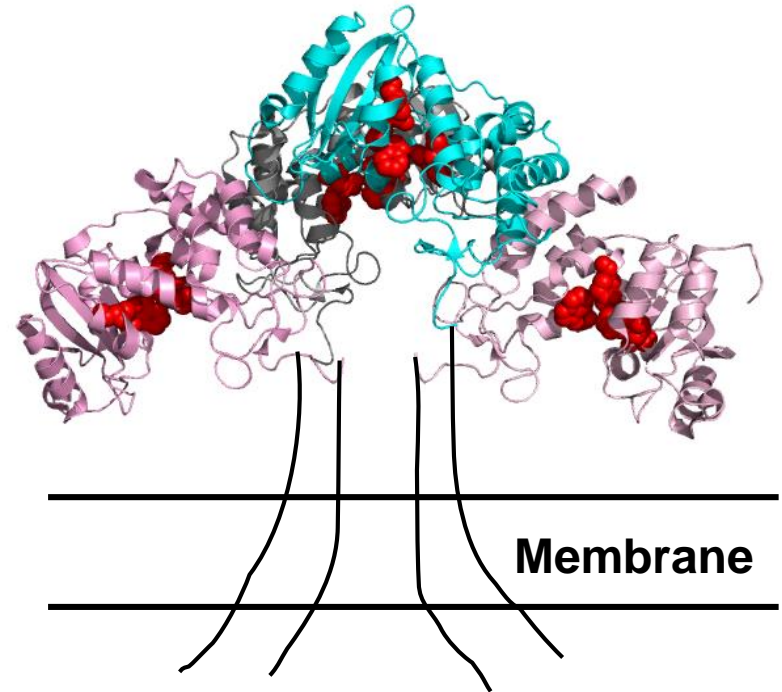
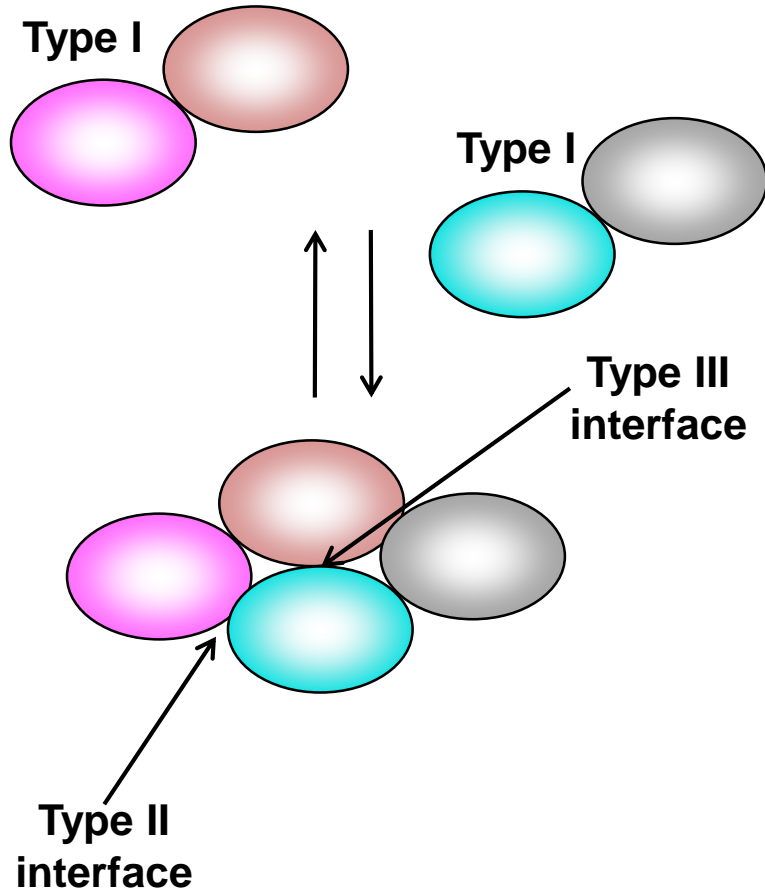
**CD38
(transmembrane
protein)
1YH3**



2EG9

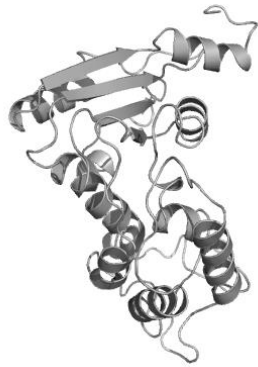
M.Hara-Yokoyama et al,
Structure 20: 1585-1595 (2012)

The dimerization of core dimers provides a structural basis for the previously reported tetramerization of CD38 on the cell-surface.



CD38の細胞膜上での四量体構造は機能と密接 に関与する

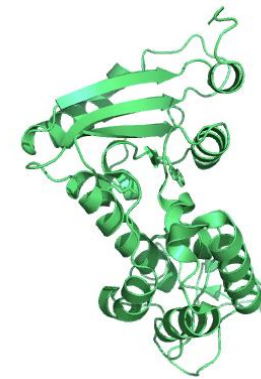
Aplysia ADP-ribosyl
cyclase
(cytosolic protein)
1LBE



BST1/CD157
(GPI-anchored
protein)
1ISF



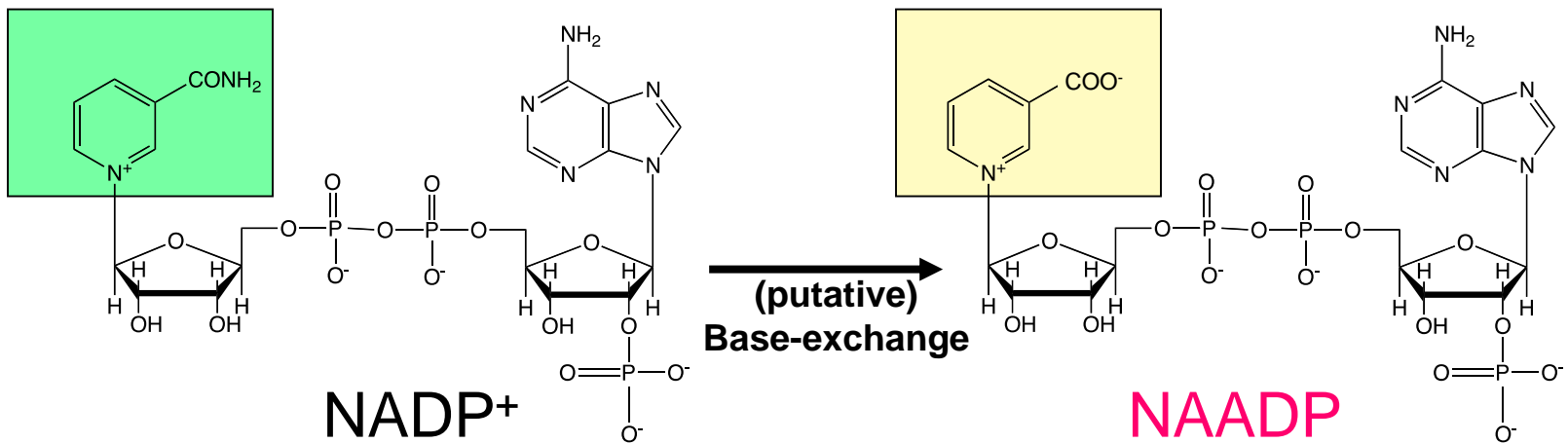
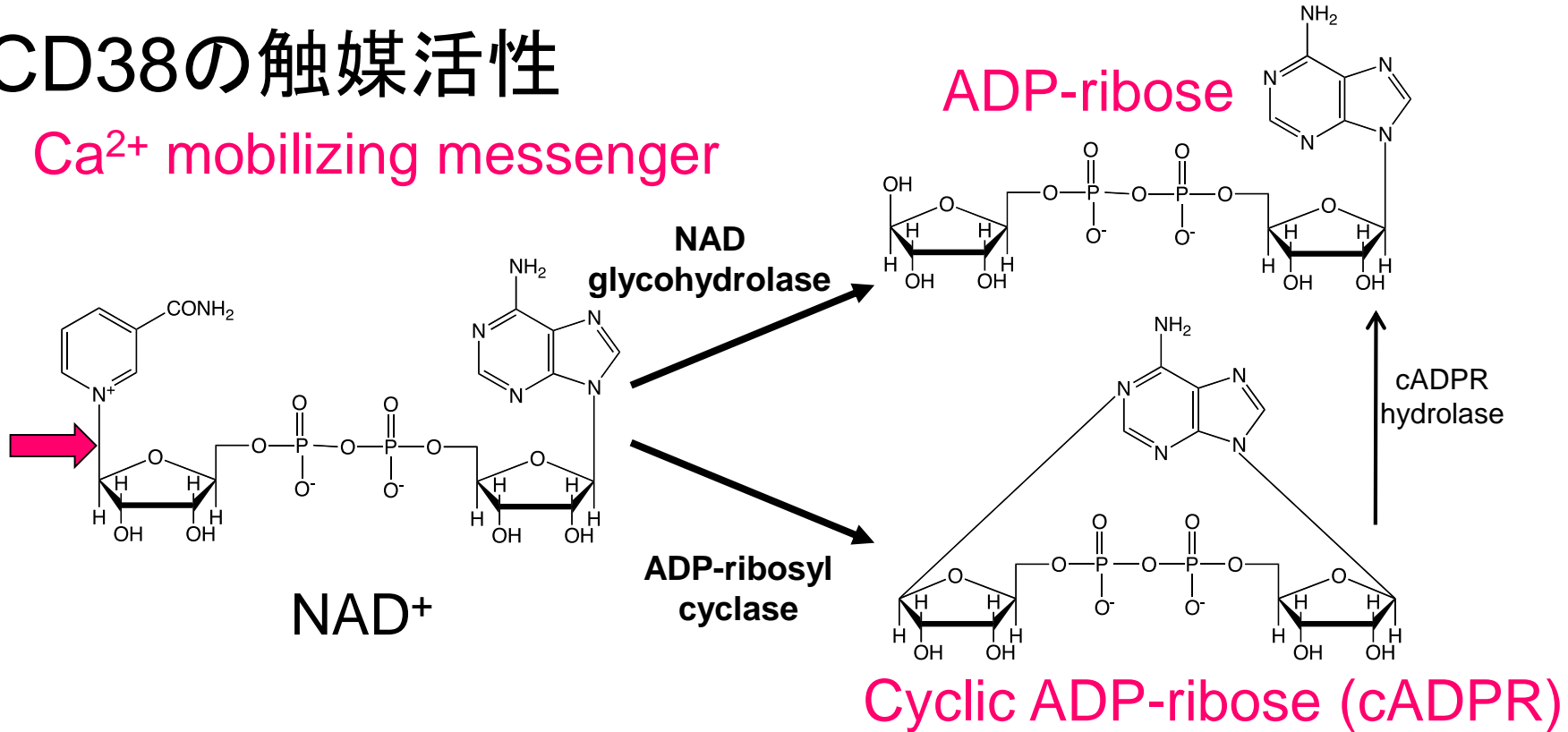
CD38
(transmembrane
protein)
1YH3



M.Hara-Yokoyama et al,
Structure 20: 1585-1595 (2012)

CD38の触媒活性

Ca²⁺ mobilizing messenger

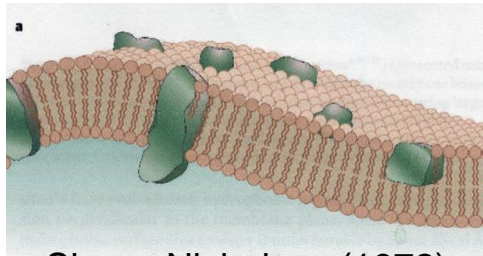


細胞内カルシウム動員活性をもつ

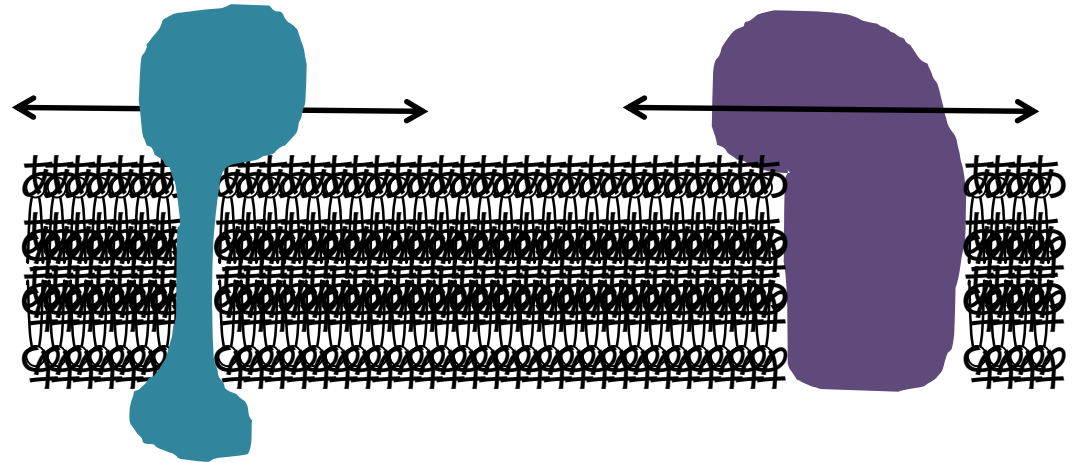
CD38は膜ドメインに存在する

膜ドメインとは

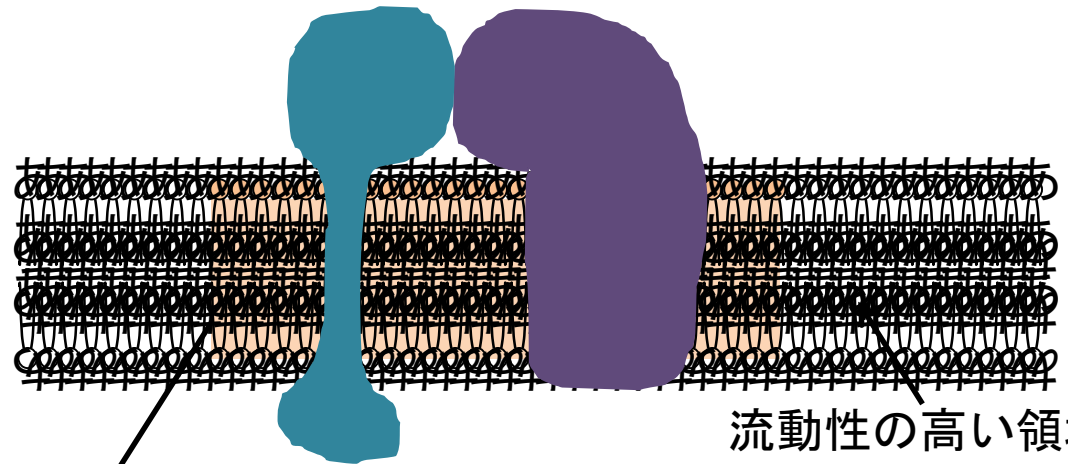
脂質二重層



Singer-Nicholson (1972)



生体膜には不均一性がある



流動性の高い領域

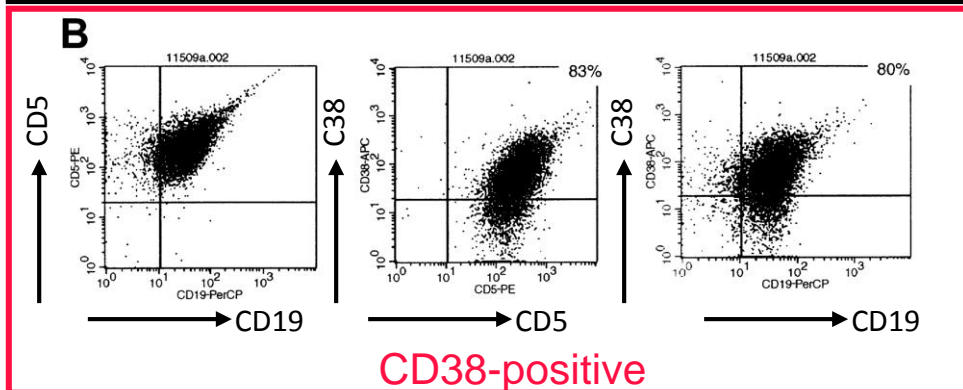
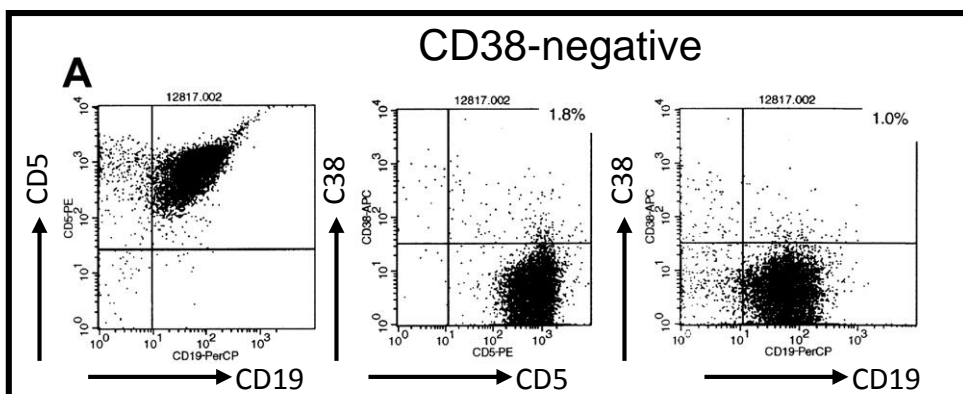
コレステロール・スフィンゴ脂質に富む流動性の低い領域
(膜ドメイン)

CD38 is recognized as a negative prognostic indicator in B-CLL patients.

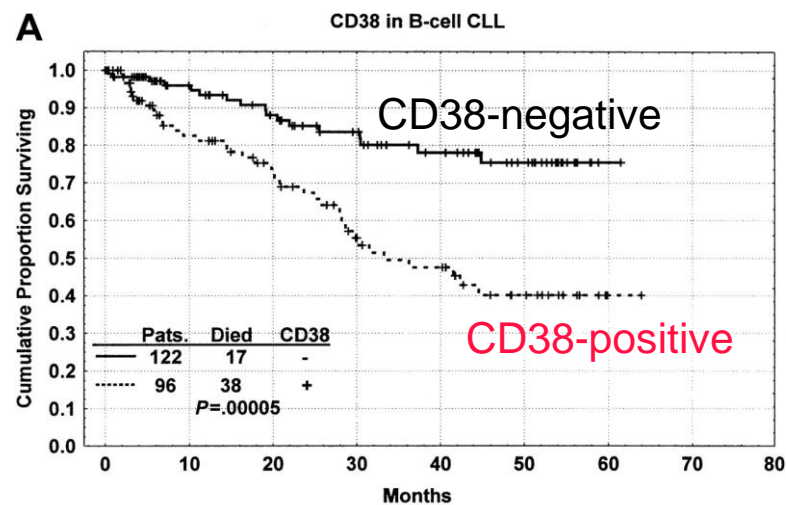
Peripheral blood



Flow cytometry (CD5/CD19/CD38)
B-CLL cells

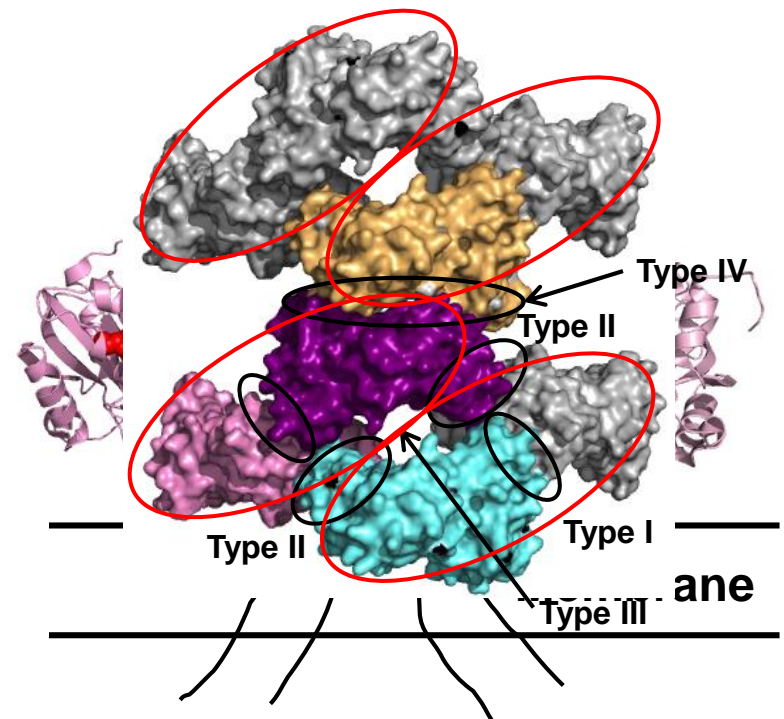
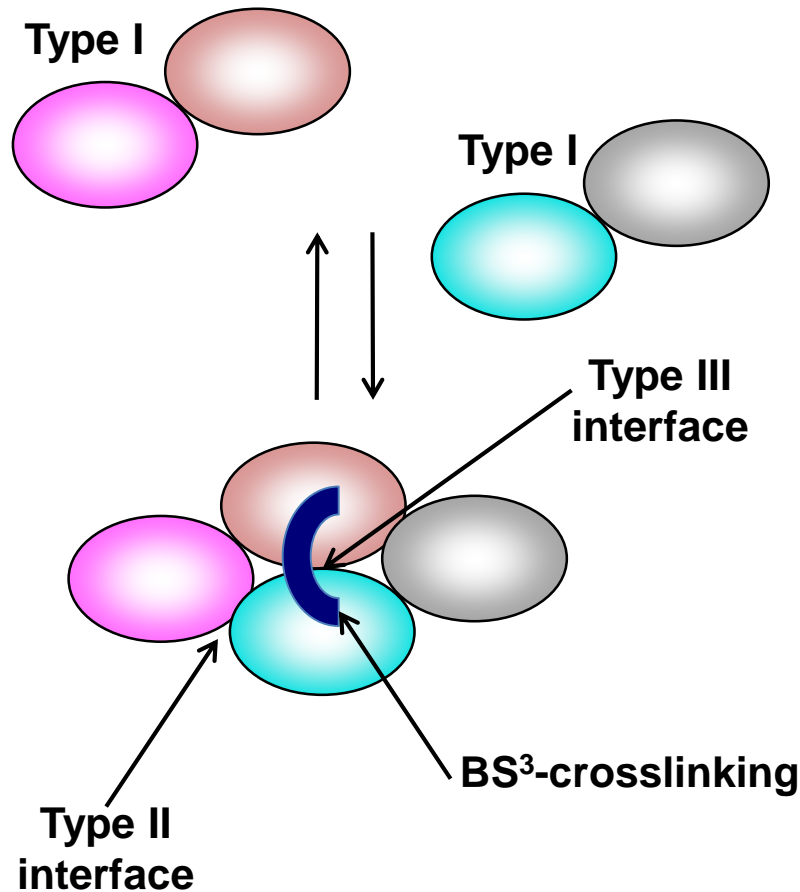


Kaplan-Meier survival curve



BLOOD 98:181-186 (2001)

The dimer via the type I interaction mode is further considered to exist in equilibrium to form a tetramer via the type II/III interaction mode, which is compatible with membrane association.



Daratumumab (anti-CD38 mAb)

Updated results of a key Phase 1/2 trial testing the potential new myeloma therapy daratumumab were released.

“Daratumumab continues to show substantial promise as potential new treatment for multiple myeloma (ASCO2015)”

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