



# Evaluation of non-instrumented nucleic acid amplification by loop-mediated isothermal amplification (NINA-LAMP) for the diagnosis of malaria in Northwest Ethiopia

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# Introduction



- Malaria is one of the deadly infectious disease worldwide especially in SSA countries like Ethiopia
- To minimize the burden of mortality and morbidity, early and accurate diagnosis of malaria is critically required

# Introd..

- Current routine diagnostic methods (Microscopy and Rapid diagnostic tests)
  - used to reduce the morbidity and mortality with malaria
  - less sensitive for detection of low level infections and asymptomatic cases

Bell D, WHO, 2010. Golassa L et al, 2013. Alemu et al , 2014.

# Introd...

- PCR assays are highly sensitive and specific method that can detect low level parasitemia
- PCR is not feasible in low resource settings (LRS)

(WHO, 2014, Cordray et al,2012 ; Noemi et al, 2000; Polley et all, 2010)







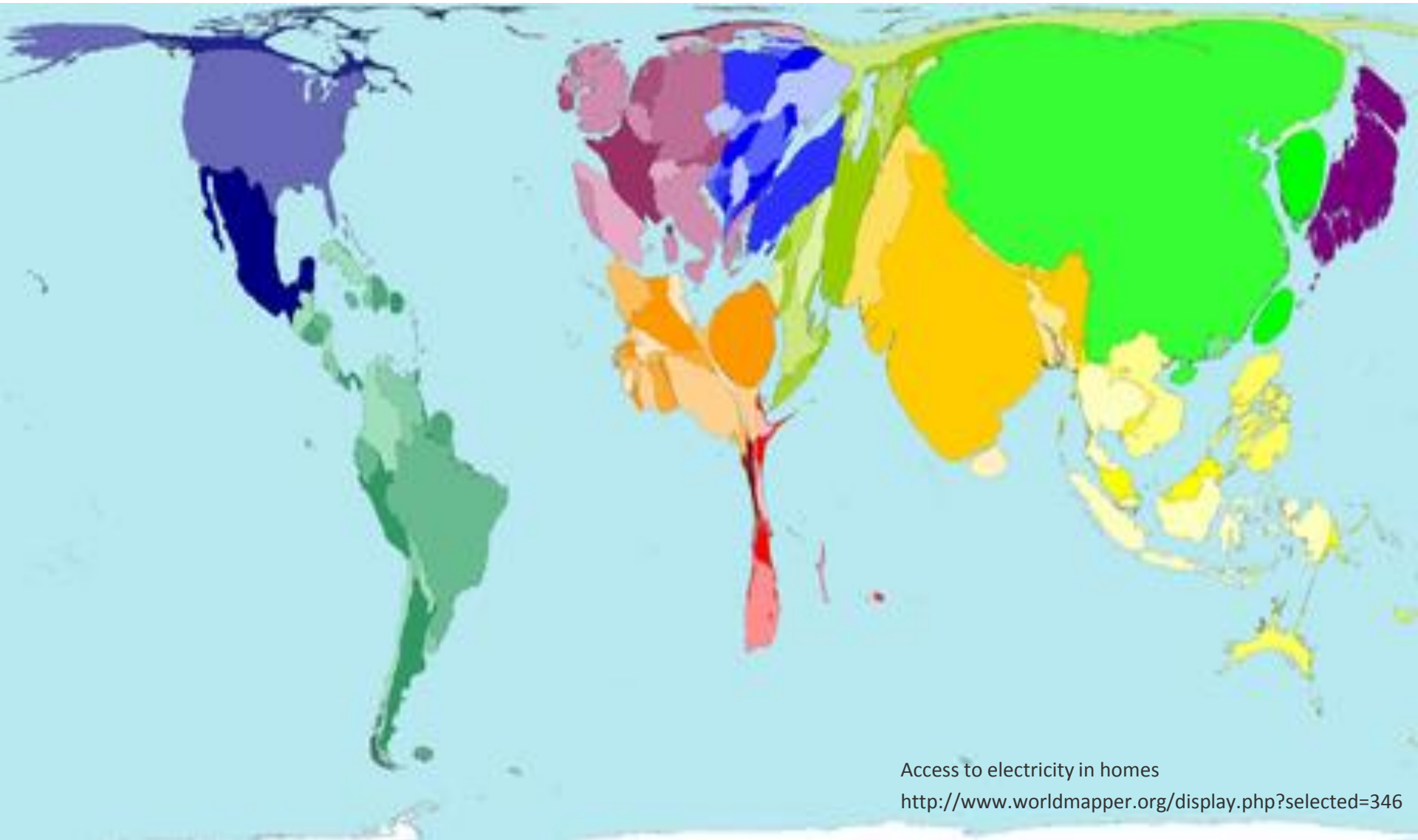






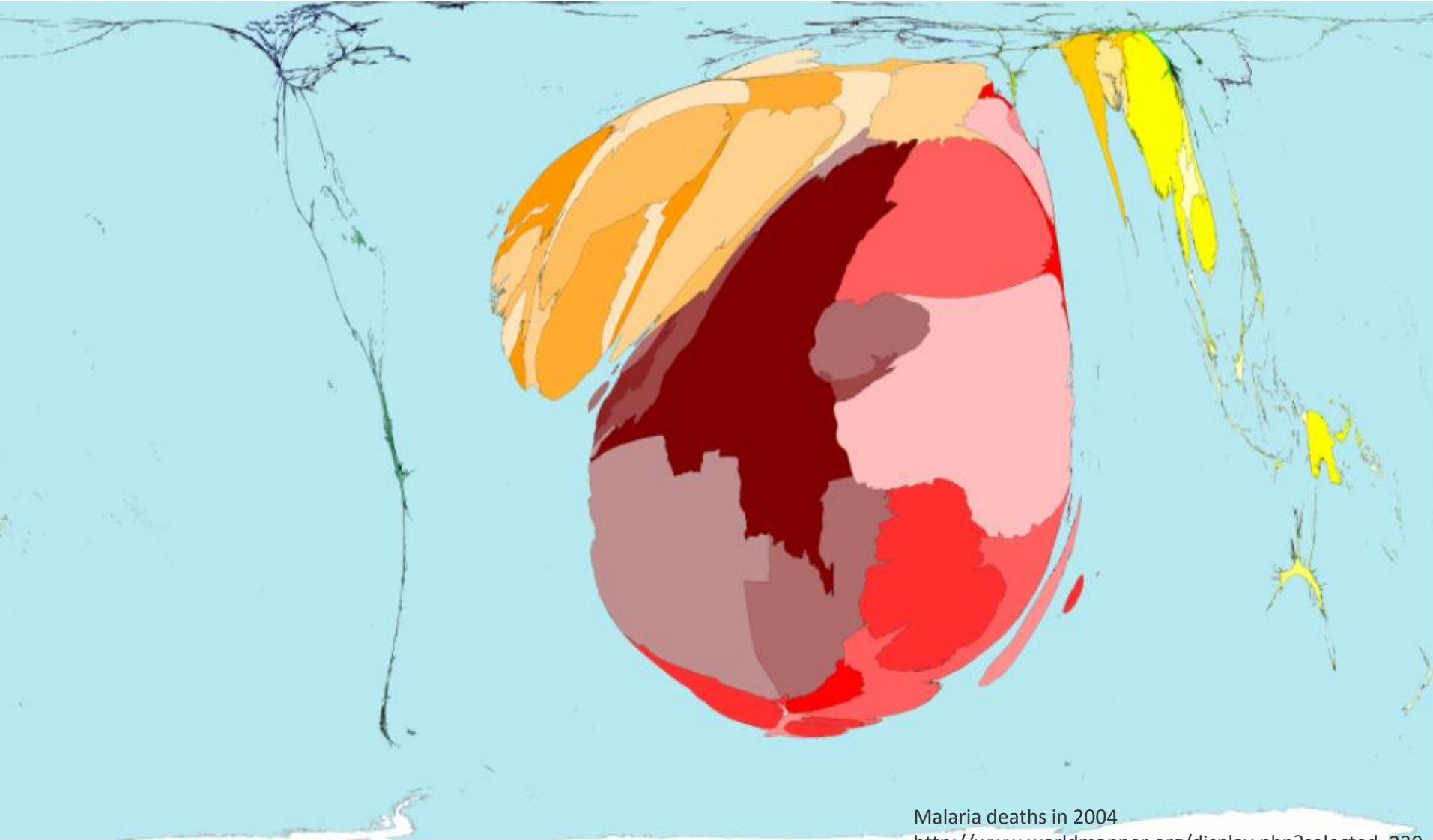


# Electricity Access



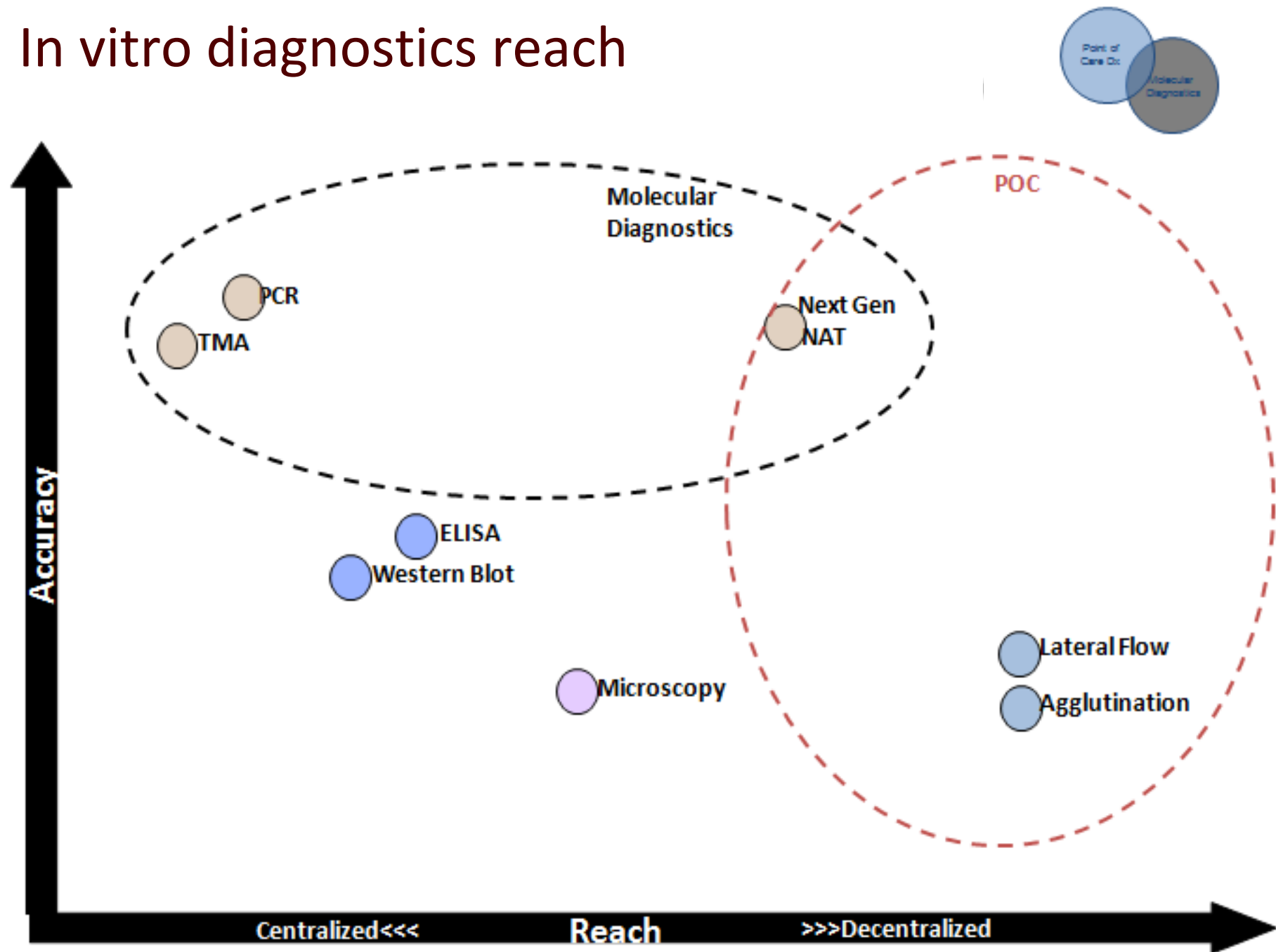


# Malaria Deaths



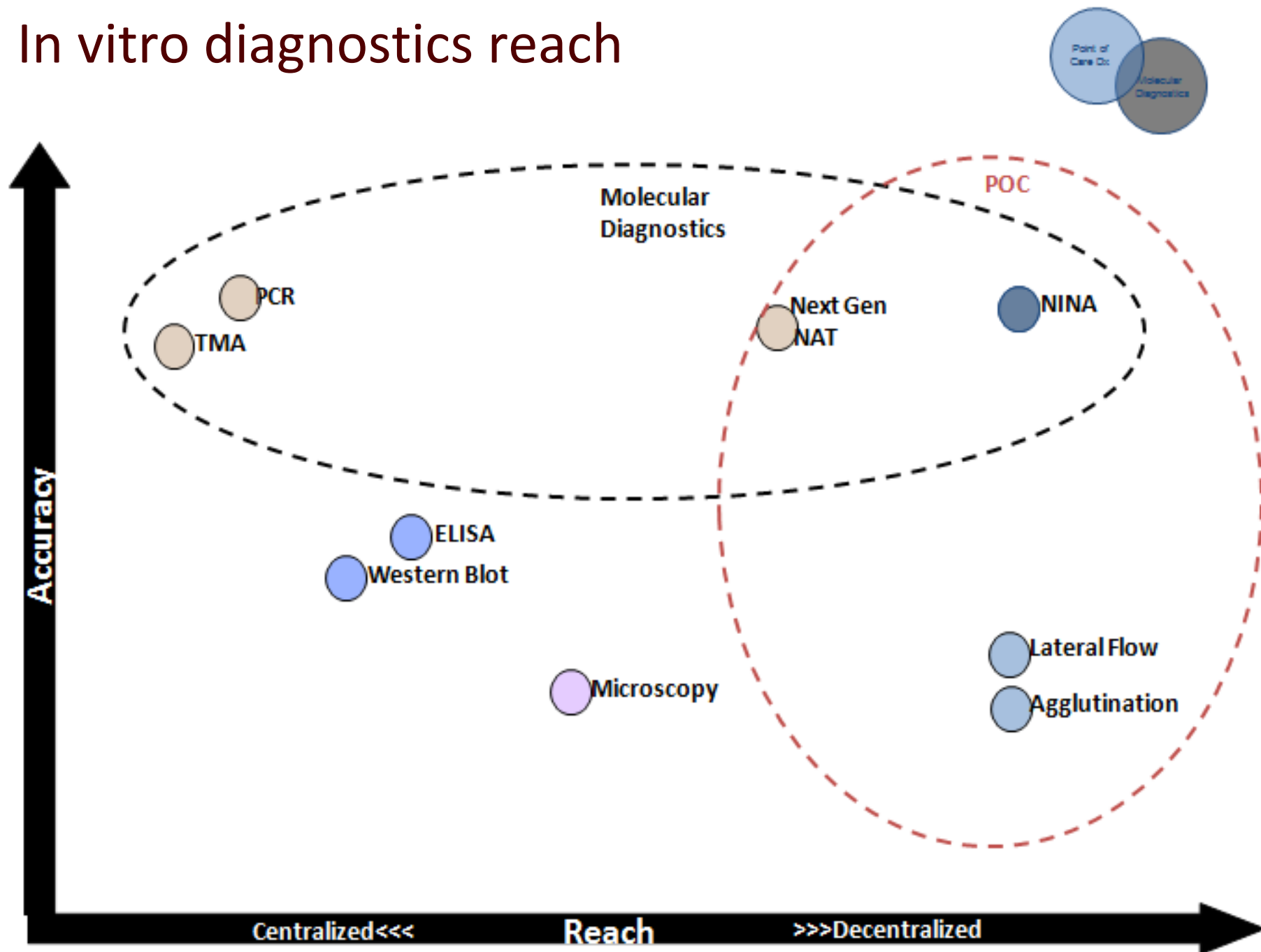
Malaria deaths in 2004  
<http://www.worldmapper.org/display.php?selected=230>

# In vitro diagnostics reach





# In vitro diagnostics reach



# Introd...

- In developing countries like Ethiopia, more sensitive, specific, cost-effective, rapid and easy method of diagnosis is crucial to the success of the NSP to eliminate and eradicate malaria
- LAMP was developed as promising simplified molecular diagnostic method appropriate in LRS

(WHO, 2014, Cordray et al,2012 ; Noemi et al, 2000; Polley et all, 2010)

# Introd...

- PATH developed a variety of NINA heaters to facilitate pathogen detection via LAMP.



## Insulation

- Foam lid
- Vacuum thermos

## Sample

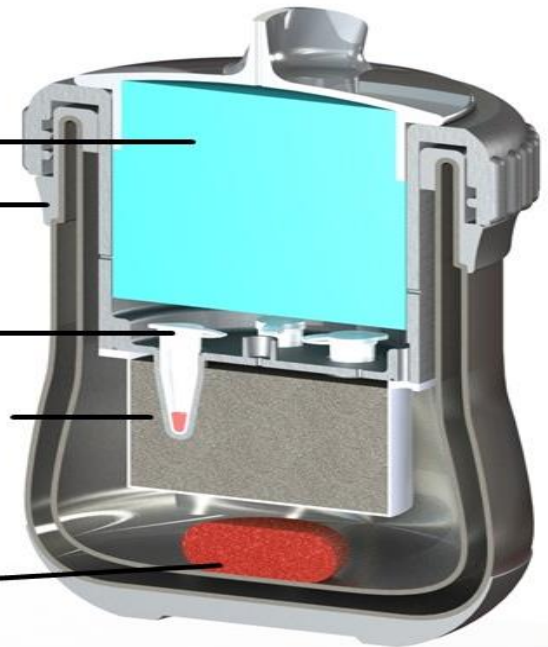
- Micro PCR tubes

## Phase Change Material (PCM)

- Custom PCM thermal enhancement additive

## Exothermic Reaction

- Disposable pouch



(Weigl et al, 2008; LaBarre et al, 2010, 2011; Singleton et al, 2014)

# What is NINA?



US008431387B2

(12) **United States Patent**  
**LaBarre et al.**

(10) **Patent No.:** **US 8,431,387 B2**  
(45) **Date of Patent:** **Apr. 30, 2013**

(54) **CHEMICAL TEMPERATURE CONTROL**

(56)

**References Cited**

(75) Inventors: **Paul Donald LaBarre**, Suquamish, WA (US); **Jay Lewis Gerlach**, Kenmore, WA (US); **Bernhard Hans Weigl**, Seattle, WA (US); **Gonzalo Jose Domingo-Villegas**, Seattle, WA (US)

U.S. PATENT DOCUMENTS

3,903,011 A 9/1975 Donnelly  
3,976,049 A 8/1976 Yamashita et al.

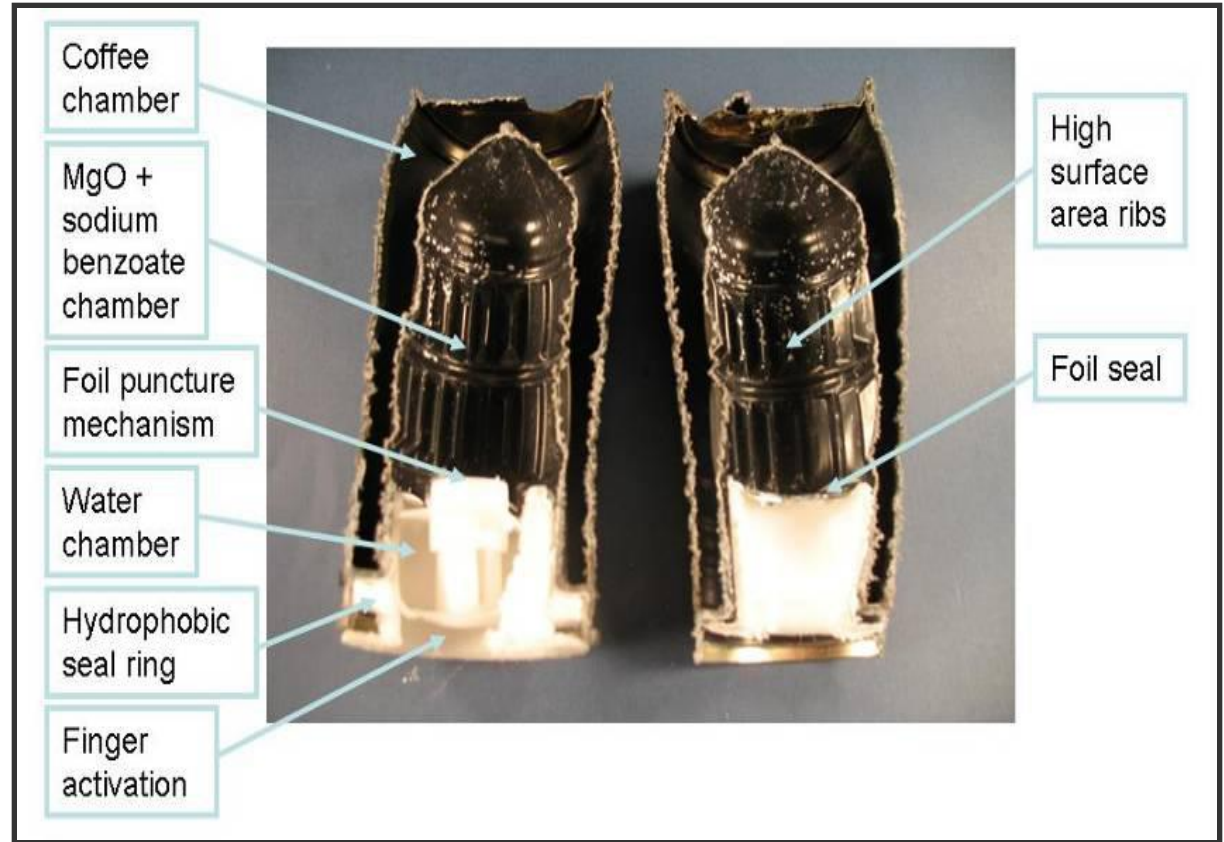
(Continued)

- **NINA:** stands for “non-instrumented nucleic acid amplification”
- **The NINA mission:** expanding access to accurate diagnostics wherever they are needed
- **The NINA vision:** 100% electricity free, infrastructure-free, sample to results, molecular diagnostics
- **Strategy**

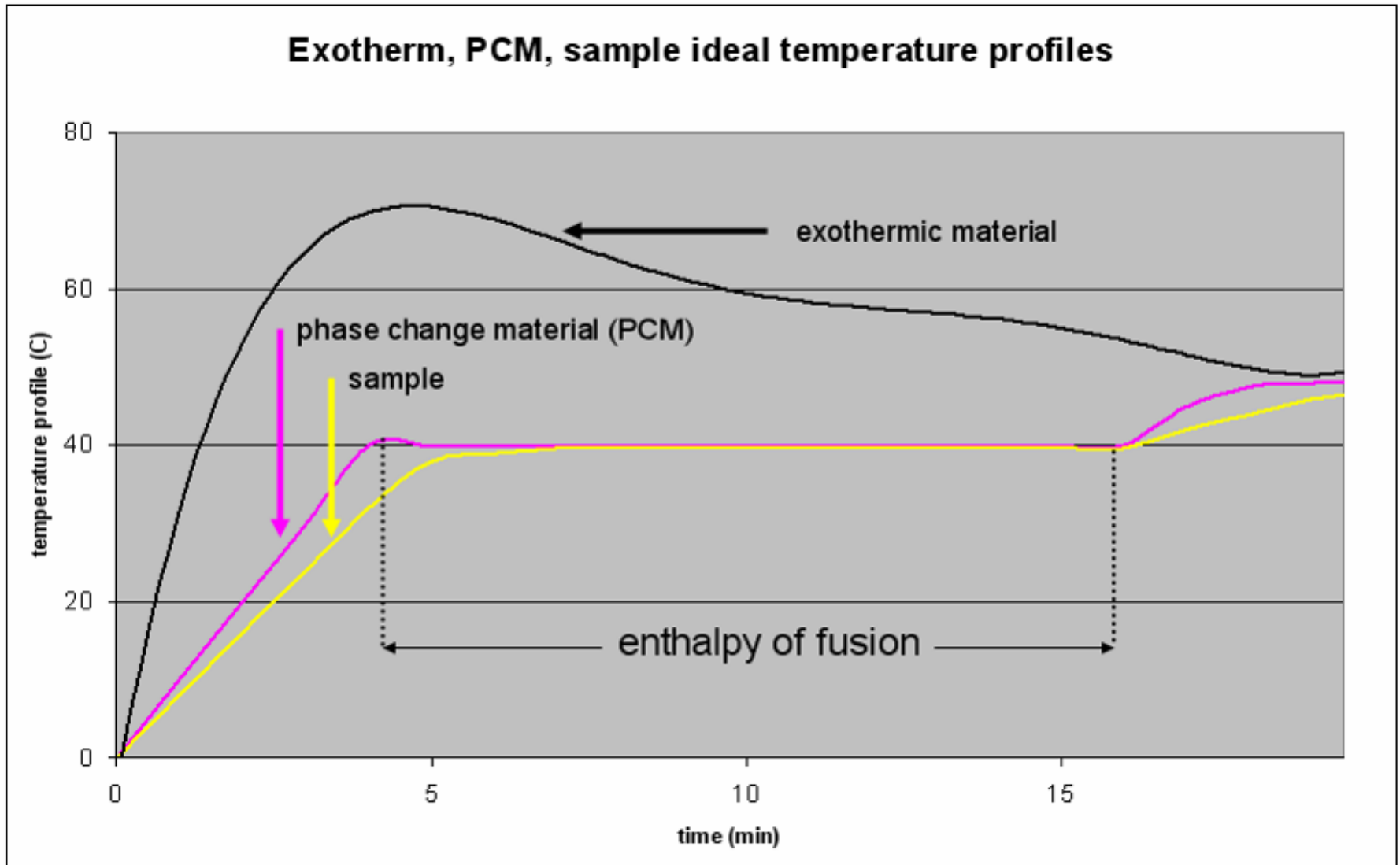


# NINA: Inspiration from a consumer product

## Inspiration from a consumer product



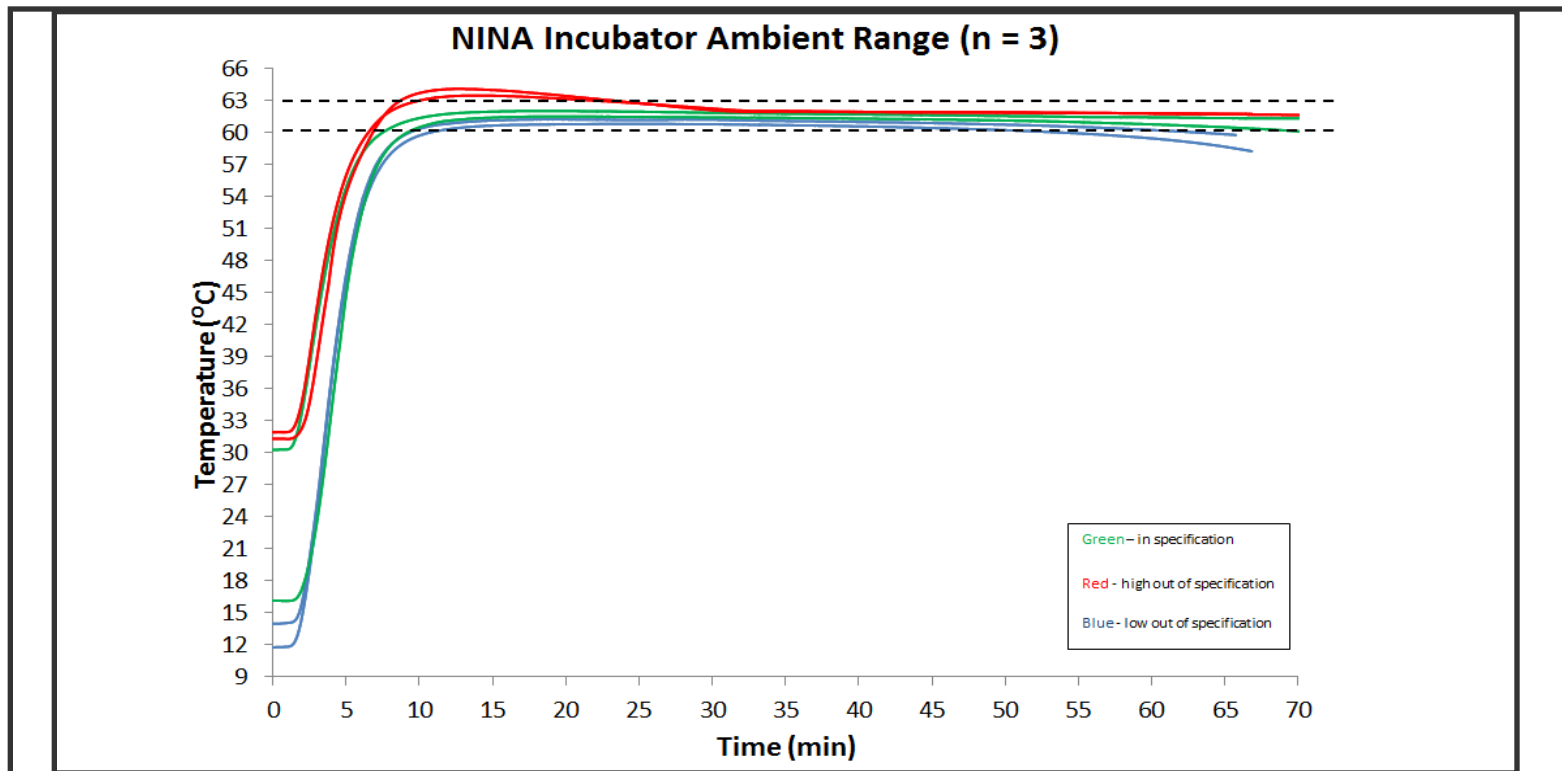
# NINA: Theory of operation



# Electricity-Free Amplification and Detection for Molecular Point-of-Care Diagnosis of HIV-1



Jered Singleton<sup>1</sup>, Jennifer L. Osborn<sup>1</sup>, Lorraine Lillis<sup>1</sup>, Kenneth Hawkins<sup>1</sup>, Dylan Guelig<sup>1</sup>, Will Price<sup>2</sup>, Rachel Johns<sup>1a</sup>, Kelly Ebels<sup>1</sup>, David Boyle<sup>1</sup>, Bernhard Weigl<sup>1</sup>, Paul LaBarre<sup>1\*</sup>



# Objectives

## ❑ General objective

- To examine the diagnostic performance of NINA-LAMP compared to microscopy and nested PCR for the diagnosis of malaria

## ❑ Specific objectives

- To compare parasite positivity by Giemsa microscopy and NINA-LAMP and nested PCR
- To determine the diagnostic accuracy of NINA-LAMP and Giemsa microscopy as compared to nested PCR



# Materials and methods

<b>Study characteristic</b>	<b>description</b>
<b>Study area</b>	Kola Diba Health center
<b>Study design and period</b>	Cross-sectional study, March to July 2014
<b>Study population</b>	All febrile malaria suspected outpatients in HC during the study period
<b>Sample size</b>	200 malaria suspected febrile patients
<b>Sampling technique</b>	Convenient sampling

# Methods...

## Sample collection and laboratory methods

**Capillary blood (200 participants)**

**Microscopic examination (Kola Diba H.C)**

**82 participants were selected**

**~4 ml venous blood**

**DBS on filter paper**

**LAMP testing(UoG)**

**Loopamp™ Pan/Pf detection kits**

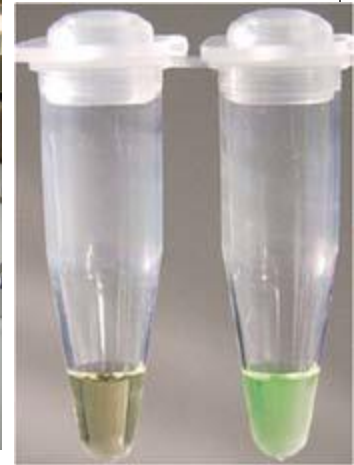
**Nested PCR (UofC)**

**LAMP re-testing**

**Data analysis and interpretation (Nested PCR - Reference method)**

# Methods...

## NINA-LAMP principle and procedure



**Boil and spin method**

**NINA H.V6 prototype heater device**      **Visual detection of turbidity**

**R. Time**

10-15 minutes

45-60 minutes



# Results and discussion

## Parasite positivity by microscopy, NINA-LAMP and nested PCR

<b>Giemsa microscopy (n)</b>	<b>Primary NINA- LAMP result (n)</b>	<b>Nested PCR (n)</b>
Positives (30), <i>P. falciparum</i> (13) <i>P.vivax</i> (17)	Positives (38), <i>P. falciparum</i> (26) Non- <i>P. falciparum</i> (12)	Positives (31), <i>P. falciparum</i> (13) <i>P. vivax</i> (11), <i>P. ovale</i> (7)
Negatives (52)	Negatives (44)	Negatives (51)

# Result...

## Diagnostic accuracy of NINA-LAMP and microscopy using nested PCR as reference method

Method		Sensitivity(%)	Specificity(%)	%agreement
NINA -LAMP	Pan (Gondar)	<b>96.8</b>	<b>84.3</b>	<b>89.0</b>
	Pf (Gondar)	<b>100</b>	<b>81.2</b>	<b>84.1</b>
	Pan (Calgary)	<b>96.8</b>	<b>98.0</b>	<b>97.6</b>
	Pf (Calgary)	<b>100</b>	<b>100</b>	<b>100</b>
Microscopy	Pan	<b>93.6</b>	<b>98.0</b>	<b>96.3</b>
	Pf	<b>92.3</b>	<b>100</b>	<b>98.8</b>



# Result...

## □ NINA- LAMP

- Satisfies the WHO recommendation of diagnostic kits sensitivity of  $> 95\%$
- Comparable performance with nested-PCR and superior to Microscopy
- Comparable accuracy to other LAMP evaluation studies conducted using nested PCR as the reference method.

# Result...

## □ **Good operational characteristics**

- Previous experiences was not required to perform
- Reading of test result and interpretation was easy
- The whole test procedure takes about 60-80 minutes
- Special equipments were not required
- The reagents were thermostable

# Limitations of the study

- Use of relatively small sample size
- The use of microscopy for case detection
- Failure to obtain “asymptomatic” or low parasitemia cases.
- sample preparation still requires a centrifugation step

# NINA Reusable Housing Platform (RHP)

- \$34.37/prototype
- \$0.22/test
- 5x 0.2mL PCR tubes
- Maintains 62-65°C for 60min
- Vacuum thermos insulation
- Simple activation
- 240cc device volume



# NINA Single-Use Disposable (SUD)

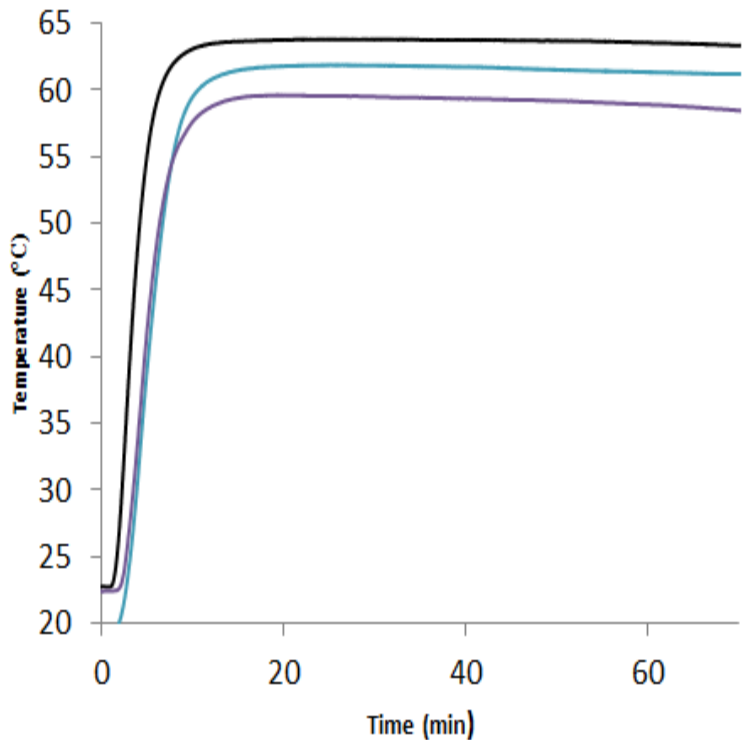
- \$1.83/prototype
- 1x 0.2mL PCR tube
- Maintains 58-60°C for 45min
- Closed-cell foam insulation
- 100cc device volume





# Current collaborations & operating temps

Range of Operating Temperatures



Collaborator	Assay	Temperature Requirement	Phase Change Material (PCM)
Centers for Disease Control, USA	LAMP, HIV-1	58 – 61C	PureTemp 60
Center Pasteur, Cameroon	RT-LAMP, <i>Pf</i> gametocytes	62-67C	PureTemp 65
New England Biolabs	LAMP, <u>Filariasis</u>	62-64C	Palmitic Acid
University of Calgary, Canada	LAMP, <i>Plasmodium</i> spp.	62 – 65C	Puretemp 65
PATH, USA	LAMP, HIV-1	60 – 63cC	Palmitic Acid
Noguchi Research Institute, Ghana	LAMP, Buruli Ulcer	62 – 65C	PureTemp 65
<i>*Proposed, Lucigen</i>	<i>OmniAmp (LAMP), Pf and EBOV</i>	69-72C	<i>Na2SiO3-5H2O (proposed)</i>

# Acknowledgements

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Sema et al. Malaria Journal (2015) 14:44

<http://www.malariajournal.com/content/>

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- Kola Diba health center laboratory staffs and study participants

Thank you!