Efficacy of *Bacillus subtilis* based probiotic growth performance, fecal microbiota and intestinal morphology of broiler chickens

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>Importance of probiotic in digestive trace

- > What makes *Bacillus subtilis* different that other microbes
- Materials and methods
- Results and discussion
- Conclusions

Significance of gut

- Barrier to ingested chemicals, feed contaminants and pathogens
- Absorption of nutrients and water
- Components: physical, chemical, immunological and microbiological
- Intestine is the largest immune organ
- Gut microbes perform functions such as:
 - nutrient digestion and absorption
 - gut health and integrity
 - competitive exclusion of pathogens
 - immunomodulation







Development of the microflora



Microbes in GIT



Under commercial rearing conditions birds are exposed to more stressful situations like transportation, vaccination, high flock density, and heat stress.





"Live microorganisms which when administered in adequate amounts confer a health benefit on the host"

FAO/WHO, 2002

The most frequently used organisms for probiotic preparations are:

- Bacillus
- Lactic acid bacteria (LAB)
- Live yeast



Probiotics... Mode of Action?



1. Competitive exclusion of pathogens



 Improved immune status: production of antibacterial defensins and mucin

3. Maintain epithelial integrity and barrier function 4. Nutritional effects:

- Production of enzymes
- Production of vitamin B₁₂ and K





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- Definition of Bacillus
 - Gram positive organisms
 - Spore formers
- *Bacillus* are found in environmental samples obtained from virtually anywhere on Earth (soil, plants, fresh and salt water, rocks etc.).
- *Bacillus* are administered in the form of spores, a dormant resistant stage which transform to vegetative cells when entering the intestinal tract of the animal.
- Bacillus spores are
 - pH stable
 - Trypsin stable
 - Thermo-tolerant
 - Storage and in-feed stable





Spores of a carefully selected *Bacillus subtilis* with the following major characteristics



- Heat tolerance
- pH stability, even at pH 2-3
- Compatibility with other feed ingredients including organic acids, coccidiostats and antibiotic growth promoters
- Approved by the European Food Safety Authority (EFSA) for application in feed

Rationale for the experiment



- Under commercial feeding conditions birds are exposed to a variety of stress factors which make them more susceptible to gastrointestinal and metabolic disturbances.
- The inclusion of sub-therapeutic antibiotic growth promoters (AGPs) into poultry diets is common to overcome these gastrointestinal challenges. However, as the use of AGPs is being more and more limited and/or banned throughout the world, there is continuous search to identify alternative strategies
- Probiotics, prebiotics, synbiotics, organic acids and phytogenic feed additives could be such alternatives with probiotics being one of the most favorable.
- A Bacillus subtilis (DSM 17299) spore-forming probiotic seem to be most suitable candidates for in-feed applications because of their spore forming ability. These spores are tolerant to heat, harsh pH conditions, pressure, coccidiostats and antibiotics..





 To evaluate the effects of feeding *Bacillus subtilis* (DSM 17299) or antibiotic growth promoters (AGP) alone or in combination on growth performance, fecal microbiota and gut morphology of broiler chickens.





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Experimental Design

- Birds: 480 Cobb male broilers
- Housing: Open house-Floor pen
- **Growth phases :** 0-21 d (starter), and 22-42 d (grower)
- Duration of experiment:0-42 days of age
- Feed form: Mash
- Temperature and humidity: 26-34°C and 82-93%
- Diets of each growth phase:
 - Diet 1: Control
 - Diet 2: Control + AGP
 - Diet 3: Control + Bacillus subtilis (500 g/ton)
 - Diet 4: Control + AGP* + Bacillus subtilis (500 g/ton)
 - *AGP = a combination of oxytetracyclin and neomycin at 100 ppm (w/w)
- Replication: 6 Replicates/treatment (20 birds/replicate)
- Parameters: Growth performance, Fecal microbial counts [Lactic acid bacteria (LAB) and Enterobacteriaceae (ENT)] and Intestinal morphology

Ingredient and nutrient composition of basal diets



Ingradianta	Dietary		
ingreatents	Starter	Finisher	
Corn	506.00	549.00	
Soybean	293.82	269.00	
Wheat Pollard	60.72	63.75	
СРО	36.00	32.00	
Fish Meal 55%	76.00	50.00	
L-Lysine	2.50	2.50	
DL-Methionine	2.00	2.00	
Monodicalciumphosphate 21	10.00	14.00	
Calcium carbonate	6.80	9.92	
Choline chloride	0.60	0.58	
Salt	2.50	2.50	
Other (Minerals, Vitamin and toxin binder)	3.05	3.05	
Calculated analysis:			
Crude protein, %	22.50	20.34	
ME, MJ/kg	12.22	12.18	



- Body weight and feed intake on weekly basis and FCR was calculated accordingly.
- A total of 18 samples from each treatment in triplicates were determined for the faecal LAB and ENT population using the method as described by Foo et al. (2003b).
- A total of 20 samples from each treatment were used to study the intestinal morphology. The procedure was a modified method as described by Hair-Bejo (1990).
 - Segments of 5 to 6 cm long were removed from the duodenum, jejunum, and ileum collected and flushed with 10% neutral buffered formalin solution and were then used for morphometric analysis.
 - The morphometric variables examined included: villus height (from the tip of the villi to the villi crypt junction), crypt depth (defined as the depth of the invagination between adjacent villi). Values are means from the best 20 villi and only vertically oriented villi and crypts from each slide were measured.





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	Control (C)	C+AGP	C + <i>Bacillus</i> <i>subtilis</i> DSM 17299	C+AGP + <i>Bacillus subtilis</i> DSM 17299	SEM
BWG, kg	2.09	2.13	2.15	2.18	0.01
FI, kg	3.65 ^a	3.66 ^a	3.55 ^b	3.60 ^a	0.05
FCR, kg/kg	1.75 ^a	1.73 ^{ab}	1.66 ^b	1.65 ^b	0.02

*BWG = Body weight gain, FI = Feed intake and FCR = Feed conversion ratio

^{a-b} Values with different superscript letters within a row indicate significant differences (P < 0.05).

Results and discussion: fecal microbiology



Microbial count, log ₁₀ CFU/g	Control (C)	C+AGP	C+ <i>Bacillus</i> <i>subtilis</i> DSM 17299	C+AGP + <i>Bacillus</i> <i>subtilis</i> DSM 17299	SEM
LAB	6.15 ^b	6.42 ^b	6.89 ^a	6.98 ^a	0.09
ENT	5.73 ^a	5.40 ^{ab}	4.64 ^c	5.21 ^b	80.0

^{a-c} Means within a row with common superscripts are not significantly different (P > 0.05); LAB=lactic acid bacteria; ENT=*Enterobacteriaceae*.

Ileal total bacteria and E.coli counts





(Doranalli et al., 2013)

Digestive tract micro flora analysis



- A total of 3000 day-old male chicks (Ross 308) were housed in 12 pens (250 birds/pen)
- Two dietary treatments: corn-soy based standard feed -/+ Bacillus subtilis
 DSM 17299
- At age 35 days of age, 3 birds were randomly selected per pen unit for gut excision and collection of ileal digesta for determination of micro flora profile



Increases beneficial bacteria in the intestine



Dice coefficient analysis of the ileal profiles from control birds and birds fed *Bacillus* subtilis DSM 17299



Results and discussion: Intestinal morphology



	Control (C)	C+AGP	C+Bacillus subtilis DSM 17299	C+AGP+Bacillus subtilis DSM 17299	SEM
Villi height, µm					
Duodenum	1682.47 ^c	1756.38 ^b	1752.00 ^b	1871.62 ^a	9.70
Jejunum	1183.38°	1200.43 ^c	1407.74 ^a	1310.50 ^b	9.00
lleum	738.83 ^b	676.96 ^c	872.35 ^a	843.80 ^a	7.00

^{a-c} Means within a row with common superscripts are not significantly different (P > 0.05); VH=villus height

Conclusions



- It can be concluded that inclusion of *Bacillus subtilis* (DSM 17299) to broiler diets improved growth performance and small intestinal morphology.
- Synergistic effects were observed in combination of *Bacillus subtilis* and AGP for some response variables.
- In addition, *Bacillus subtilis* supplementation showed its effects on modulating microbial populations, which was evidenced by increased amounts of LAB, mostly accounting for beneficial gut microbiota and reduced counts of *Enterobacteriaceae*, the latter group containing gut pathogenic bacteria like *E. coli* and *Salmonella*

