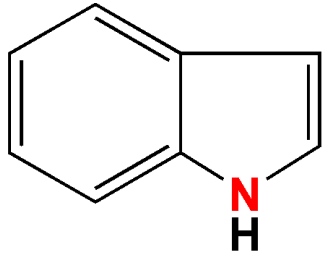


# Applications of signaling molecule **indole** to control biofilm formation and host immunity



**Indole as a signaling molecule**



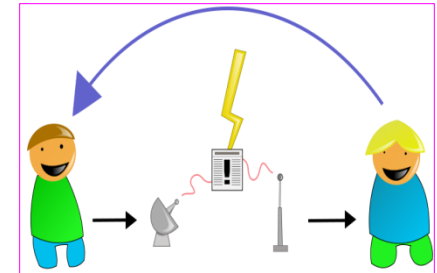
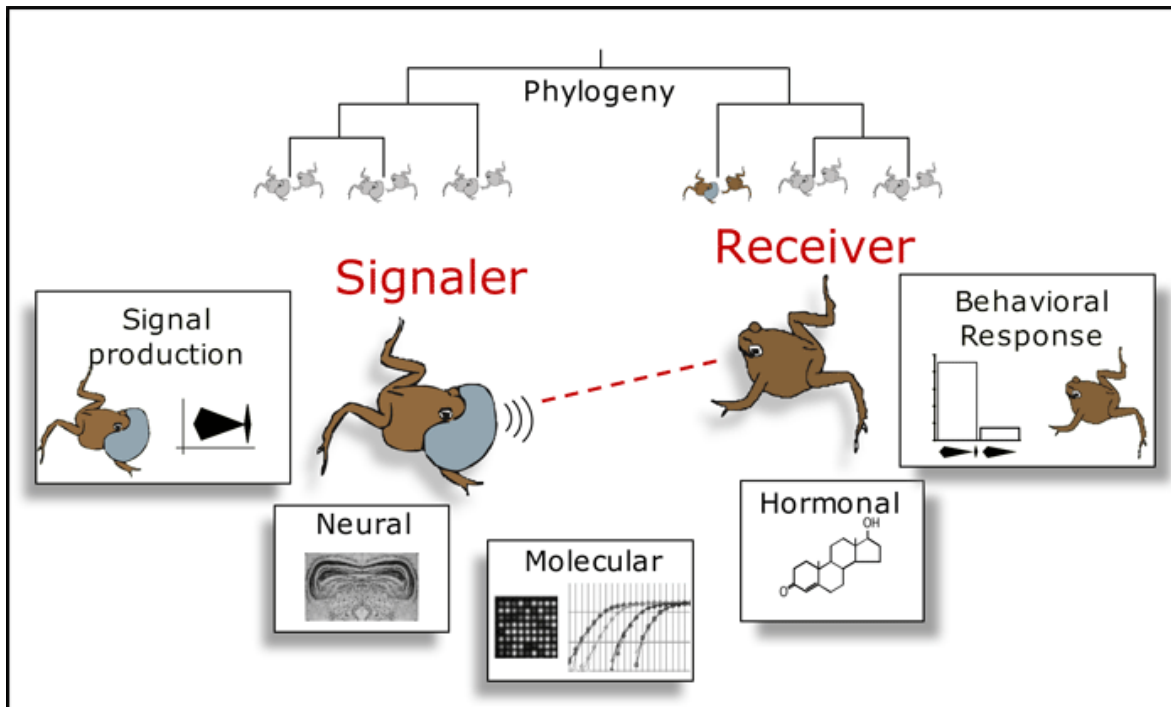
**Jintae Lee**  
**Yeungnam University, Korea**

**2017. 2. 27.**

**7<sup>th</sup> Clinical Microbiology**



# Animal Communication



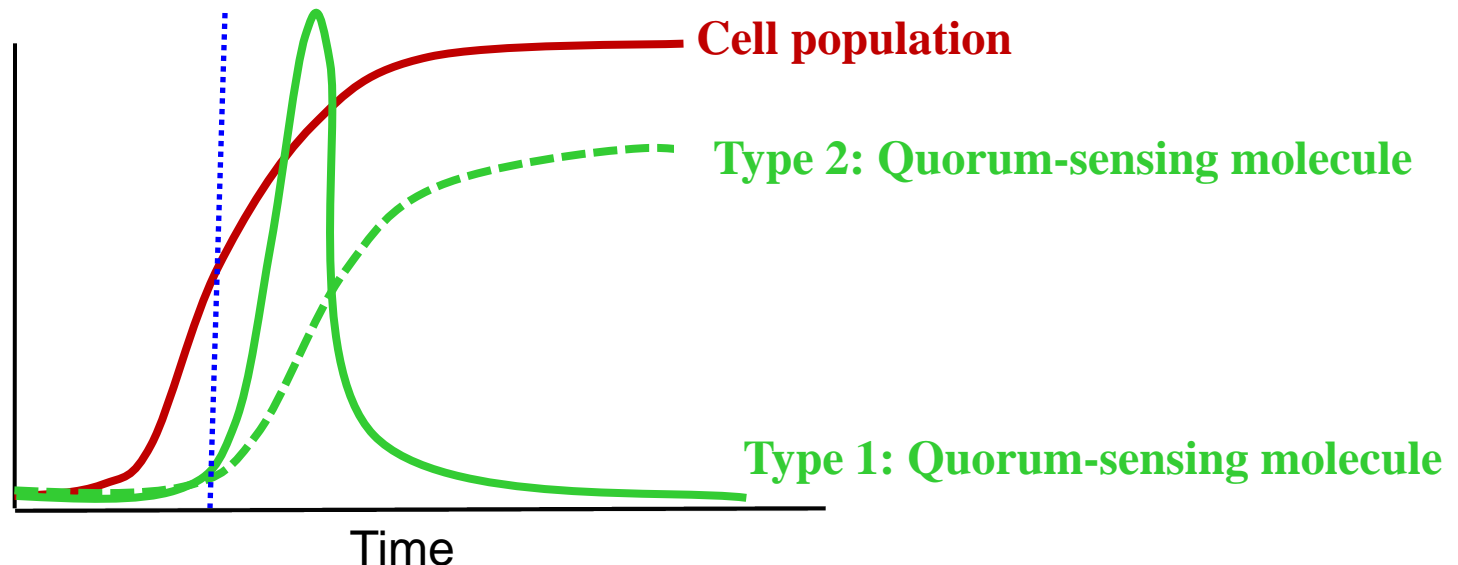
# Bacterial Communication: Quorum Sensing (QS)

## Sensing neighbors

**Quorum sensing (bacterial social behavior):**

**the ability of gene expression regulation in response to increasing cell population density by producing small diffusible chemicals (Fuqua *et al.*, 1994, *J. Bacteriology*)**

**Quorum-sensing molecule ~ Autoinducer**



# Why Quorum Sensing (QS)?

Well-being

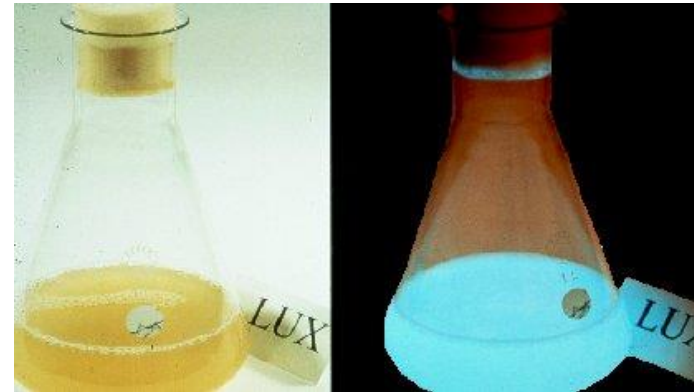
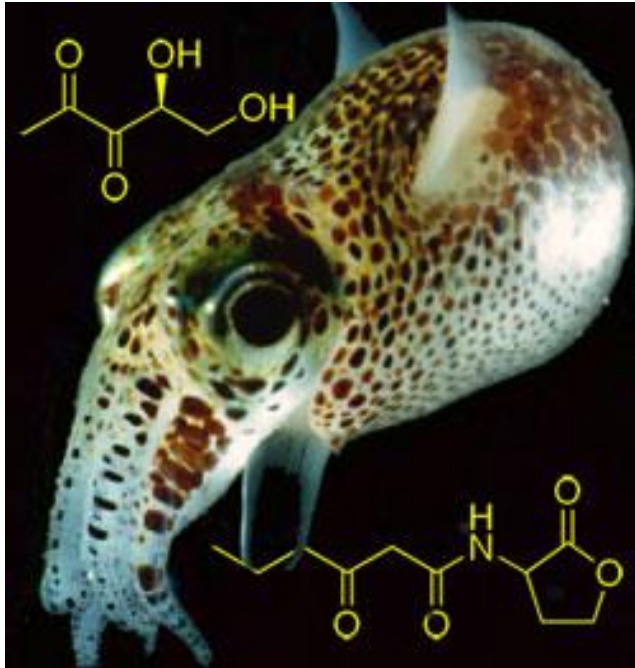
To sense and response to environmental stimuli including cell population

Quorum-sensing controlled processes → well-survival !

- 1) Bioluminescence (symbiosis)
- 2) Virulence factor production (pathogenesis)
- 3) Biofilm formation
- 4) Competence (conjugation)

# Bioluminescence (Symbiosis)

Win-Win



LUX (luminescence phenotype): luciferase

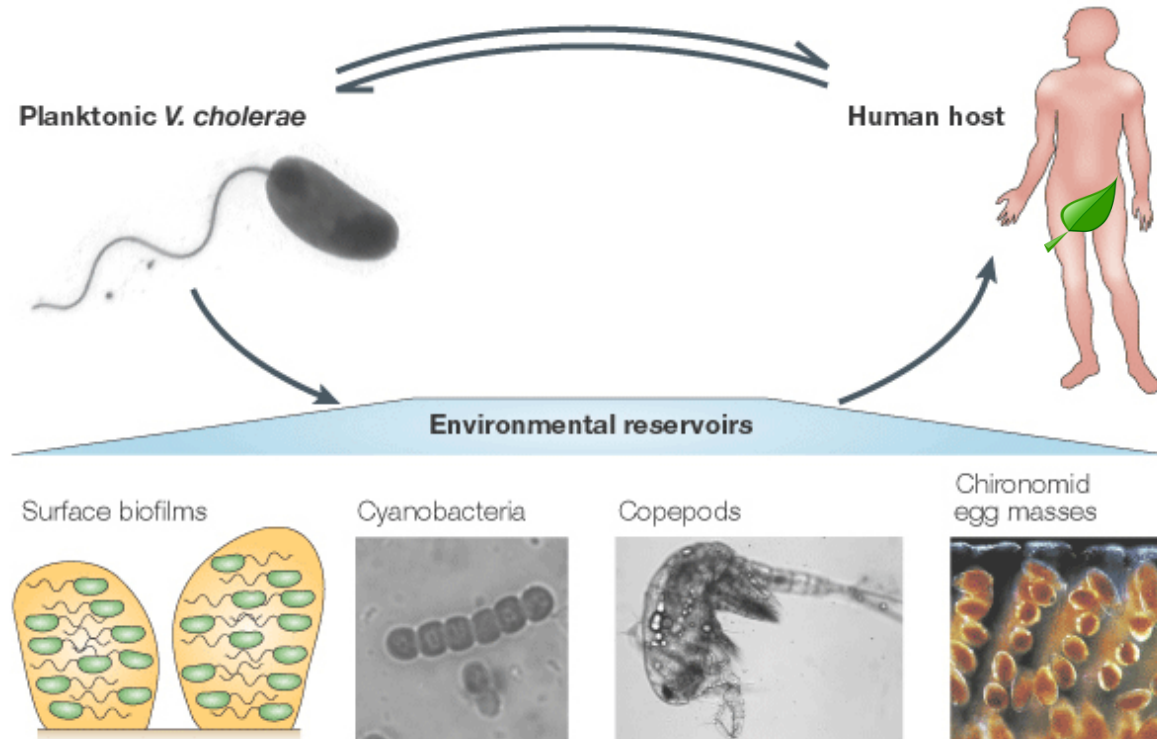
*Vibrio harveyi* in Hawaiian bobtail squid

Bacteria provide a light to the squid at night and the squid provides a shelter to bacteria

Engebrecht *et al.*, 1983, *Cell*

# QS Regulates Virulence (Disease)

Weapons



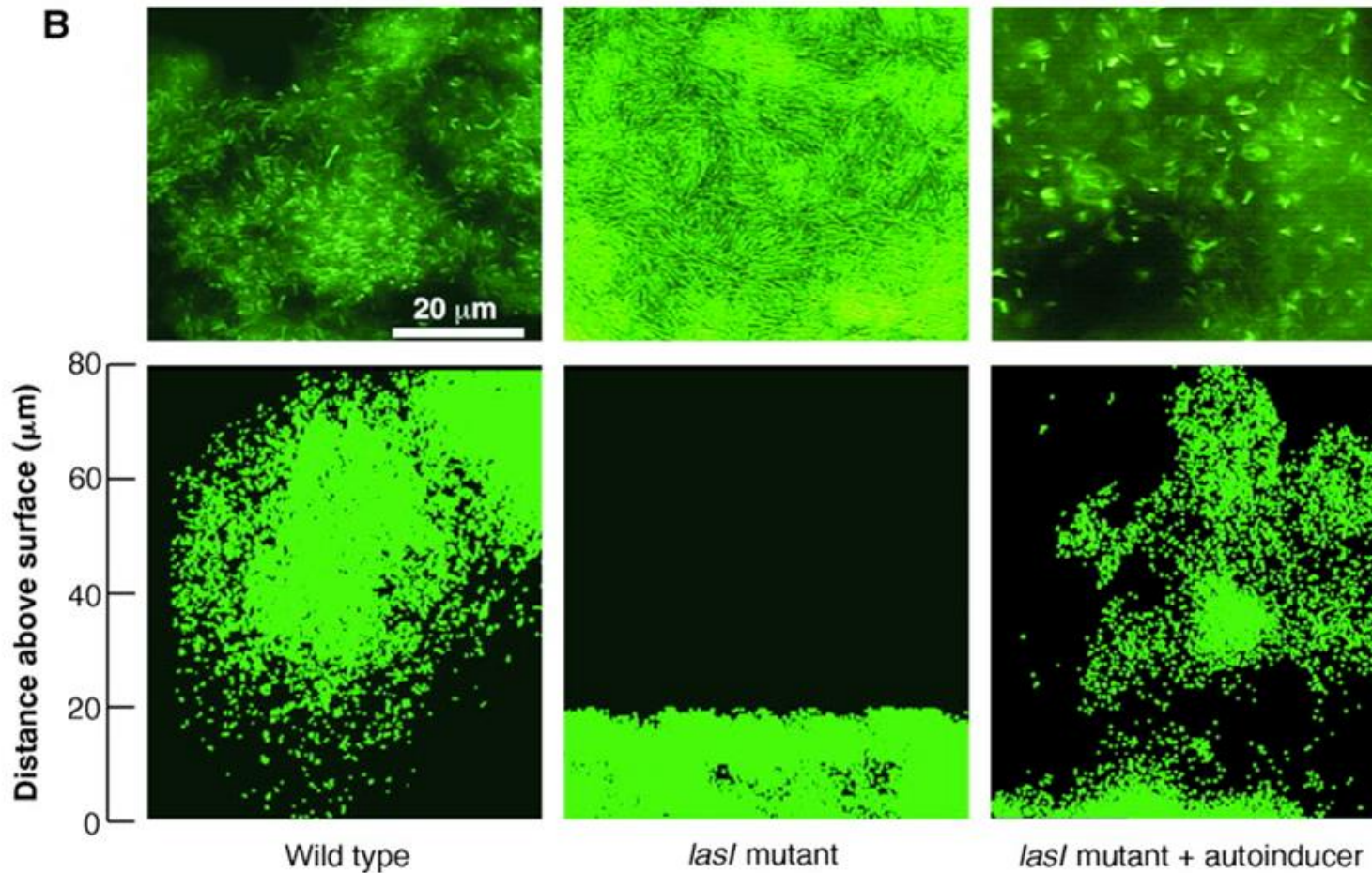
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Nature Reviews | Microbiology

QS regulates virulence in *Vibrio cholerae* (Miller *et al.*, 2002, *Cell*)



# QS Controls Biofilm Formation

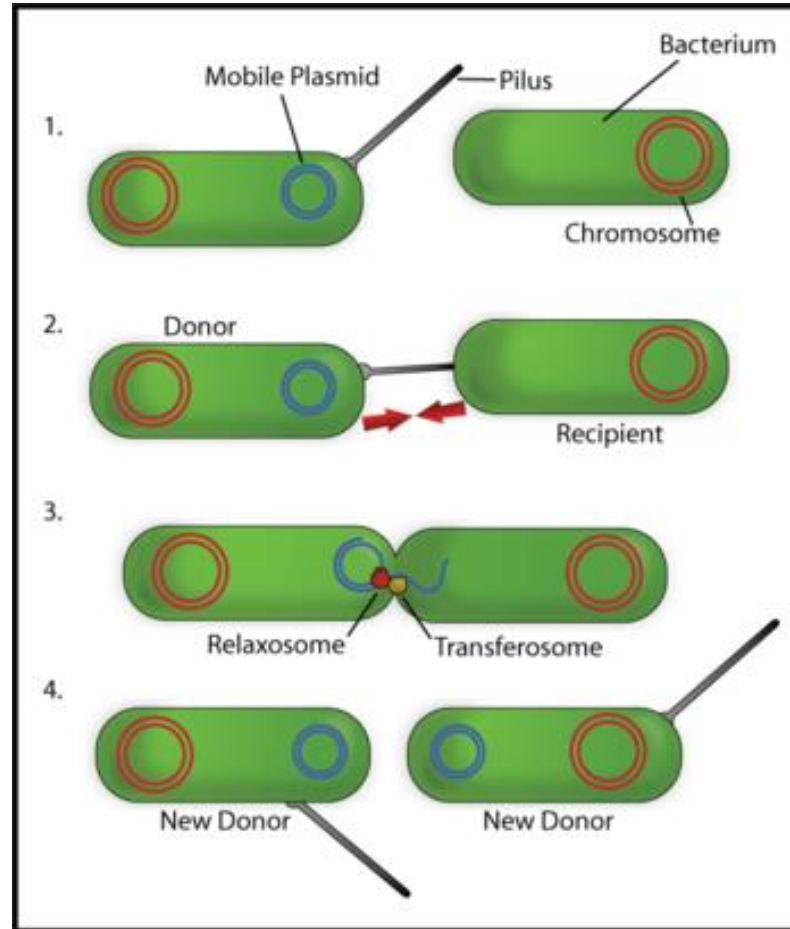
Castles



QS molecules control biofilm formation of *Pseudomonas* (Davies *et al.*, 1998, *Science*)

# QS Increases Competence (Conjugation)

Sex



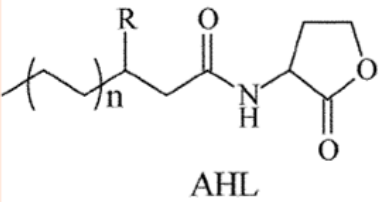
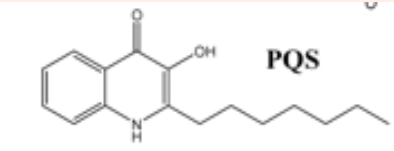
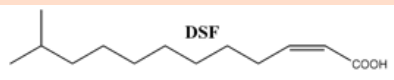
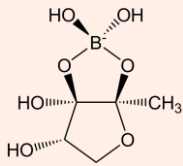
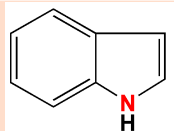
**Mobile plasmid helps bacteria survive in the presence of antibiotics.**

**Pappas and Winans, 2003, *Mol. Microbiol.***



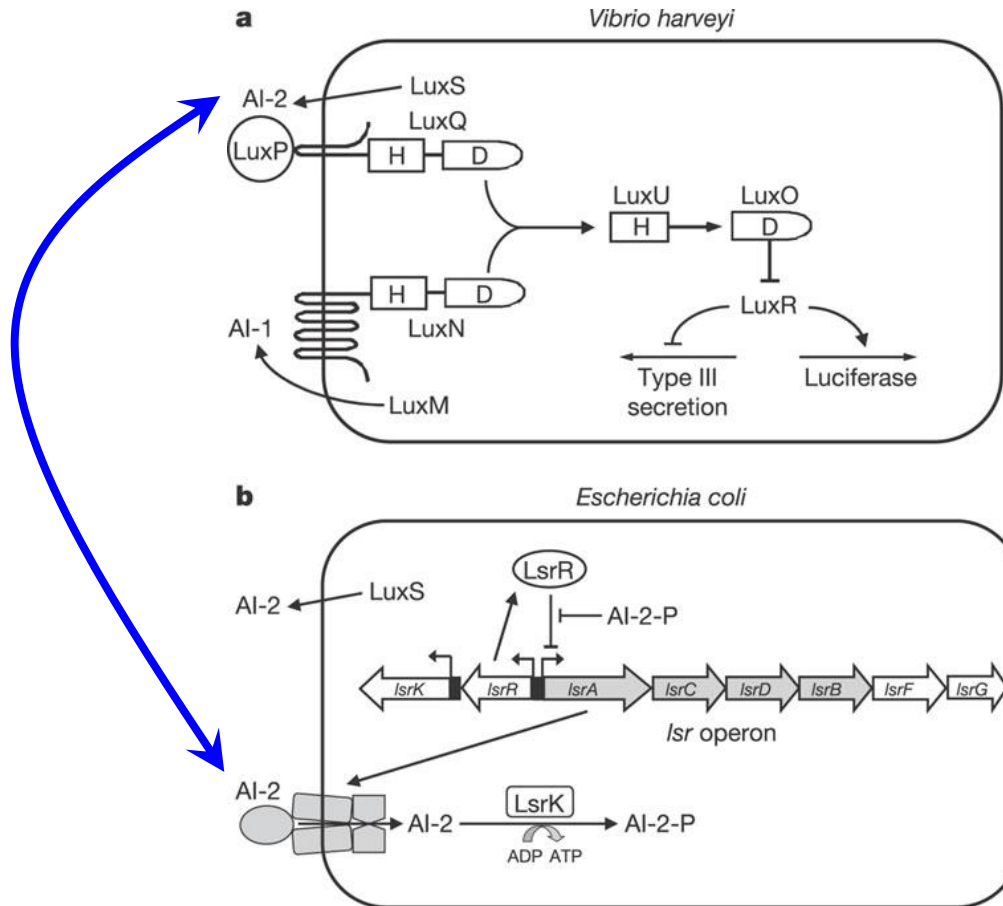
# What Kinds of QS Molecules?

Many languages

Molecules	Structure	Strains	Reference
<i>N</i> -acyl homoserine lactones (AHL)		<i>Vibrio fischeri</i> <i>Pseudomonas aeruginosa</i>	Gambello <i>et al.</i> , 1981, <i>Cell</i> Latifi <i>et al.</i> , 1991, <i>J. Bacteriol.</i>
<i>Pseudomonas</i> quinolone signal (PQS)		<i>Pseudomonas aeruginosa</i>	Pesci <i>et al.</i> , 1999, <i>PNAS USA</i>
Diffusible signal factor (DSF)		<i>Xanthomonas campestris</i>	Barber <i>et al.</i> , 1997, <i>Mol. Microbiol.</i>
Autoinducer 2 (AI-2)		Some of Gram-positive and -negative	Chen, <i>et al.</i> , 2002 <i>Nature</i>
<b>Indole</b>		<i>Vibrio cholerae</i> <i>E. coli</i>	<b>Smith, 1897,</b> <b><i>J. Exp. Med.</i></b>

# Interspecies QS Molecule AI-2

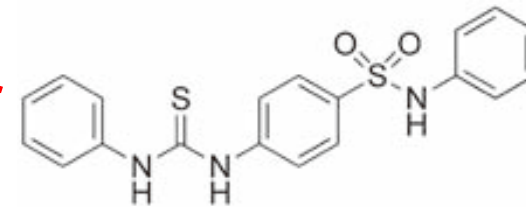
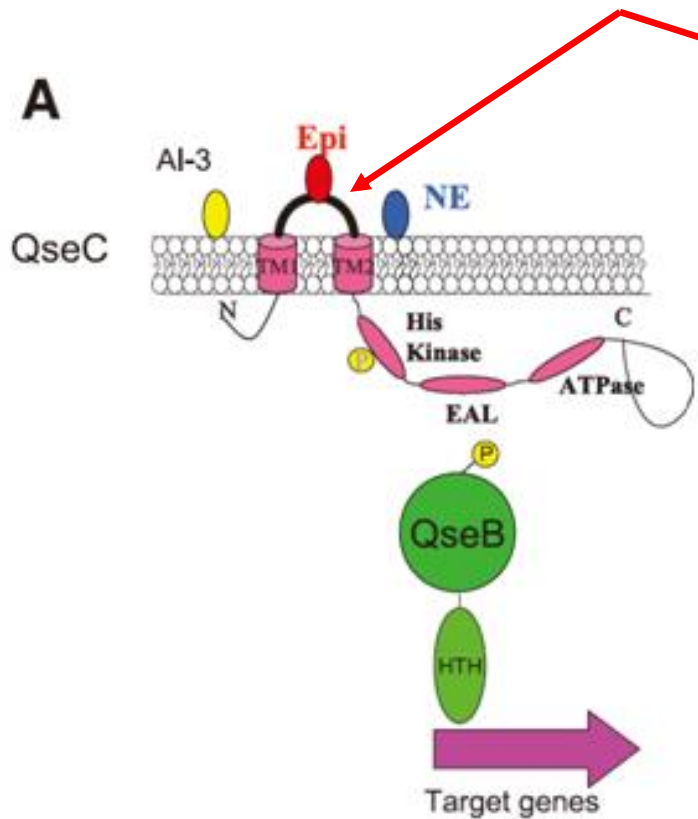
Global language



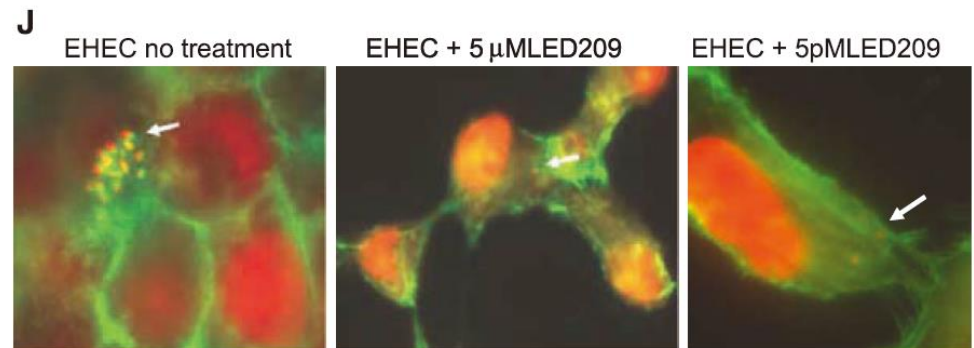
*E. coli* and *V. harveyi* communicate using AI-2 (Xavier and Bassler, 2005, *Nature*)

# Application of QS: New Type of Antibiotic

No communication → No disease !



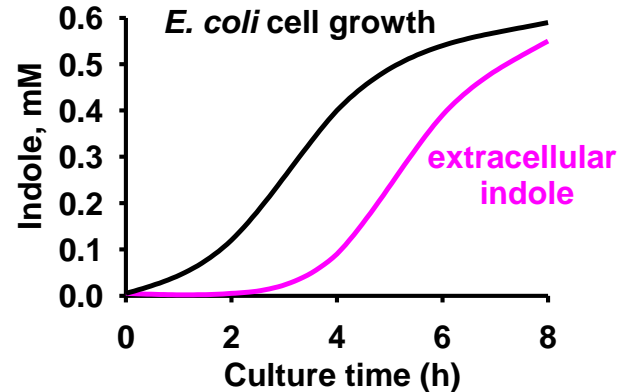
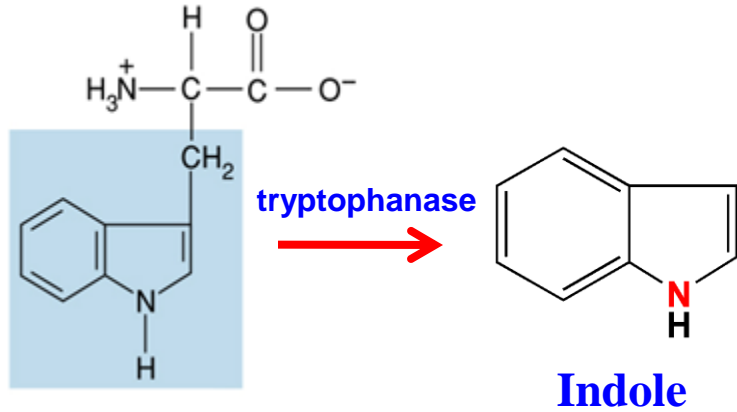
LED209 screened from  
a library of 150,000 small organic compounds



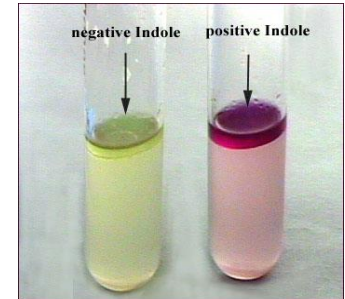
An inhibitor of QS sensor kinase decreases virulence of *E. coli* O157: H7

(Rasko *et al.*, 2008, *Science*)

# Indole Overview



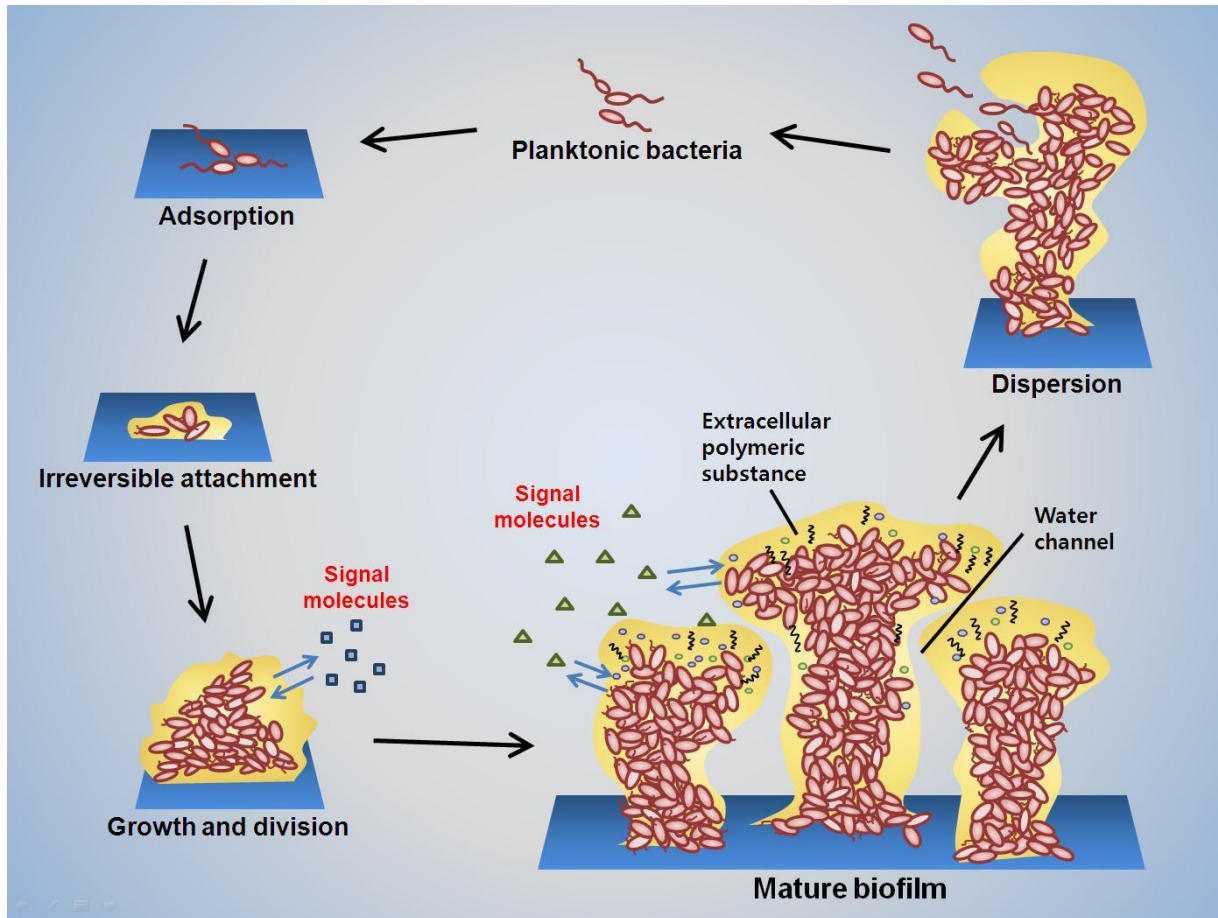
**A language**



- The average human intestine contains more than 1 kg of bacteria (~100 trillion bacterial cells).
- *E. coli* produces a large amount of indole during stationary growth phase (Smith, 1897, *J. Exp. Med.*)
- Indole enhances plasmid stability (Chant and Summers, 2007, *Mol. Microbiol*)
- Indole increases population-wide antibiotic resistance (Collins *et al*, *Nature*, 2010; Collins *et al*, *Nature Chemical Biology*, 2012; Collins *et al*, *PNAS* 2013)
- Prof. Thomas. K. Wood and I have found that indole is a QS molecule during biofilm study.

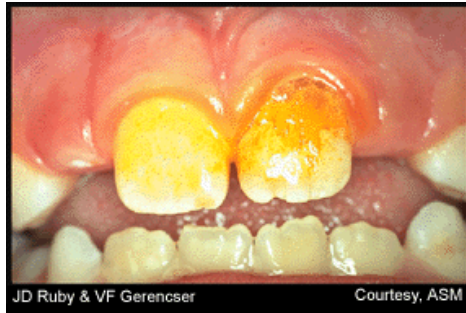
# Bacterial Biofilm

Castle

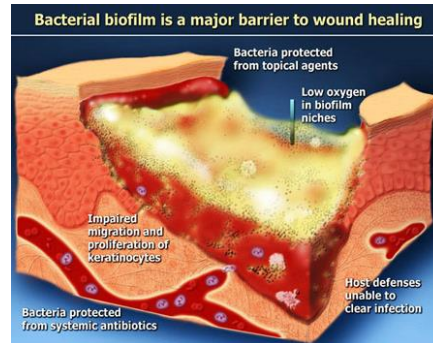


**Bacterial biofilms: a surface attached community of bacteria made of extracellular polymeric substances (carbohydrates, adhesive fibers, eDNA)**

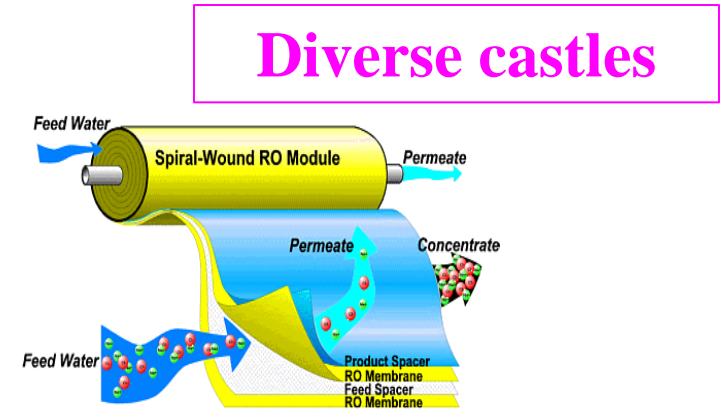
# Where are Biofilms?



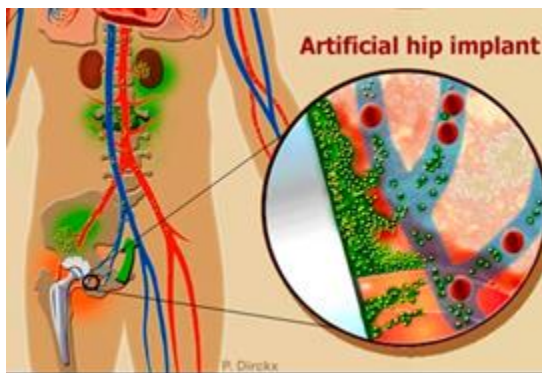
Dental plaque



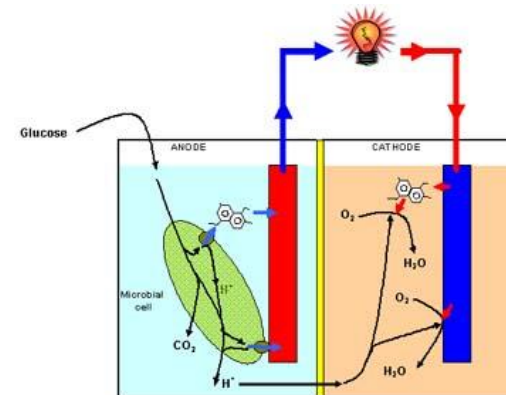
Wound biofilm



Reverse osmosis membrane



Biofilm on medical devices



Microbial fuel cell

99% of bacteria grow as biofilms. Biofilms are important for **medicine (80% of bacterial infections occur in biofilms)** and **engineering (corrosion and bioreactors)**.

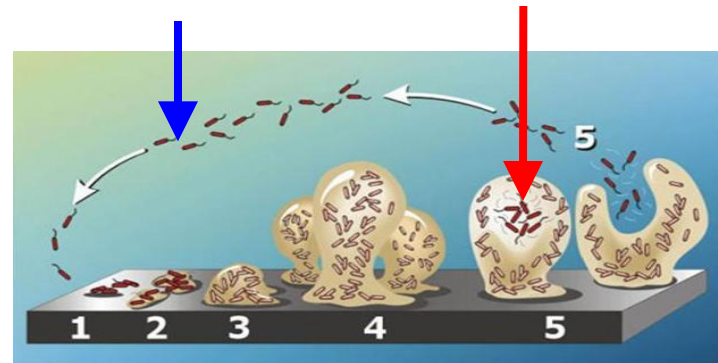


# Issue of Biofilm

- **Biofilm cells can be up to 1000 times more resistant to antibiotic treatment than planktonic (free-floating single cells).**
- **Objectives in biofilm research: to discover new biofilm inhibitors and to understand its genetic mechanism.**
- **More than 5000 biofilm papers from PubMed in 2016.**

**Very secure castle !**

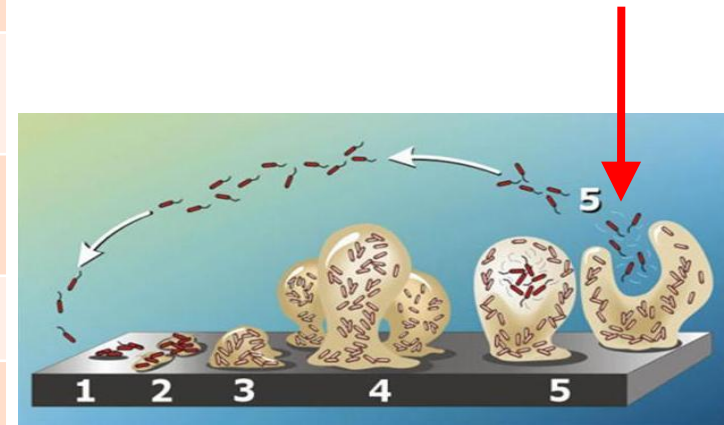
**Planktonic cells** : easy to kill      **Biofilm cells** : antibiotic resistant



# Methods of Biofilm Reduction

Agents	Target and mechanisms
Protease, DNase, glucosaminidase, chitinase, dispersin, etc	Matrix-targeting enzymes
QS modulators	Interference of quorum-sensing (QS)
Small natural molecules	D-amino acids, <i>cis</i> -2-decanoic acid, indole, etc
Nitric oxide, DSF	Controlling c-di-GMP
Nutrients (increase or limitation), oxygen, iron, etc	Controlling c-di-GMP
Plant derived compounds (furanons, indole derivatives, flavonoids, etc)	Plant's defense system
Bacterial derived compounds	Possible weapon in bacterial competition
Artificial chemicals or polymers	Surface modification

## Biofilm reduction and its mechanism





# Biofilm vs. Planktonic in *E. coli*

biofilm cells



DNA Microarray

Comparison of 4,000 genes



planktonic cells

Gene	b#	Expression ratio	Function of the protein
<i>hha</i>	b0460	+25	haemolysin modulating protein
<i>ybaJ</i>	b0461	+36	unknown
<i>yliH</i>	b0836	+3	unknown
<i>yceP</i>	b1060	+4	unknown
<i>ycfR</i>	b1112	+12	stress response protein
<i>ydgG</i>	b1601	+6	AI-2 transporter
<i>b2377</i>	b2377	+12	unknown
<i>b3022</i>	b3022	+8	unknown
<i>hslS</i>	b3686	+32	heat shock protein
<i>hslT</i>	b3687	+4	heat shock protein
<i>soxS</i>	b4062	+49	regulation of superoxide response regulator

My observation : pink mutants produced less indole (↓) and formed more biofilm (↑).

# *E. coli* O157:H7

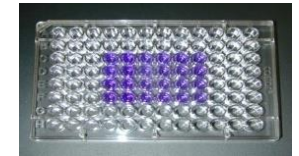
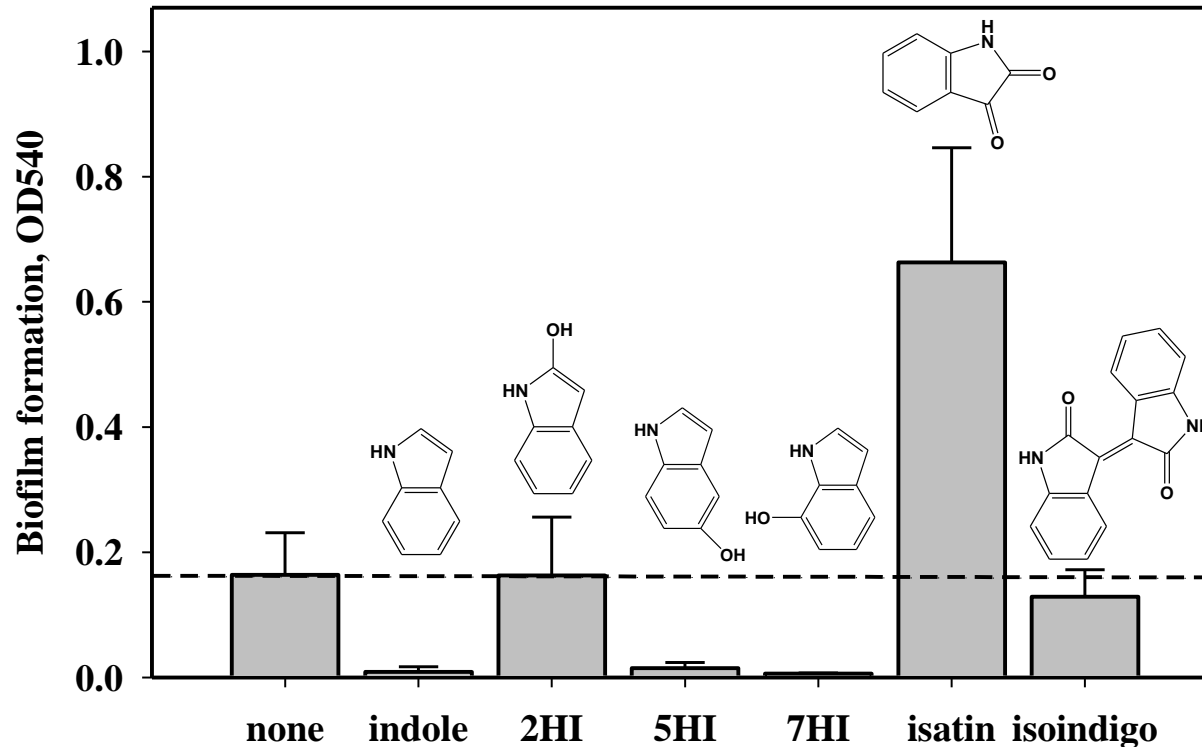
- Enterohemorrhagic (**EHEC**) *E. coli* - food borne pathogen that causes **bloody diarrhea** and hemorrhagic colitis. Approximately 70,000 infections (USA) and 2,100 hospitalizations with a cost of \$405 million (2003 value).
- No treatment currently available.
- *E. coli* O157:H7 makes strong biofilm on any surface.



# Indole and Hydroxyindoles

## Control EHEC Biofilm

Sophistication of words

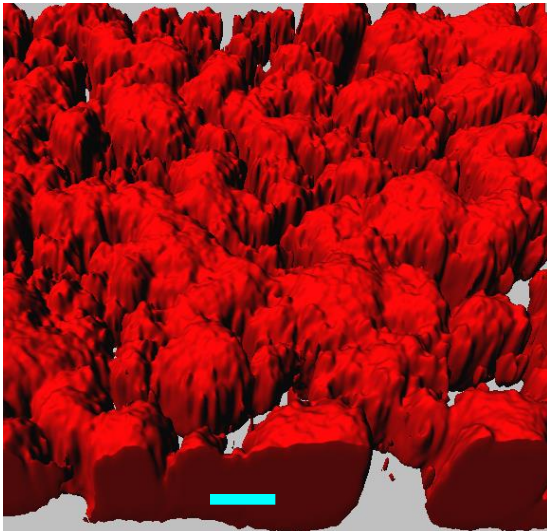


**Result:** Indole, 5-HI, and 7-HI decrease biofilm formation at 30°C for 7 hrs.  
Oxidation position is important for controlling biofilm formation.

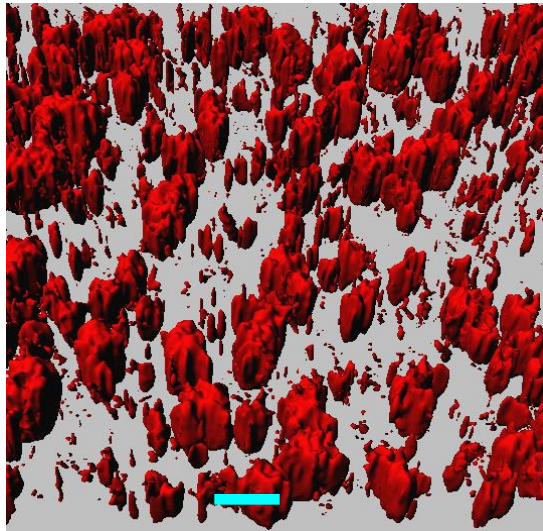
# In Continuous Flow Chamber



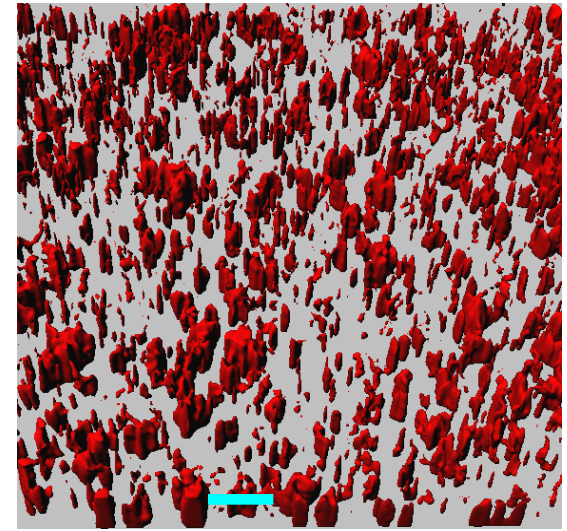
None



1.0 mM indole



1.0 mM 7-hydroxyindole



*E. coli* O157 biofilm formation in LB at 30°C after 24 hr in continuous flow chambers. Scale bar indicates 20  $\mu\text{m}$ .

**Results:** Confirmed the results in continuous flow chambers.



# Indole Controls Many Genes



Gene	b #	Fold-change <i>yceP</i> + indole	Fold change $\Delta$ <i>trpE</i>	Fold change $\Delta$ <i>tnaA</i>	Description
<b>Transcription</b>					
<i>sdiA</i>	<i>b1916</i>	2.9	1.0*	1.2	transcriptional activator for cell division ( <b>LuxR/UhpA family</b> )
<i>soxS</i>	<i>b4062</i>	2.3	2.8	-1.2	regulation of superoxide response regulon
<i>cspA</i>	<i>b3556</i>	1.8	-4.6	-1.2*	cold shock protein 7.4, transcriptional activator of hns
<b>Cell motility and secretion</b>					
<i>hha</i>	<i>b0460</i>	4.9	-1.4	-1.4	haemolysin expression modulating protein
<i>ybaJ</i>	<i>b0461</i>	5.2	-1.1*	1.0*	conserved hypothetical protein
<i>mqsR</i>	<i>b3022</i>	2.8	1.1*	1.4	master regulator of chemotaxis via AI-2; interacts with QseB
<b>Colanic acid synthesis genes</b>					
<i>ypdI</i>	<i>b2376</i>	1.1*	1.3*	-2.5	YpdI colanic acid synthesis lipoprotein
<b>Acid resistance</b>					
<i>gadE</i>	<i>b3512</i>	-5.3	-1.6	1.1*	activator acid resistance genes and putative LuxR transcriptional activator
<i>gadA</i>	<i>b3517</i>	-4.9	-2.3	1.2*	glutamate decarboxylase A, isozyme, PLP-dependent
<i>gadB</i>	<i>b1493</i>	-3.3	-2.6	1.2*	glutamate decarboxylase isozyme
<i>gadC</i>	<i>b1492</i>	-3.7	-1.6	1.2*	acid sensitivity protein, putative transporter
<i>hdeA</i>	<i>b3510</i>	-4.6	-2.3	1.0*	periplasmic chaperone of acid-denatured protein
<i>hdeB</i>	<i>b3509</i>	-4.3	-2.3	1.2*	10K-L protein, periplasmic protein related to acid resistance protein
<i>hdeD</i>	<i>b3511</i>	-2.6	-1.6	-1.5	protein involved in acid resistance
<i>ymgC</i>	<i>b1167</i>	-2.3	1.2*	1.5	hypothetical protein
<i>ymgA</i>	<i>b1165</i>	-3.3	1.1	1.7	hypothetical protein
<i>ymgB</i>	<i>b1166</i>	-6.1	1.2	1.5	hypothetical protein
<b>Phage-related genes</b>					
<i>ypjF</i>	<i>b2646</i>	1.2*	1.2*	-12.1	CP4-57 prophage
<i>ymfI</i>	<i>b1143</i>	1.0*	1.1*	-5.7	e14 prophage
<i>ydaY</i>	<i>b1366</i>	1.0*	1.0*	-4.6	Rac prophage

**Result:** Indole influences gene expression of cell motility and the acid resistance genes.

Lee, Jayaraman, and Wood, *BMC Microbiology* (2007, >290 citations)

# Indole against Other Species

## Defense against other species

**Hypothesis: indole as a defense system in multispecies nature.**

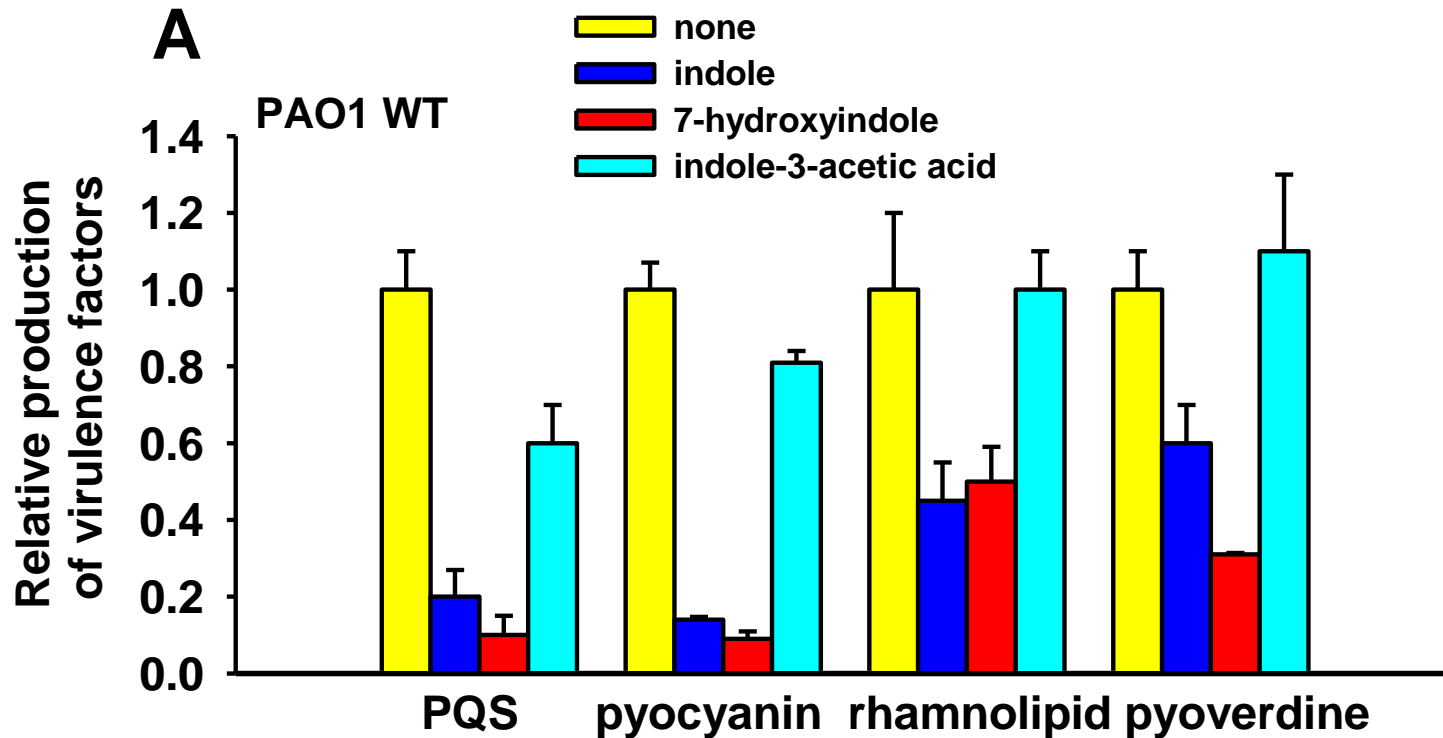
- System: pathogenic *Pseudomonas aeruginosa* + Indole



- *P. aeruginosa* is a notorious human pathogen that can cause chronic infection in the lungs, ears, wounds, and burns. It easily acquires antibiotic resistance. **No production of indole**
- *P. aeruginosa* produces various toxic chemicals for human and other bacteria.

# Reduction of Toxins by Indole

## Defense tools



**Result:** indole and 7-HI decreased the production of virulence factors without affecting cell growth. Maybe, *E. coli* utilizes indole to compete against *P. aeruginosa*.

# Mechanism of Indole in *P. aeruginosa*



PA#	Gene	Fold change <sup>1</sup>		Description
		indole vs. no indole	7HI vs. no 7HI	
<b>Multidrug efflux transporter</b>				
PA4205	<i>mexG</i>	-13.0	-7.0	Hypothetical protein, part of the <i>mexGHI-opmD</i> cluster
PA4206	<i>mexH</i>	-11.3	-5.7	Efflux membrane fusion protein precursor
PA4207	<i>mexI</i>	-6.1	-4.0	Efflux transporter
PA4208	<i>opmD</i>	-7.5	-4.6	Probable outer membrane efflux protein
<b>Phenazine synthesis</b>				
PA1902	<i>phzD2</i>	-3.2	-3.0	Phenazine biosynthesis protein PhzD
PA1903	<i>phzE2</i>	-3.5	-2.6	Phenazine biosynthesis protein PhzE
PA1904	<i>phzF2</i>	-4.0	-4.3	Phenazine biosynthesis protein
PA1905	<i>phzG2</i>	-3.2	-3.2	Pyridoxamine 5'-phosphate oxidase
PA2274		-4.9	-3.2	Phenazine biosynthesis protein
PA4210	<i>phzA1</i>	-2.5	-3.7	Phenazine biosynthesis protein
PA4217	<i>phzS</i>	-2.8	-2.8	Flavin-containing monooxygenase
<b>PQS synthesis</b>				
PA0996	<i>pqsA</i>	-1.9	-2.0	PqsA, probable coenzyme A ligase
PA0997	<i>pqsB</i>	-2.3	-2.5	PqsB,
PA1000	<i>pqsE</i>	-2.0	-2.1	PqsE, quinolone signal response protein
PA1003	<i>mvrR</i>	1.3	-1.1	MvR (PqsR), PQS transcriptional regulator
<b>Pyoverdine synthesis</b>				
PA2393	<i>pvdM</i>	-2.5	-1.1	pyoverdine biosynthesis protein PvdM
PA2394	<i>pvdN</i>	-2.5	-1.2	pyoverdine biosynthesis protein PvdN
PA2424	<i>pvdL</i>	-2.3	-1.3	pyoverdine biosynthesis protein PvdV
PA2426	<i>pvdS</i>	-6.1	-2.6	Sigma factor PvdS, transcriptional regulator
<b>Metabolism</b>				
PA0283	<i>sbp</i>	-8.6	-1.7	Sulfate-binding protein precursor
PA1838	<i>cysI</i>	-6.1	-2.1	Sulfite reductase
PA3441	<i>ssuF</i>	-7.5	-5.7	SsuF, part of the <i>ssu</i> locus,
PA3442	<i>ssuB</i>	-4.6	-2.6	SsuB, part of the <i>ssu</i> locus
PA3443	<i>ssuC</i>	-3.2	-2.6	SsuC, part of the <i>ssu</i> locus
PA3444	<i>ssuD</i>	-6.1	-2.1	SsuD, part of the <i>ssu</i> locus
PA3445	<i>ssuA</i>	-6.5	-2.8	SsuA, part of the <i>ssu</i> locus
PA3446	<i>ssuE</i>	-6.5	-1.6	SsuE, part of the <i>ssu</i> locus
PA4442	<i>cysN</i>	-4.6	-2.0	ATP sulfurylase GTP-binding subunit
PA4443	<i>cysD</i>	-4.9	-1.7	ATP sulfurylase small subunit
PA5025	<i>metY</i>	-2.6	-1.3	Homocysteine synthase
PA5427	<i>adhA</i>	2.1	2.6	Alcohol dehydrogenase
<b>Motility and TypeIV pilus</b>				
PA2131		-1.9	-2.6	Probable pili assembly chaperone
PA2570	<i>paIL</i>	1.6	-2.8	PA-I galactophilic lectin
PA4084	<i>cupB3</i>	-2.8	-4.0	Probable fimbrial biogenesis usher protein
PA4085	<i>cupB2</i>	-3.0	-2.1	Probable pili assembly chaperone
PA4297	<i>tadG</i>	2.3	3.5	TadG
PA4299	<i>tadD</i>	1.7	-2.6	Flp pilus assembly protein TadD
PA4300	<i>tadC</i>	1.7	-2.3	Flp pilus assembly protein TadC
PA4301	<i>tadB</i>	1.6	-3.0	Flp pilus assembly protein TadB
PA4302	<i>tadA</i>	1.9	-2.5	Flp pilus assembly protein, ATPase CpaF
PA4303	<i>tadZ</i>	1.7	-2.3	Flp pilus assembly protein, ATPase CpaE
PA4304	<i>repA</i>	2.0	-1.9	Flp pilus assembly protein, secretin CpaC
PA4651		2.0	-3.5	Probable pili assembly chaperone
<b>Transport of small molecules</b>				
PA1108		-1.9	-5.7	Probable MFS transporter
PA2092		-2.6	-4.0	Probable MFS transporter
PA2204		-1.9	-9.8	Binding protein component of ABC transporter
PA2328		-6.1	-5.3	Hypothetical protein in the cluster of ABC transporter
PA2329		-5.7	-5.7	Probable ATP-binding component of ABC transporter
PA2330		-4.9	-4.3	Hypothetical protein in the cluster of ABC transporter
PA2331		-5.7	-4.0	Hypothetical protein in the cluster of ABC transporter
PA3531	<i>bfrB</i>	3.2	2.3	Bacterioferritin, transport of small molecules
PA3926		-1.7	-2.0	Probable MFS transporter
PA4514	<i>piuA</i>	-4.3	-2.5	Probable outer membrane receptor for iron transport
<b>Oxygenases</b>				
PA2512	<i>antA</i>	5.3	-2.0	Anthraniolate dioxygenase large subunit
PA2513	<i>antB</i>	6.1	-2.6	Anthraniolate dioxygenase small subunit
PA2514	<i>antC</i>	3.0	-2.8	Anthraniolate dioxygenase reductase
PA0106	<i>coxA</i>	3.2	1.4	Cytochrome c oxidase, subunit I
PA0107		2.8	1.3	Predicted cytochrome oxidase assembly factor
PA0111		3.0	1.3	Hypothetical protein in the cluster of oxidase
PA0112		3.0	1.1	Hypothetical protein in the cluster of oxidase
PA0113		3.0	1.4	Probable cytochrome c oxidase assembly
<b>Others</b>				
PA0716		5.7	7.0	Probable bacteriophage protein
PA0740	<i>sdsA1</i>	-8.6	-3.0	Probable beta-lactamase
PA1109		-2.6	-5.7	Transcriptional regulator
PA3234	<i>yjeG</i>	4.3	2.0	Probable sodium:solute symporter
PA3337	<i>rfaD</i>	2.0	2.5	ADP-L-glycero-D-mannoheptose 6-epimerase
<b>Hypothetical proteins</b>				
PA0284		-13.9	-1.6	Hypothetical protein, induced by MvR
PA0492		-4.3	-3.5	Hypothetical protein
PA0696		-4.3	-2.5	Hypothetical protein
PA0939		-1.1	-5.3	Hypothetical protein
PA1190		5.7	1.6	Hypothetical protein
PA1837		-4.6	-2.1	Hypothetical protein
PA1914		1.1	-6.1	Hypothetical protein
PA1953		-3.2	-6.1	Hypothetical protein
PA2036		-2.0	-7.0	Hypothetical protein
PA2078		-3.0	-5.7	Hypothetical protein
PA2419		-1.6	6.1	Hypothetical protein
PA3235		4.0	2.5	Hypothetical protein
PA3237		-1.9	-6.1	Hypothetical protein
PA3719		4.6	1.3	Hypothetical protein
PA3931		-5.3	-1.7	Hypothetical protein

**Result: Indole and 7-HI prevent bacterial communication of *P. aeruginosa*.  
quorum quenching!**

# Inhibition of Swarming (not Growth) by Indole and 7-HI

none

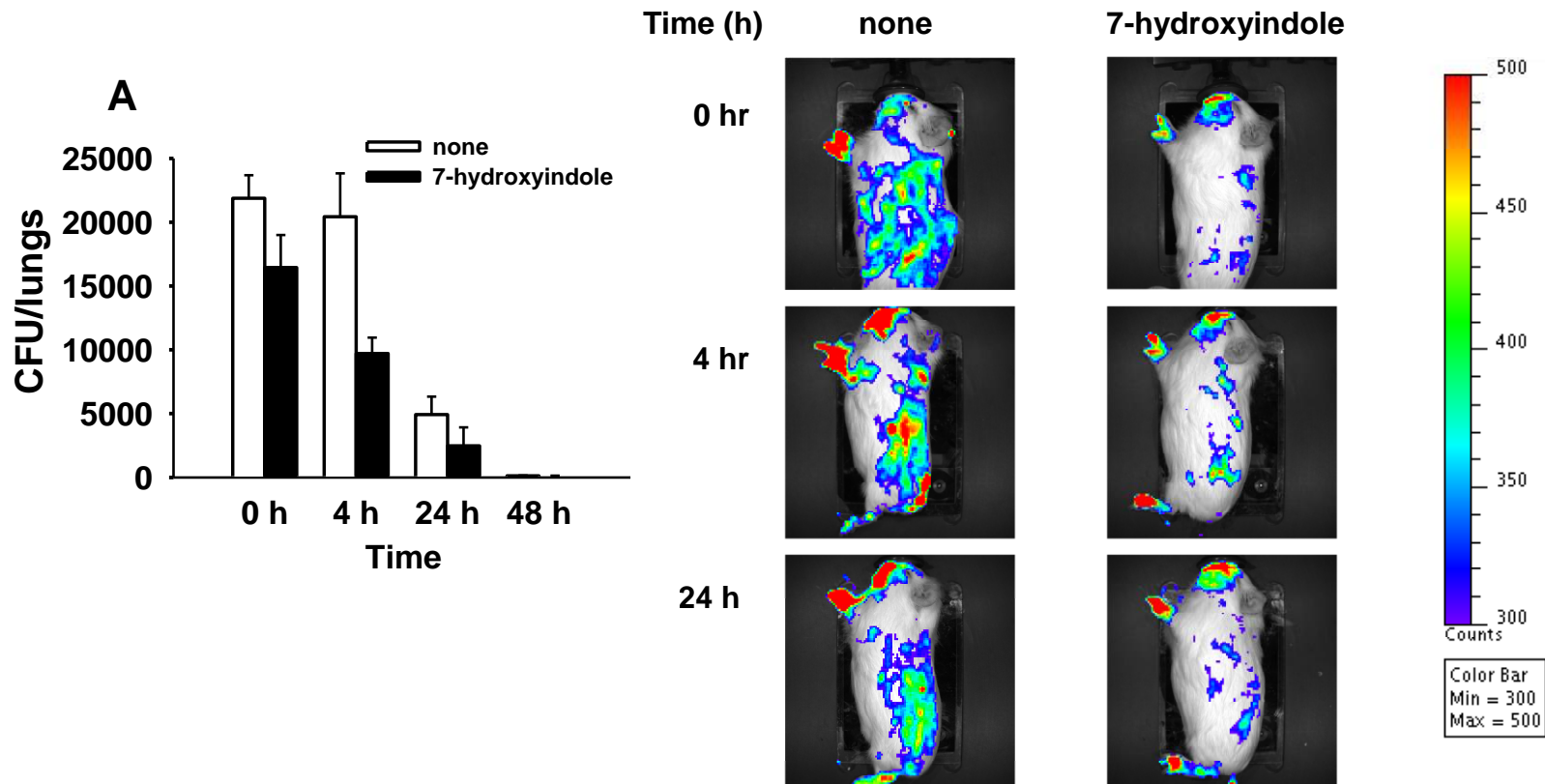
indole

7-hydroxyindole



**Result:** Indole decreases moving and 7-HI stops moving of *P. aeruginosa*.

# Animal Study (Guinea Pigs)

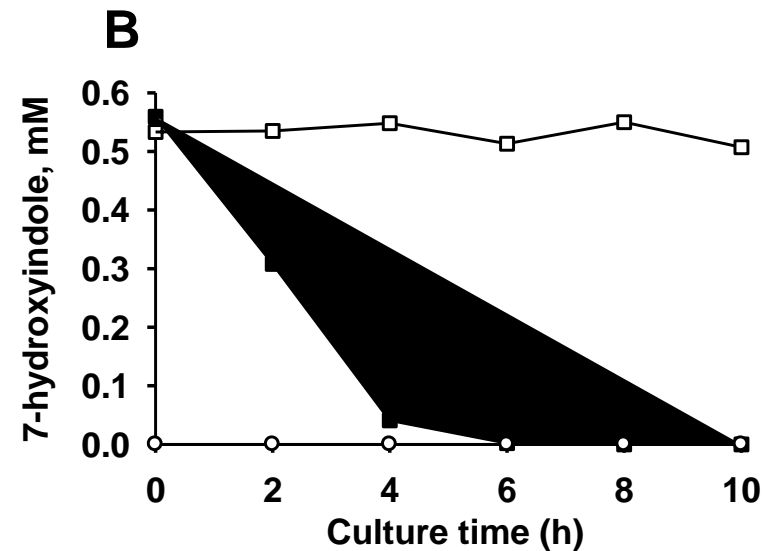
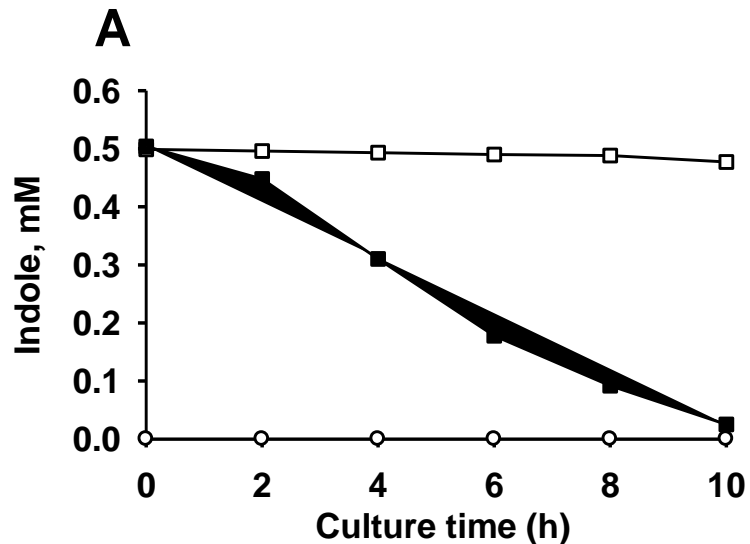


**Result:** 7-HI reduced attachment of *P. aeruginosa* in guinea pigs. We found a potential drug.



# Problem of Indole and 7-HI: Degradation

## Spear and shield

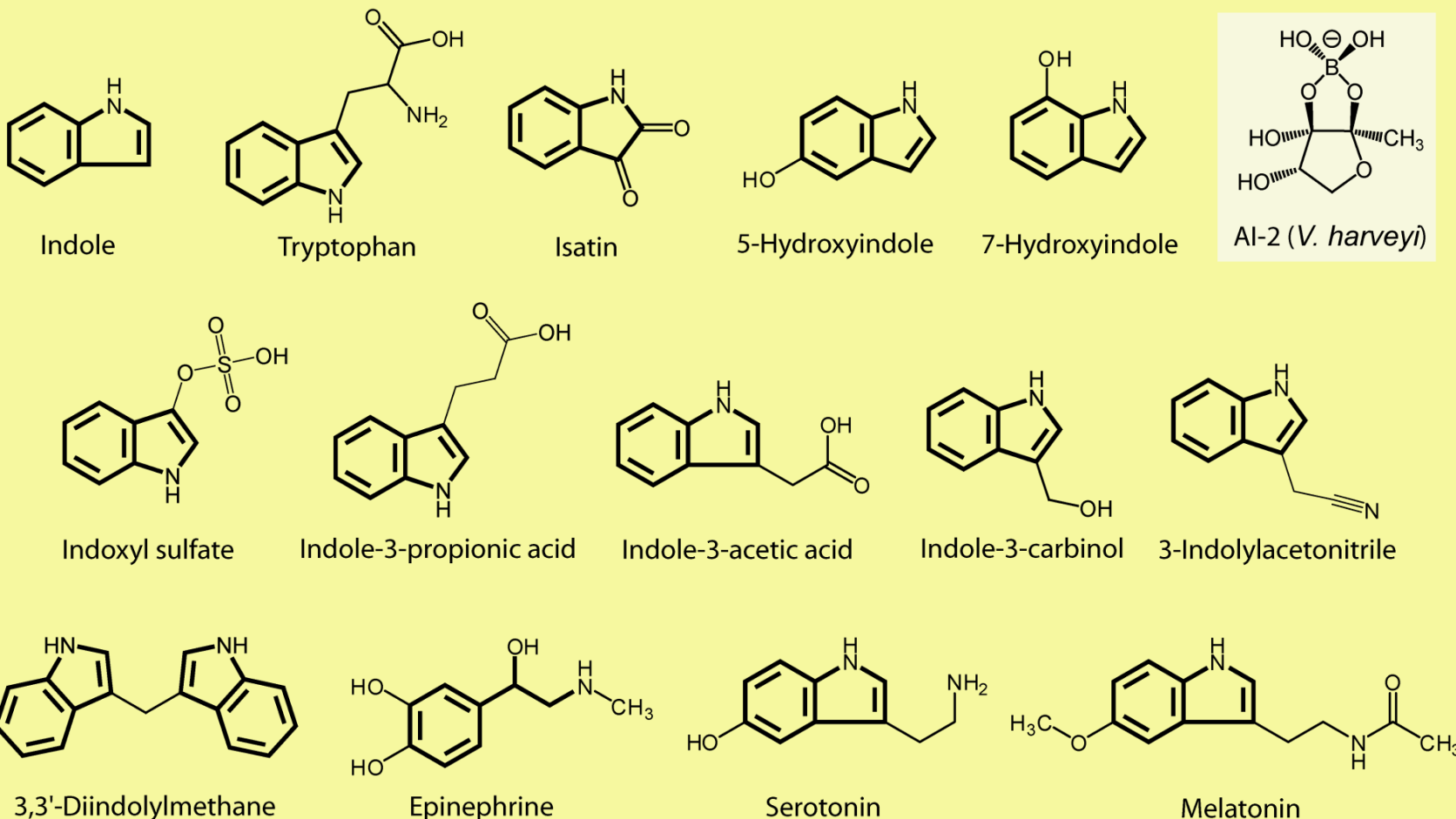


Degradation of **indole (A)** and **7HI (B)** in LB at 37°C. **Closed square data (■)** are from live cells.

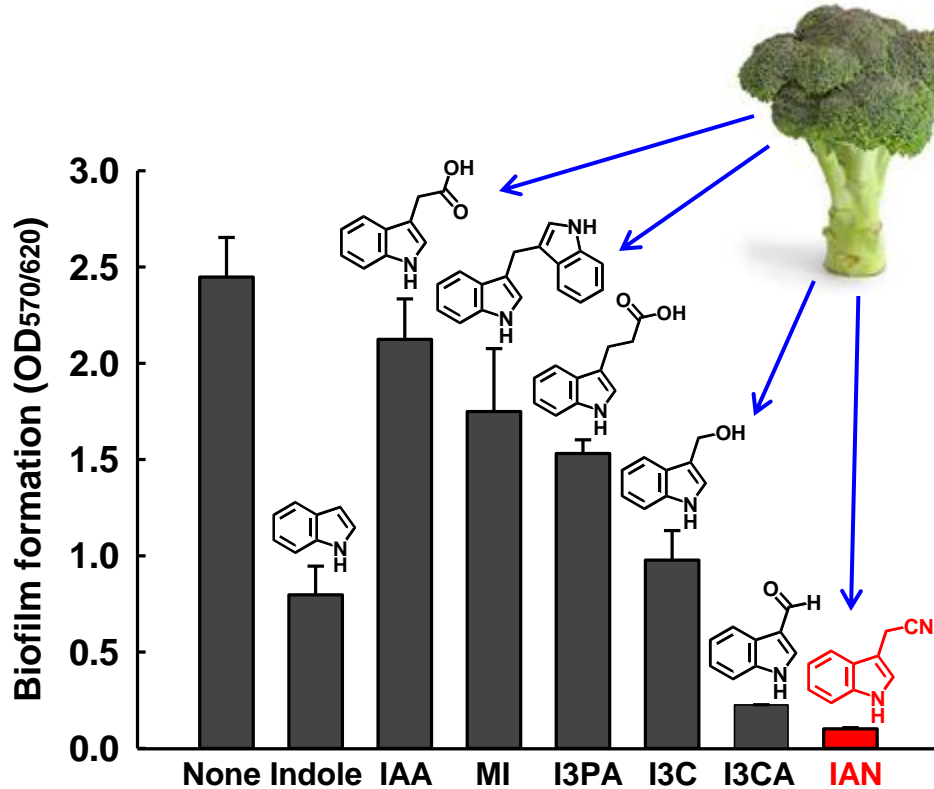
**Result:** As a defense, *P. aeruginosa* degrades indole and 7-HI.

# Natural Indole Derivatives

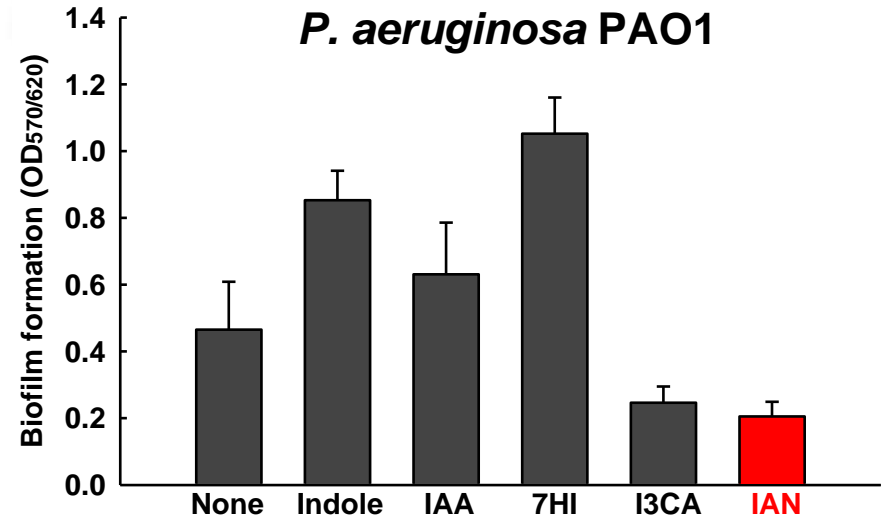
Indole motifs are in bold.



# Broccoli Contains Biofilm Inhibitors



Biofilm formation of *E. coli* O157:H7.  
No growth inhibition.  
**IAN is a plant auxin, like IAA.**

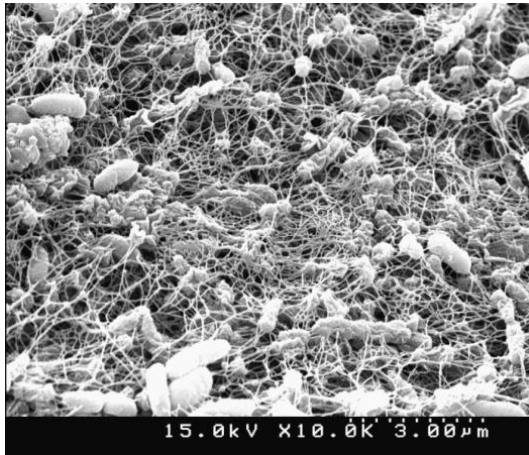


Biofilm formation of *P. aeruginosa*.  
No growth inhibition.

# Mechanism of IAN: Polymeric Matrix (Curli)

No tie → weak group

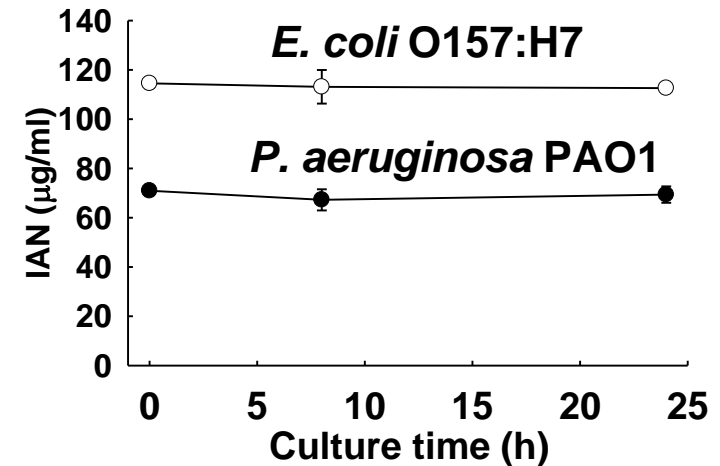
## *E. coli* O157:H7



None



IAN



Found a **stable and non-toxic biofilm inhibitor** against *E. coli* O157:H7

Lee, Cho, and J. Lee, *Environmental Microbiology* (2011)

# Indoles Affect *Staphylococcus aureus* Virulence (Golden Carotenoid Staphyloxanthin)

## A Cholesterol Biosynthesis Inhibitor Blocks *Staphylococcus aureus* Virulence

Chia-I Liu,<sup>1,2,3\*</sup> George Y. Liu,<sup>4\*</sup> Yongcheng Song,<sup>5\*</sup> Fenglin Yin,<sup>6</sup> Mary E. Hensler,<sup>7</sup> Wen-Yih Jeng,<sup>1,2</sup> Victor Nizet,<sup>7,8†</sup> Andrew H.-J. Wang,<sup>1,2,3,†</sup> Eric Oldfield<sup>5,6†</sup>

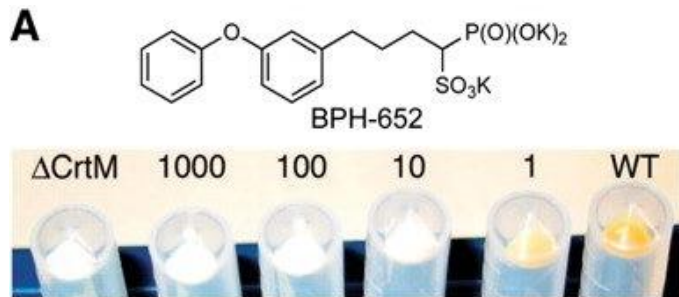
*Staphylococcus aureus* produces hospital- and community-acquired infections, with methicillin-resistant *S. aureus* posing a serious public health threat. The golden carotenoid pigment of *S. aureus*, staphyloxanthin, promotes resistance to reactive oxygen species and host neutrophil-based killing, and early enzymatic steps in staphyloxanthin production resemble those for cholesterol biosynthesis. We determined the crystal structures of *S. aureus* dehydrosqualene synthase (CrtM) at 1.58 angstrom resolution, finding structural similarity to human squalene synthase (SQS). We screened nine SQS inhibitors and determined the structures of three, bound to CrtM. One, previously tested for cholesterol-lowering activity in humans, blocked staphyloxanthin biosynthesis in vitro (median inhibitory concentration ~100 nM), resulting in colorless bacteria with increased susceptibility to killing by human blood and to innate immune clearance in a mouse infection model. This finding represents proof of principle for a virulence factor-based therapy against *S. aureus*.

(MRSA) Unfortunately, it has been a decades-long process to develop new antibiotics. One approach is to research specific molecules to re-innate

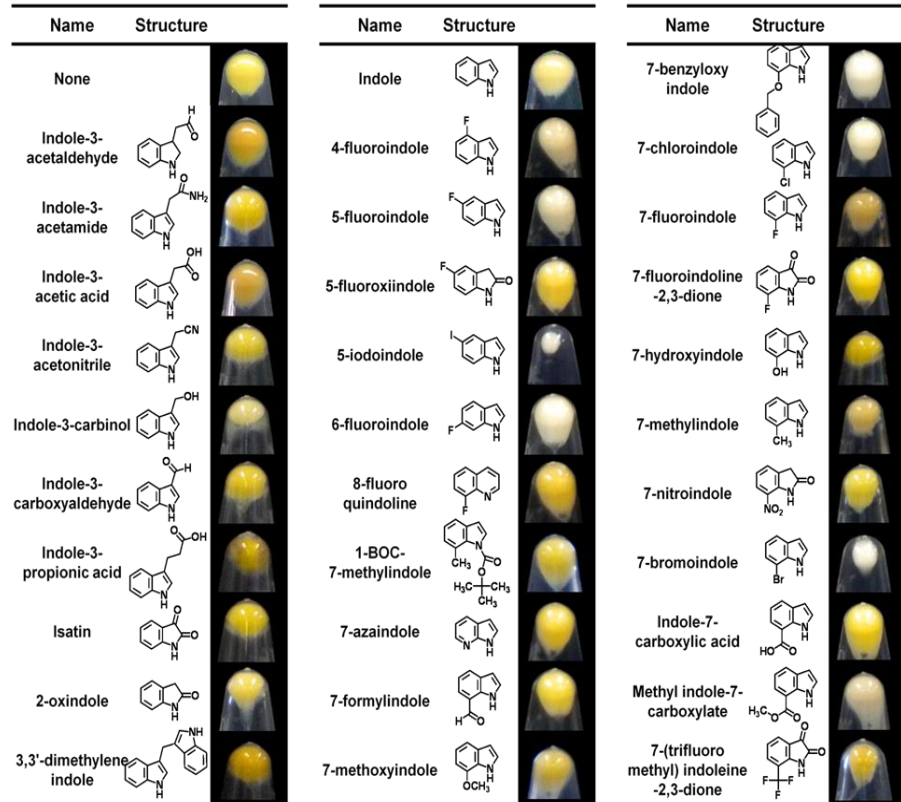
<sup>1</sup>Institute of Life Science, National Central University, Chungli, Taiwan 32001, Republic of China; <sup>2</sup>Department of Life Science, National Central University, Chungli, Taiwan 32001, Republic of China; <sup>3</sup>Department of Life Science, National Central University, Chungli, Taiwan 32001, Republic of China; <sup>4</sup>Department of Chemistry, Los Angeles Valley College, Van Nuys, California 91411, USA; <sup>5</sup>Department of Chemistry, University of Illinois, Urbana, Illinois 61801, USA; <sup>6</sup>Department of Chemistry, University of Illinois, Urbana, Illinois 61801, USA; <sup>7</sup>Department of Microbiology, University of Illinois, Urbana, Illinois 61801, USA; <sup>8</sup>Department of Microbiology, University of Illinois, Urbana, Illinois 61801, USA

\*These authors contributed equally to this work. †To whom all correspondence should be addressed. E-mail: wangahj@cc.nyu.edu; oldfield@uiuc.edu; vnizet@uiuc.edu

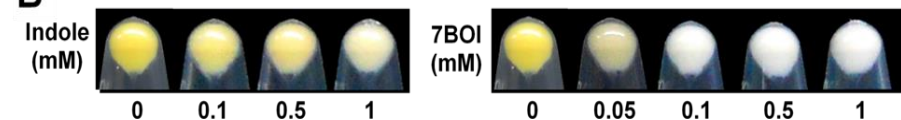
www.sciencemag.org SCIENCE VOL 319 7 MARCH 2008



## A



## B



Lee, Cho, Kim, Kim, Banskota, Cho, and J. Lee, *Applied Microbiology and Biotechnology* (2013)

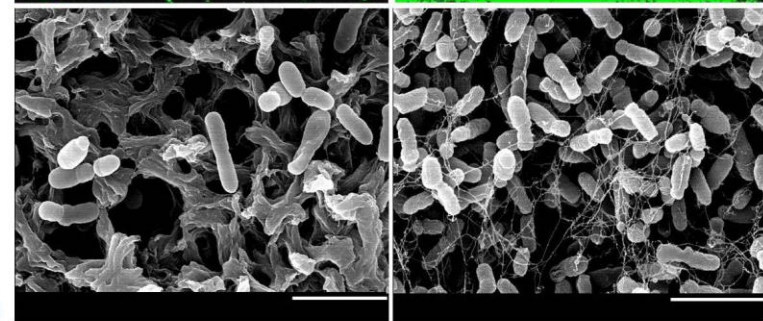
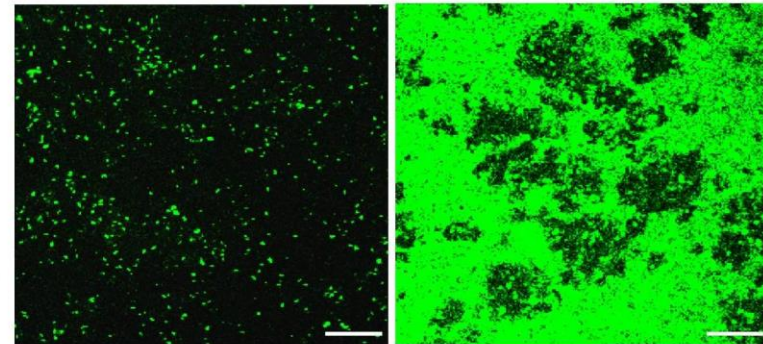
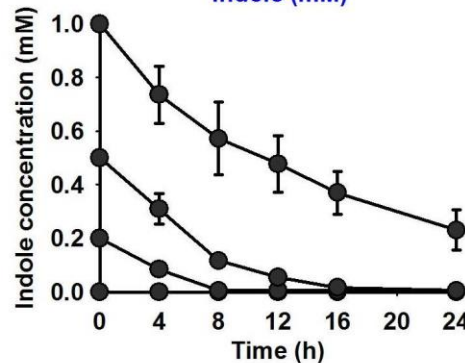
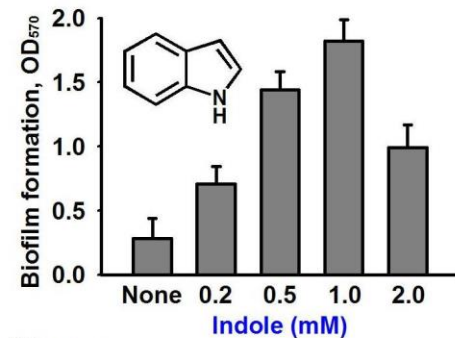


# Indole Increases Biofilm Formation of *Agrobacterium tumefaciens*



*A. tumefaciens* is a plant pathogen causing crown gall

## Roles of indole in *Agrobacterium tumefaciens*



None

Indole

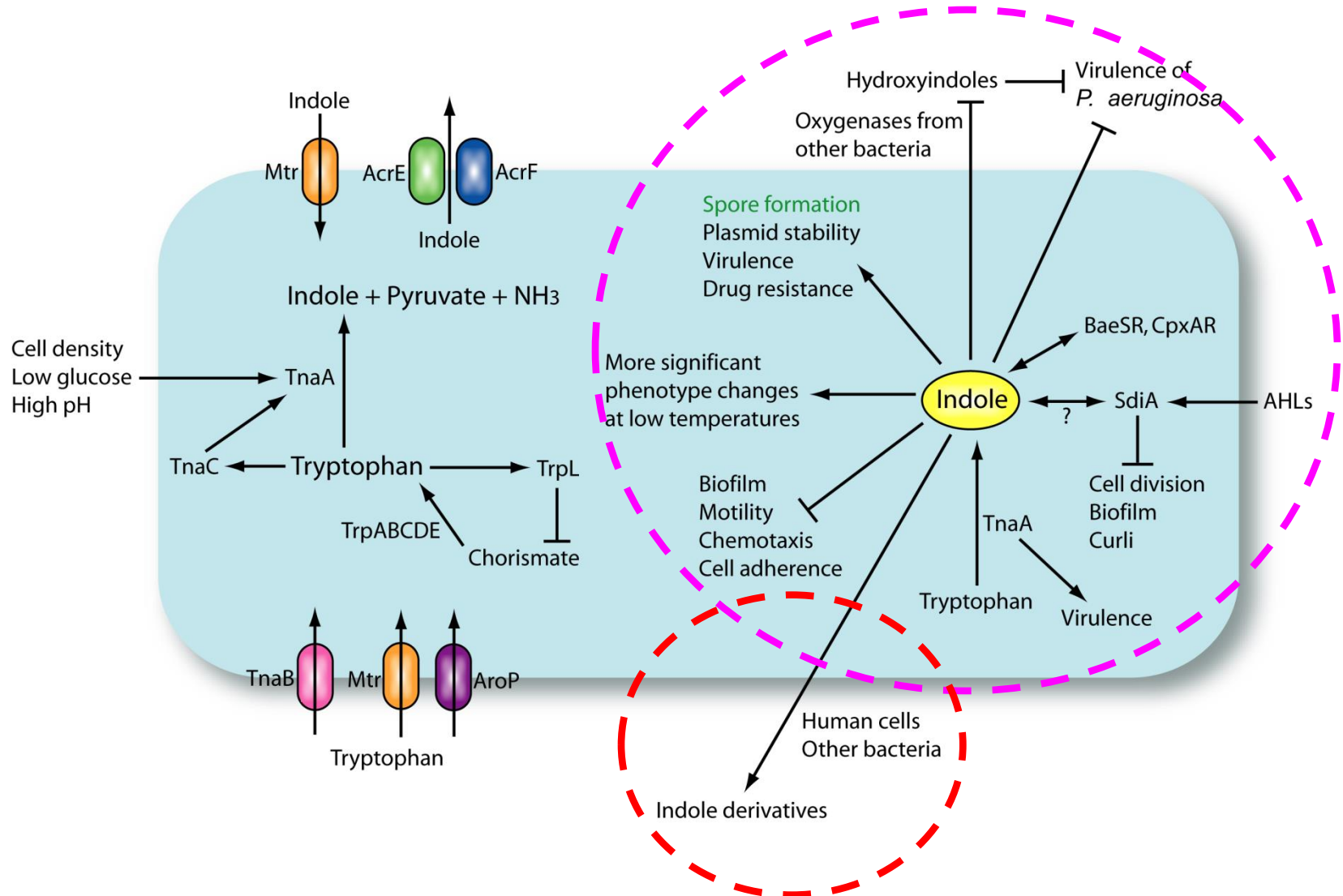
Lee, Kim, Baek, Cho, and J. Lee, *Environmental Microbiology* (2015)



# Effect of Indoles on Other Bacteria

Bacteria	Indole compound	Phenotypic change	Mechanism	Ref
<i>Acinetobacter baumannii</i>	Indole derived flustramine,	Inhibition of biofilm formation	Unknown	[46]
<i>Acinetobacter oleivorans</i>	Indole	Inhibition of biofilm formation and motility	Inhibition of QS regulator folding	[54]
<i>Bdellovibrio bacteriovorus</i>	Indole	Reduction of predation	Downregulation of flagellar and ribosome assembly genes	[60]
<i>Burkholderia unamae</i>	Indole and gallic acid	Induction of biofilm formation	Unknown	[59]
<i>Candida albicans</i>	Indole and 3-indolylacetonitrile	Reduction of biofilm formation and virulence	Stimulation of transcriptional factor	[56]
<i>Chromobacterium violaceum</i> <i>Pseudomonas chlororaphis</i> <i>Serratia marcescens</i>	Indole	Inhibition of QS-regulated pigmentation	QS inhibition	[55]
<i>Cylindrotheca sp.</i>	Indole and its derivatives	Inhibition of growth and biofilm formation	Induction of cellular Ca <sup>2+</sup> efflux	[66]
<i>Salmonella enterica serovar Typhimurium</i>	Indole	Increase of antibiotic tolerance	Oxidative stress response	[61]
<i>Salmonella enterica serovar Typhimurium</i>	Indole	Increase of antibiotic tolerance and decrease of motility	Activation of efflux pump and suppression of flagellar genes	[62,63]

# Indole in Microbial Community

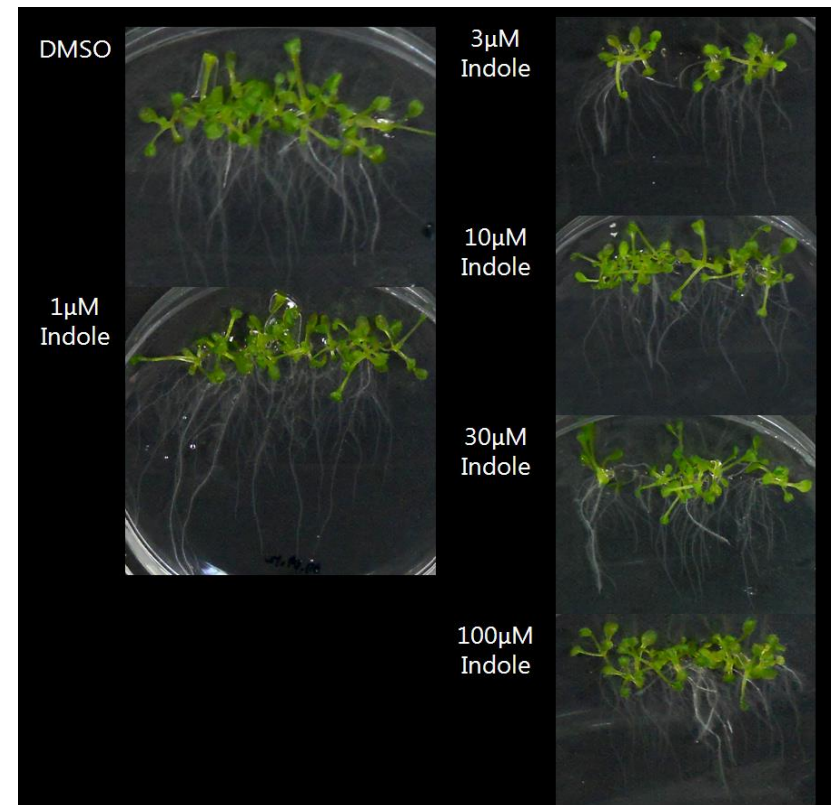
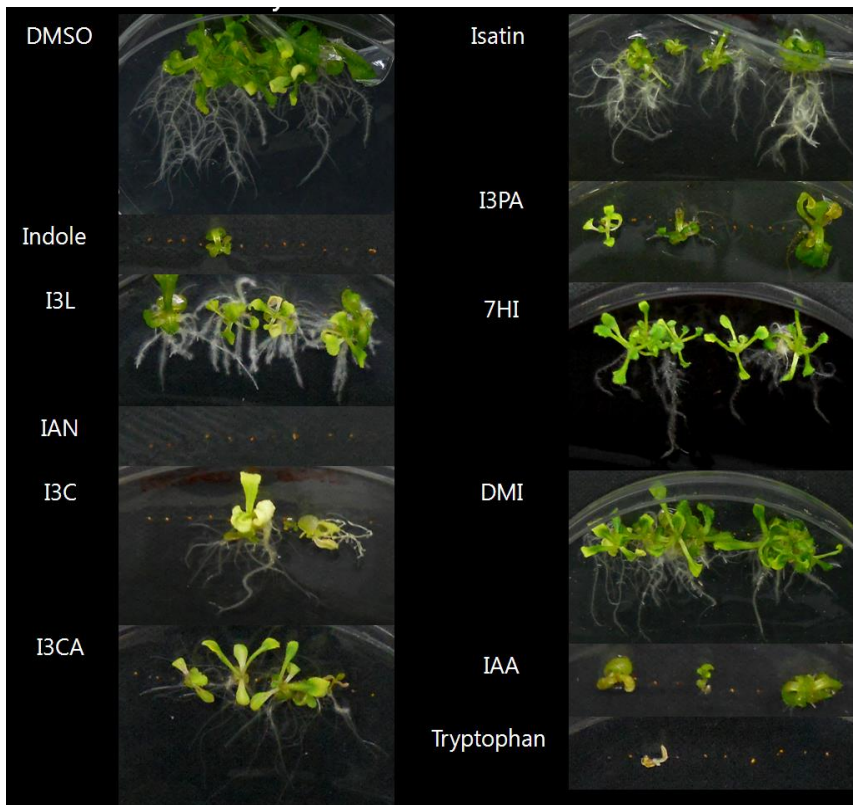




# Indoles on plants?

**Hypothesis: volatile indoles enhance plant growth and immunity.**

## *Arabidopsis thaliana*



**High concentration of indoles at 0.5 mM**

**Low concentration of indole**

**(Unpublished)**

# Functions of Indole in Animals?



**Germ-free** mouse  
(**no indole** in gut)



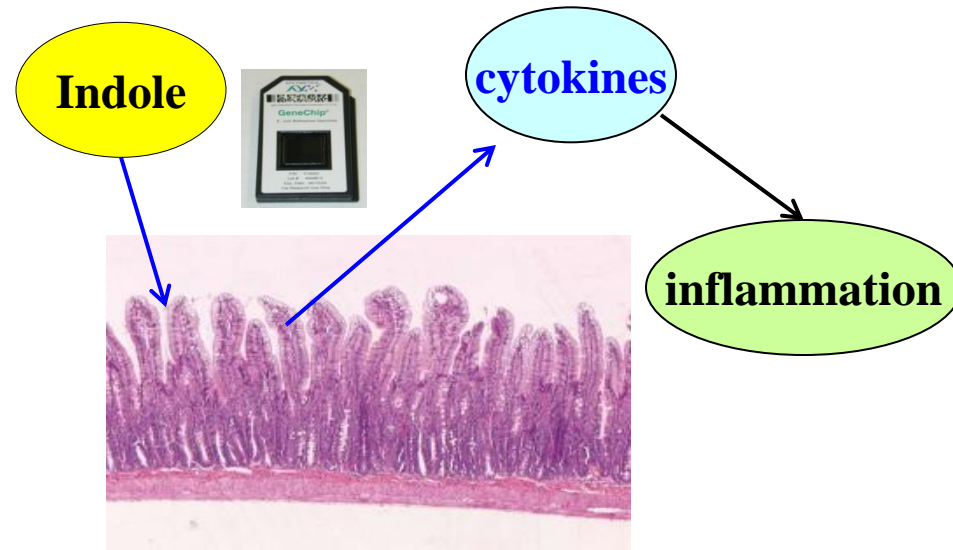
**regular** mouse  
(**indole** in gut)



**no indole-3-propionic**  
**acid in blood**

**indole-3-propionic**  
**acid in blood**

- **Indole-3-propionic acid**  
= a powerful antioxidant  
(Wikoff *et al.*, 2009, *PNAS*)



- (Wood *et al.*, 2010, *PNAS*)

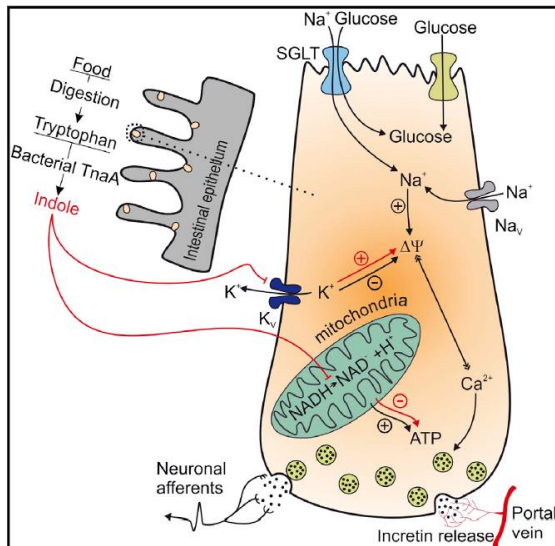


# Functions of Indole in Human?

## Cell Reports

### Bacterial Metabolite Indole Modulates Incretin Secretion from Intestinal Enteroendocrine L Cells

#### Graphical Abstract



#### Authors

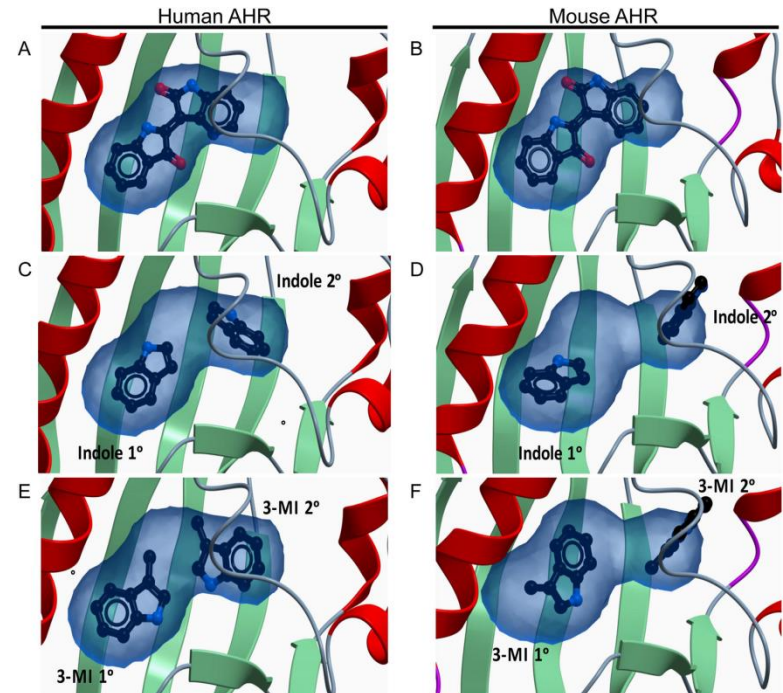
Catalin Chimerel, Edward Emery, ..., Fiona M. Gribble, Frank Reimann

#### Correspondence

cc539@cam.ac.uk (C.C.),  
fmg23@cam.ac.uk (F.M.G.),  
fr222@cam.ac.uk (F.R.)

#### In Brief

Indole is the main metabolite produced by gut bacteria from tryptophan. Chimerel et al. demonstrate that indole modulates the hormone secretion of enteroendocrine L cells and reveal the molecular mechanism behind this modulation. These findings suggest that the production of indole by bacteria could have a major impact on host metabolism.



SCIENTIFIC REPORTS

OPEN

Adaptation of the human aryl hydrocarbon receptor to sense microbiota-derived indoles

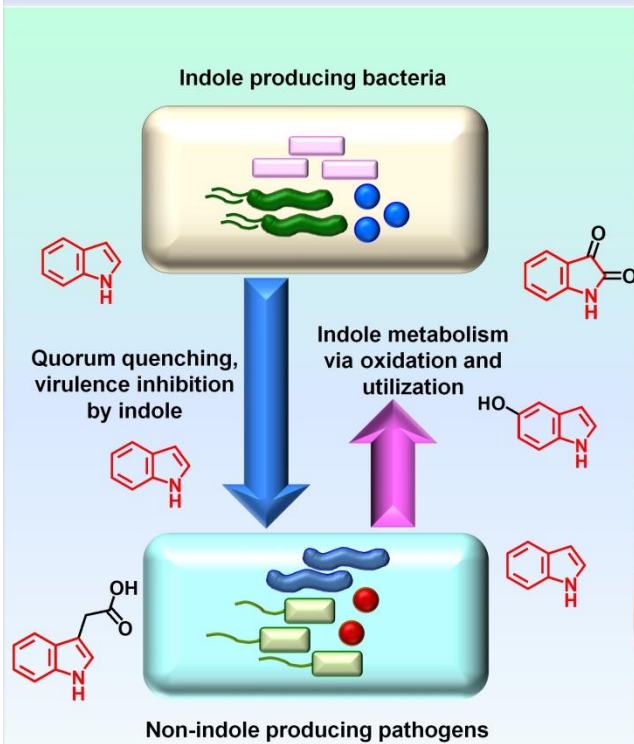
Received: 19 March 2015  
Accepted: 06 July 2015  
Published: 03 August 2015

Troy D. Hubbard<sup>1\*</sup>, Iain A. Murray<sup>1</sup>, William H. Bisson<sup>1</sup>, Tejas S. Lahoti<sup>2</sup>, Krishne Gowda<sup>2</sup>, Shantu G. Amin<sup>2</sup>, Andrew D. Patterson<sup>2</sup> & Gary H. Perdew<sup>2</sup>

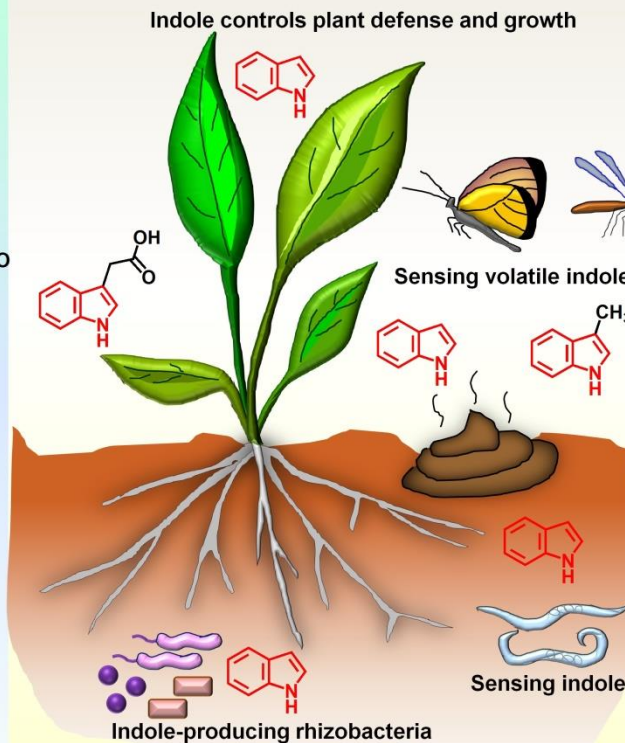


# Indole in Bacteria, Plants, and Insects, and Animals as a Signaling Molecule

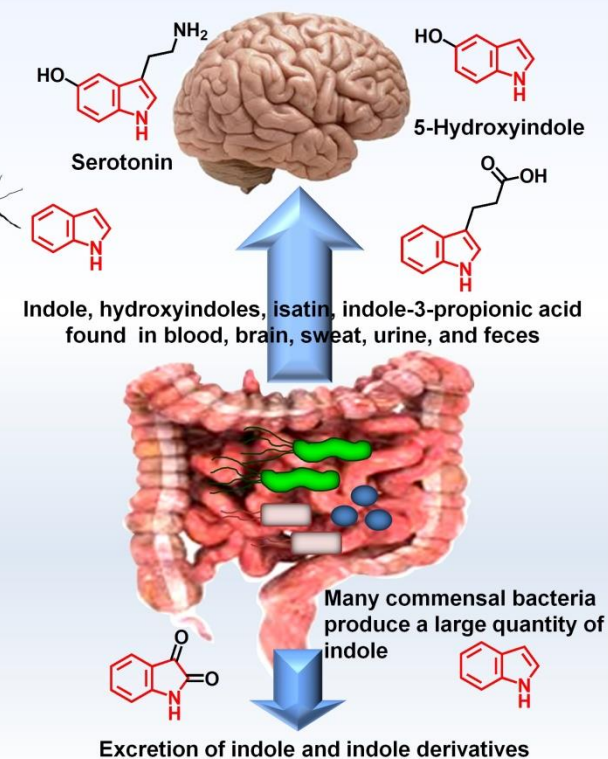
## Indoles in microbial community



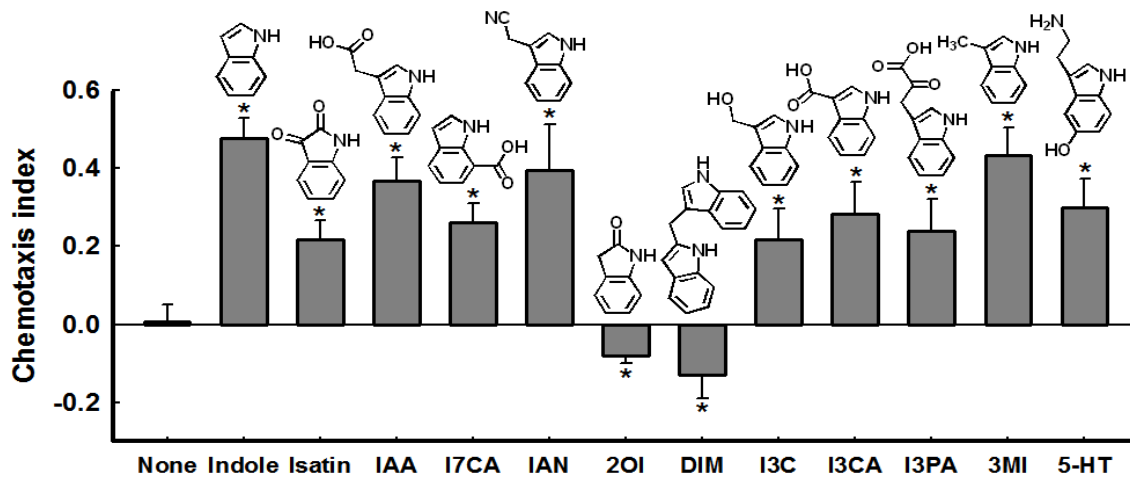
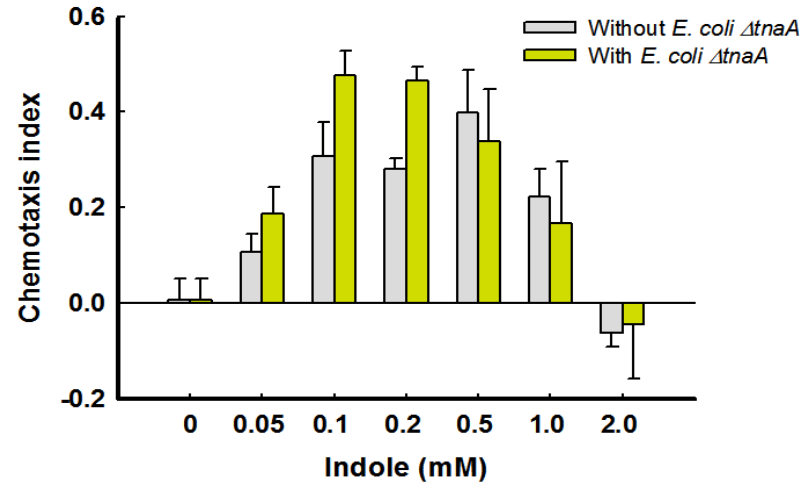
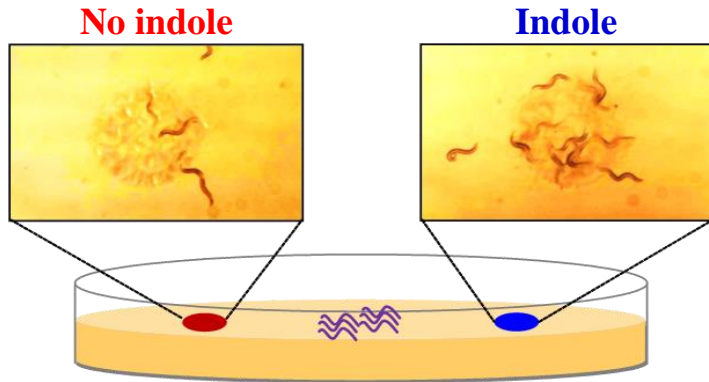
## Indoles in plants and insects



## Indoles in human

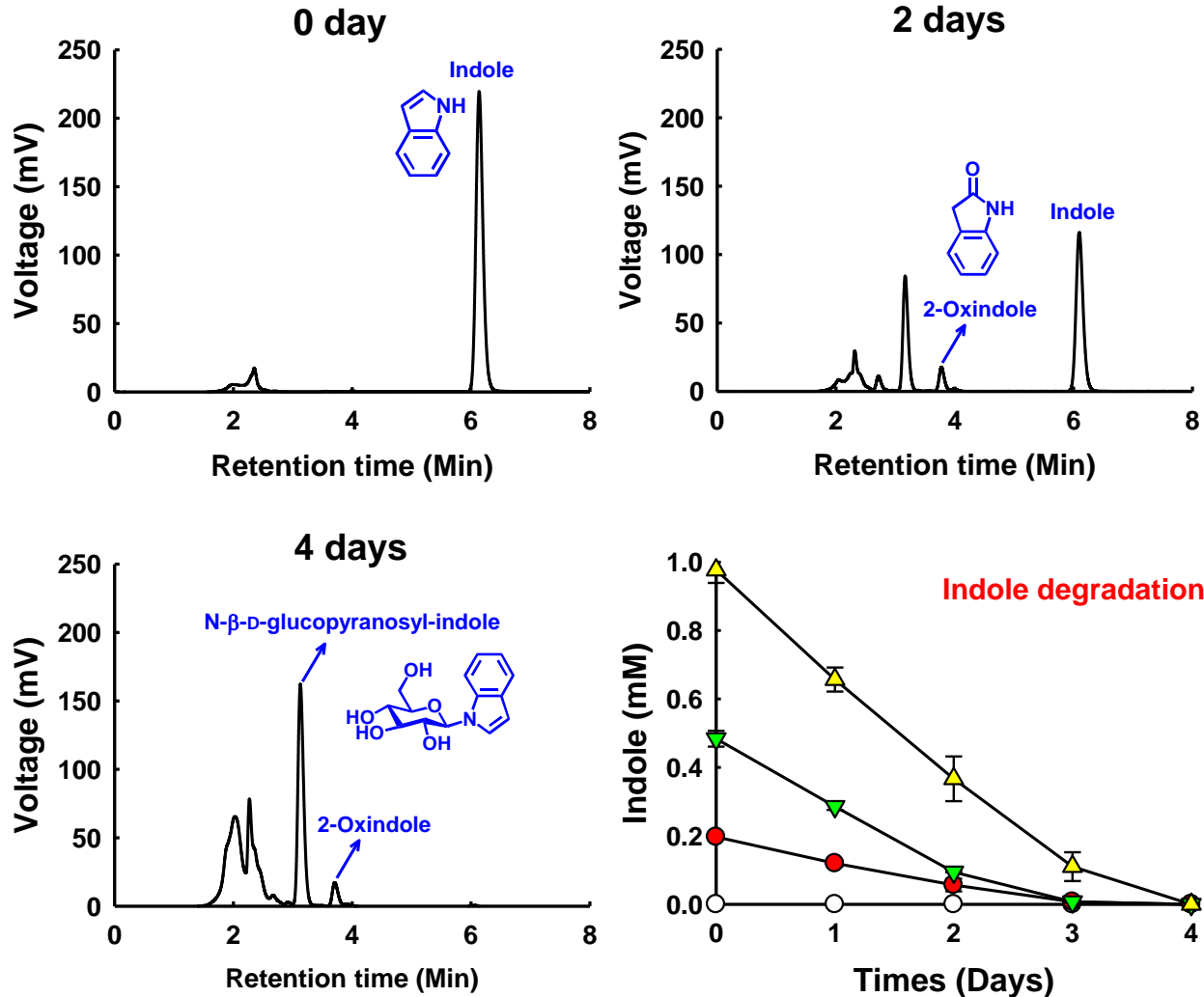


# Indoles in *Caenorhabditis elegans*



- *C. elegans* senses indole and its derivatives

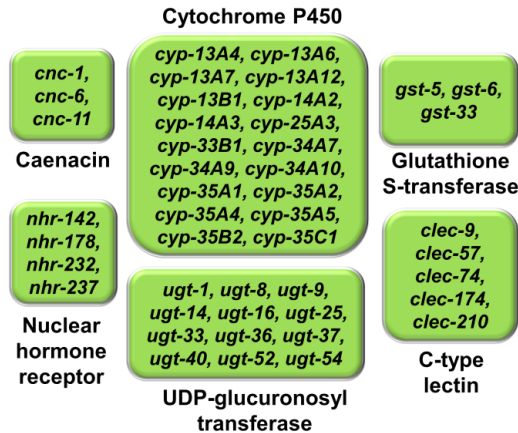
# Indole degradation by *C. elegans*



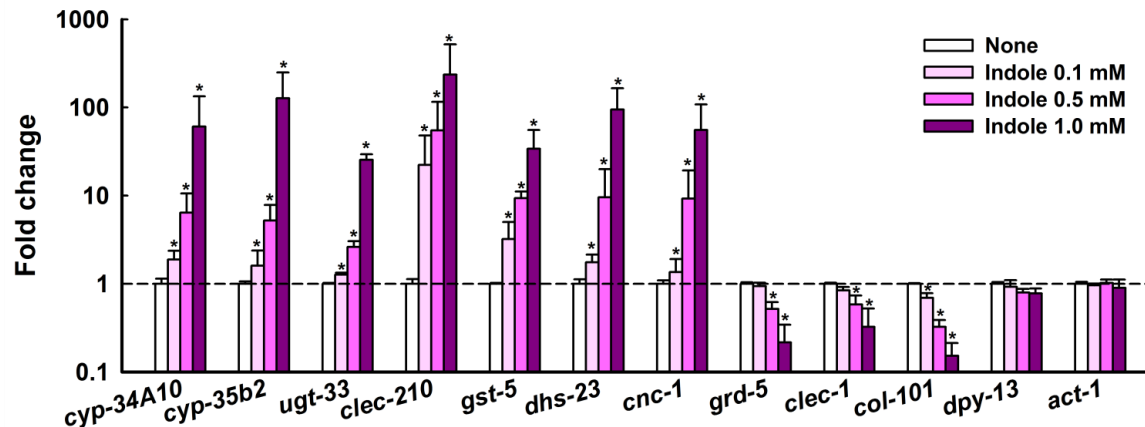
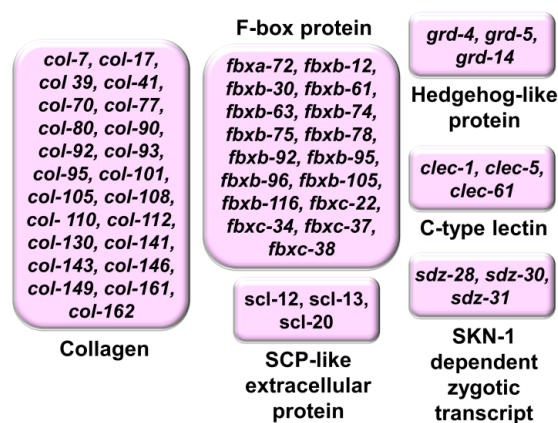
- *C. elegans* degrades indole by oxidation of glycosylation.

# Indole controls gene expression of *C. elegans*

## Up-regulated genes

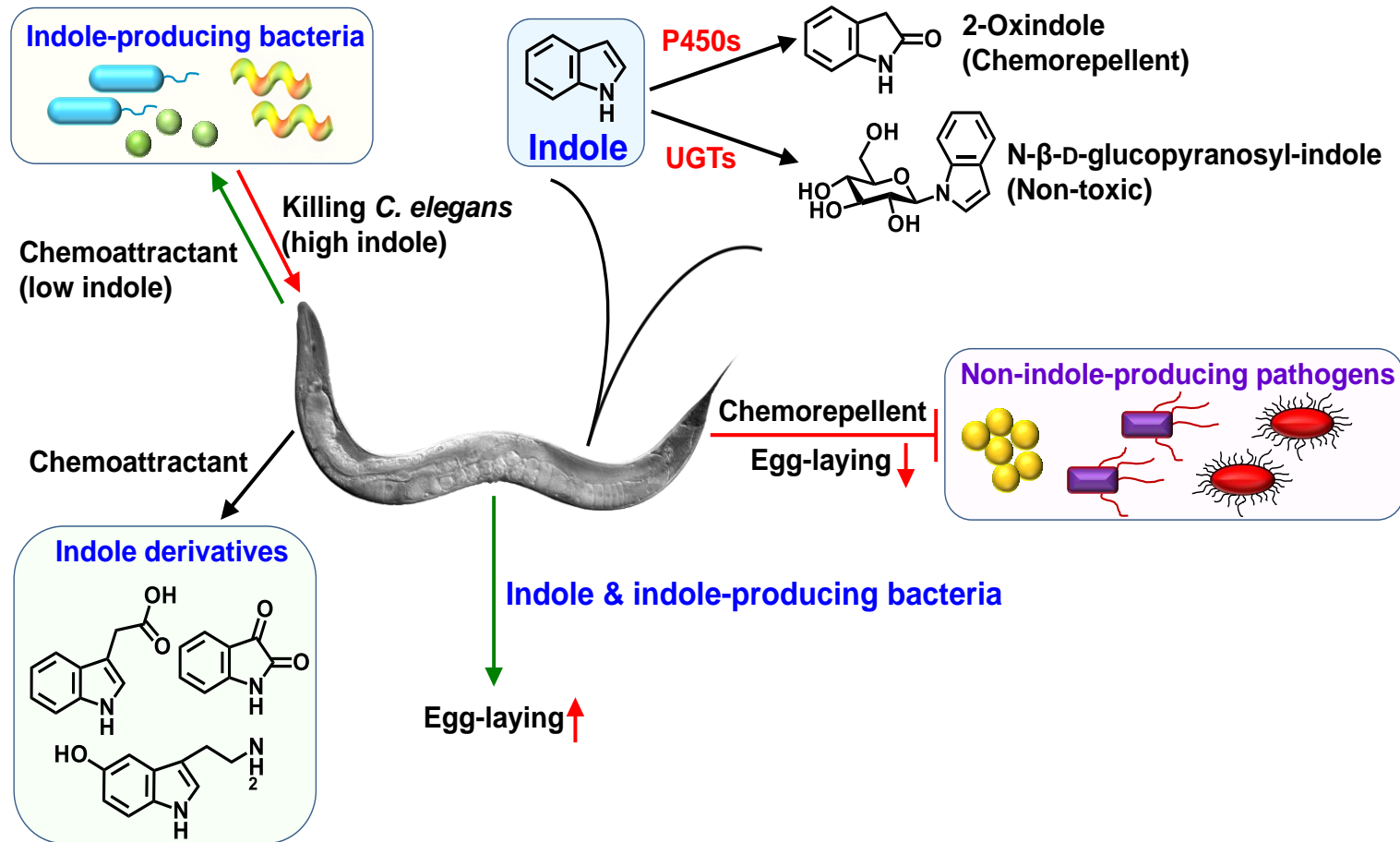


## Down-regulated genes

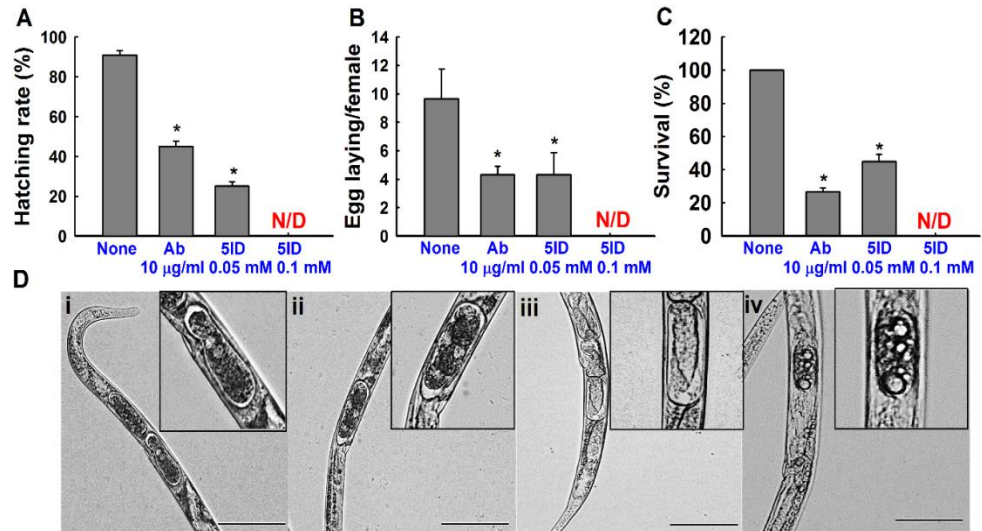
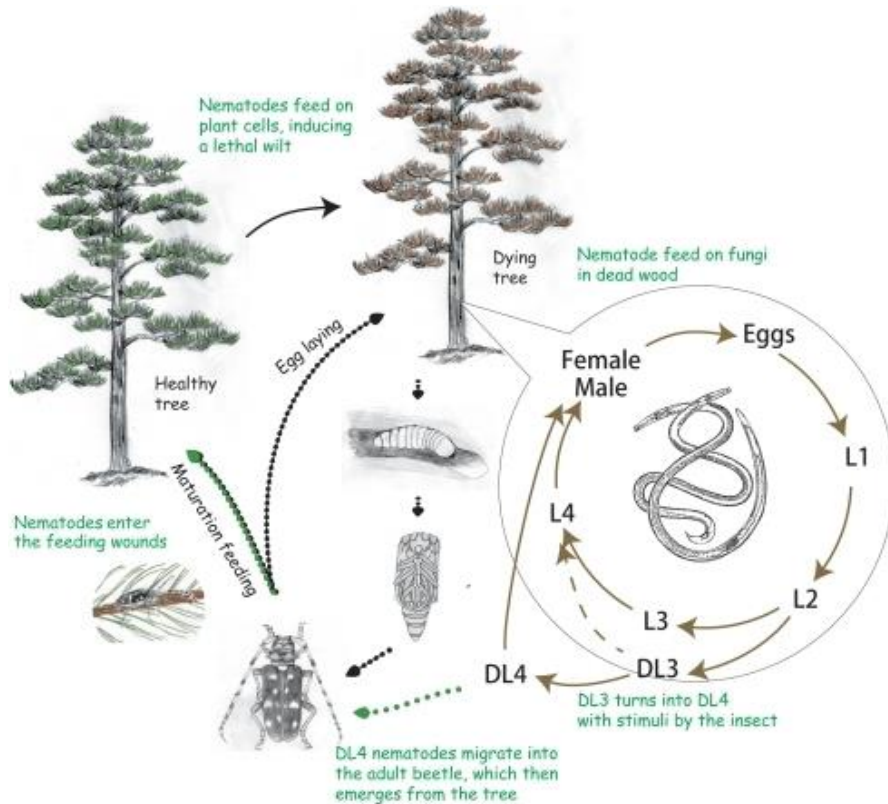


- Indole control expression of many genes in *C. elegans*.

# Model of bacteria-nematode interactions



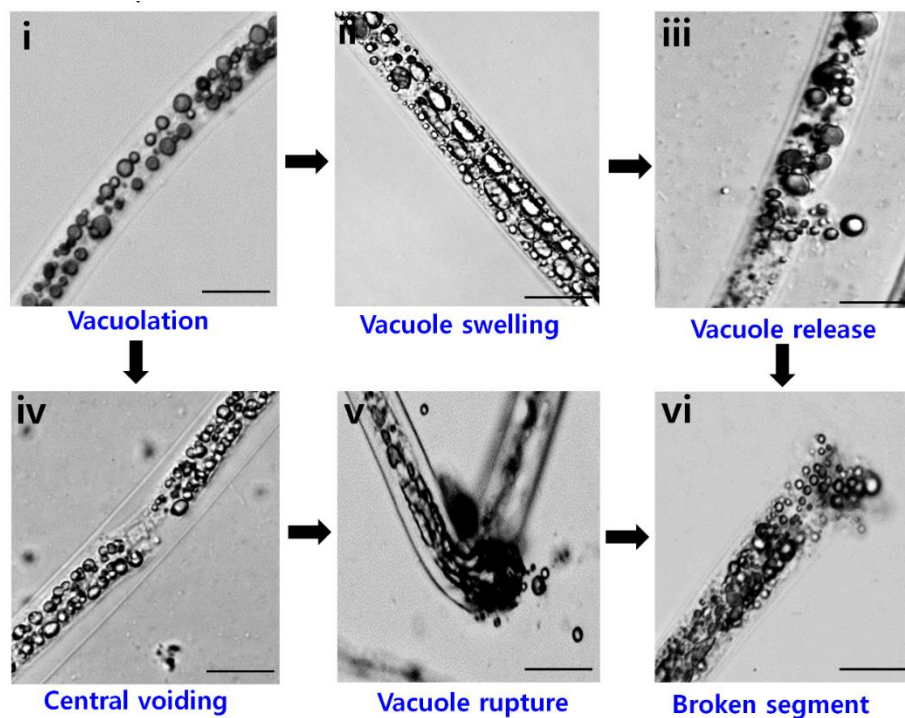
# Killing pine tree nematode with indoles



Some indole (like Abamectin) can kill pine tree nematode.

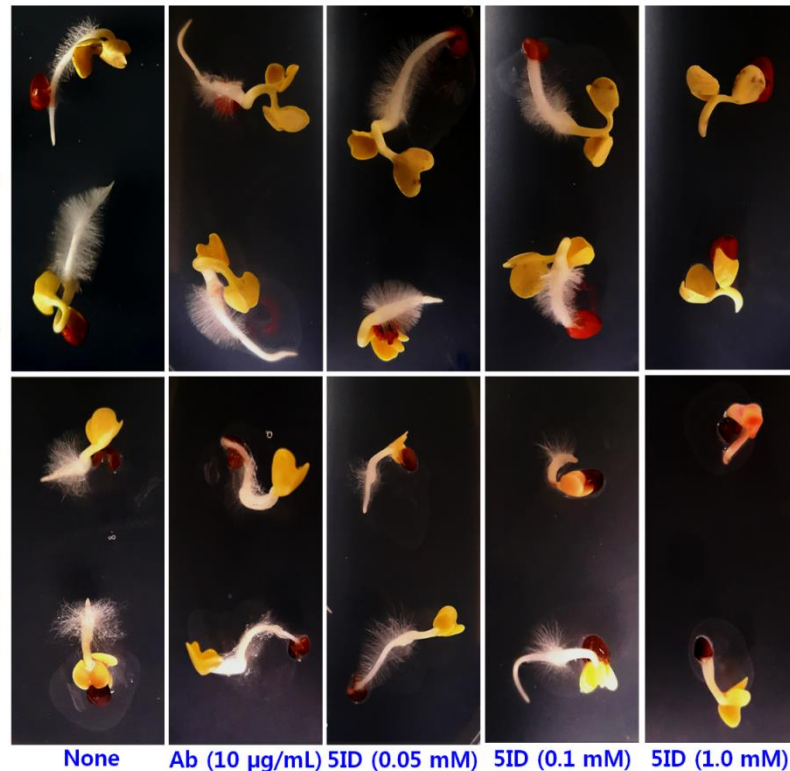


# Indoles produce vacuoles and non-toxic to plants



*Raphanus raphanistrum*

*Brassica oleracea*



Some indoles are new nematicides  
(under review)

# Conclusion of indole story

---

**Indole and its derivatives are intercellular, inter-species,  
and inter-kingdom signaling molecules.**

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- Prof. Toshinari Maeda (Kyutech, Japan)
- Prof. Younghoon Kim (Chonbuk National University, Korea)
- Prof. Rodolfo García-Contreras (UNAM, Mexico)

## Lab members:

- Post-docs: **J-H Lee**, M Ram, MR Kumar, **RK Satish**
- Ph.D. student: **YG Kim**
- M.S. students: MS Kim, GY Kwon

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