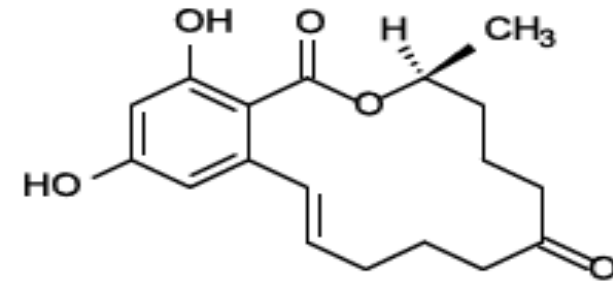


**GENOTOXICITY  
EVALUATION OF  
ZAERALENONE WITH THE  
WING SPOT TEST IN  
*Drosophila melanogaster***

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Dueñas-García  
I.E., Piedra-  
Ibarra E.**

# ZEARALENONE

- ZEA is a mycoestrogen produced as a secondary metabolite of *Fusarium* spp., that contaminates cereal crops worldwide, and produces mycotoxicosis of farm animals.



# ZEA HUMAN CONSUMPTION PER DAY

- The maximum tolerated levels of ZEA for human consumption have been established as:
  1. 20  $\mu\text{g}/\text{kg}$  bw in food intended for babies and infants.
  2. 50  $\mu\text{g}/\text{kg}$  bw in maize-based snacks and breakfast cereals.
  3. 200  $\mu\text{g}/\text{kg}$  bw in unprocessed maize and certain maize products.

# SOME REGIONAL DIETS HAVE MORE DIETARY INTAKE (WHO)

$\mu\text{g}/\text{day}$  based on 60 kg of bw

## LATIN AMERICA

- Maize < 0.51
- Rice < 0.64
- Wholemeal bread < 0.42

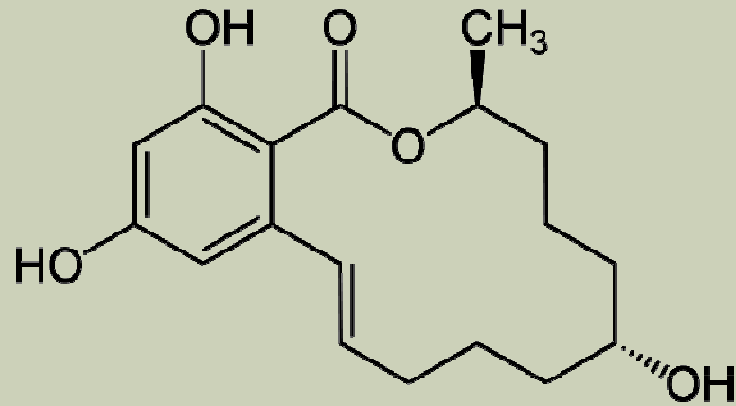
## AFRICAN

- Maize < 1.3
- Rice < 0.76
- Wholemeal bread < 0.053

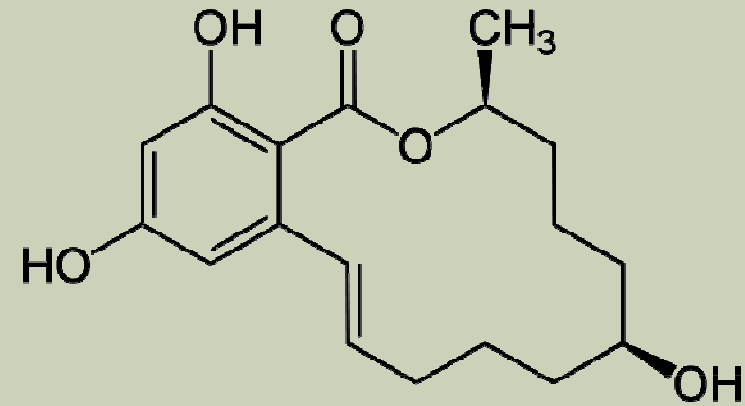
# A GROWTH PROMOTER FOR LIVESTOCK

- A synthetic commercial formulation called zeranol (Ralgro®) has been marketed successfully for use as an anabolic agent for both sheep (12 mg/3 months) and cattle (36 mg/3 months).
- In 1989, zeranol ( $\alpha$ -zearalanol) was banned by the European Union, but it is still used in other parts of the world.
- In Mexico it has official registration by the authorities (Reg. SAGARPA. Q-0552-084).

# METABOLITES



$\alpha$ -ZOL



$\beta$ -ZOL

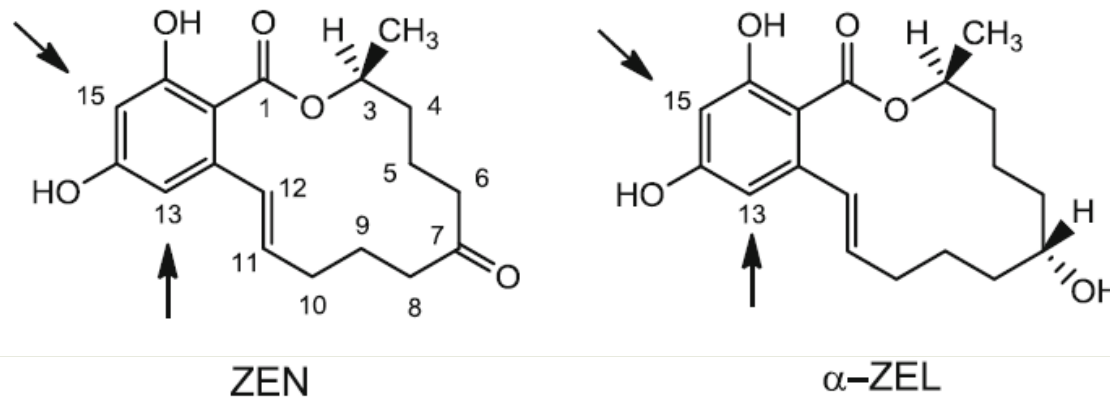
ZEA is reduced by steroid dehydrogenase metabolism to the major metabolites alpha- and beta-zearalenol ( $\alpha$ -,  $\beta$ -ZOL), that have in vertebrates a higher estrogenic activity than ZEA.

## SOME ZEA EFFECTS

- ZEA resembles  $17\beta$ -estradiol and binds to estrogen receptors in vertebrate target cells causing hyperestrogenic syndromes.
- In mammals it have been related with an increased risk for endometrial adenocarcinomas or hyperplasia and breast cancer.
- ZEA has been associated also with oxidative damage and induction of SOS repair.

Fleck et al. *Mycotoxin Res.* 2012 Nov;28(4):267-73. Sáenz de Rodríguez *et al.*, 1985; Tomaszewski *et al.*, 1998; Abid-Essefi, *et al.*, 2004; Ghedira-Chékir, *et al.*, 1998.

# HYDROXYLATION



As  $17\beta$ -estradiol, ZEA and metabolites undergo P450s hydroxylation at the positions marked with arrows that leads to catechol metabolites or glucuronides and sulfates conjugates.

Resulting catechols can cause genotoxic effects (ROS) and may contribute to an increased cancer risk in estrogen target tissues.



# IN VITRO EFFECTS OF ZEA

- Ayed-Boussema *et al.* (2007) monitored cytotoxicity, cell cycle perturbation, genotoxicity and mutagenicity in Vero cells (kidney epithelial) exposed to ZEA.
  - a) ZEA (1 -60  $\mu\text{M}$ ) reduced cell viability correlated to DNA fragmentation (DNA-laddering patterns).
  - b) The induction of apoptotic bodies and cell cycle perturbation was demonstrated in a dose-dependent manner (ZEA 10, 20, 40  $\mu\text{M}$ ) .
  - c) ZEA (5, 10, 20  $\mu\text{M}$ ) produced micronuclei and chromosome aberrations.

Ayed-Boussema *et al.* J Biochem Mol Toxicol. 2007;21(3):136-144.

# GENOTOXICITY OF ZEA , ALPHA- AND BETA-ZOL

- Ayed *et al.* (2011) demonstrated in mouse bone-marrow cells\* and in HeLa cells♦ that ZEA,  $\alpha$ - and  $\beta$ -ZOL inhibited cell viability in a dose-dependent manner.
- ZEA and its two metabolites increased the percentage of chromosome aberrations. ZEA and  $\alpha$ -ZOL were more genotoxic than  $\beta$ -ZOL.
- Authors conclude that biotransformation of ZEA may be considered as only a partial detoxification pathway since the resulting metabolites remain relatively toxic.

Ayed *et al.* Mutat Res. 2011 Nov 27;726(1):42-46. \*100  $\mu$ L of 5, 10, 20 mg/Kg; ♦ 12.5, 25 & 50% loss of cell viability.

# ZEA & P450S *IN VITRO*

- Metabolic monohydroxylation of ZEA by human cytochrome P450s (CYP2C8; CYP3A4, CYP3A5) and phenobarbital-treated rat (CYP2C11) was demonstrated *in vitro* with liver microsomes yielding several OH-ZEA products limited, and not as significant on estrogen receptors as those of ZEA and  $\alpha$ -ZOL.\*
- CYP1A2 is highly active in the hydroxylation of ZEA.\*\*

\*Ding *et al.* Toxicol Sci. 2006 Jun;91(2):448-55. Bravin *et al.* Int J Mol Sci. 2009 Apr 21;10(4):1824-37. \*\*Drzymala *et al.* Chem Res Toxicol 2014.

# THE QUESTIONS

- Is genotoxicity of ZEA caused by:
  1. Stimulation of estrogen receptors?
  2. The  $\alpha$ - and  $\beta$ -ZOL metabolites?
  3. The OH-ZEA P450s products?

In an animal model without estrogen receptors and different P450s levels (induced and high) we could address questions 2 and 3.

## *ERR, SCU & CYP1A2*

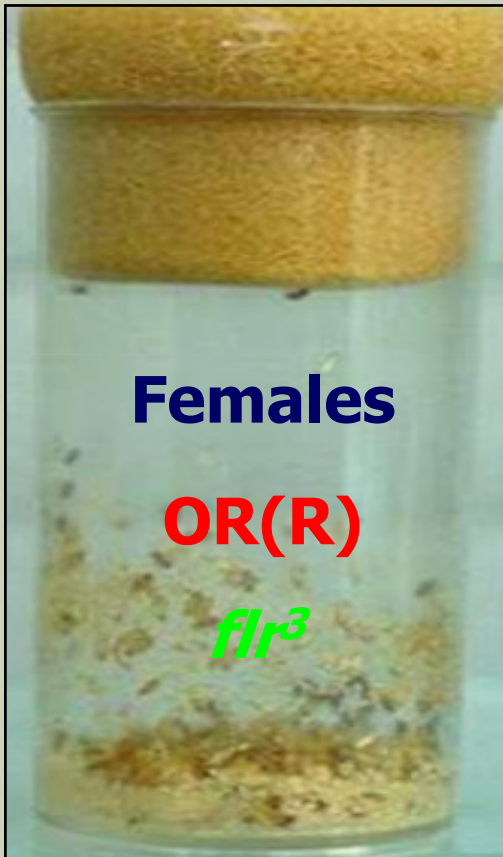
- *Drosophila melanogaster* has a single estrogen-related receptor *ERR* gene that codes for the dERR protein, which belongs to the orphan receptors family, with no known naturally occurring ligand, but is an essential regulator of carbohydrate metabolism during larval stages.
- It has a steroid dehydrogenase activity, acting on the CH-CH group of donors, by a protein product of the *scu* gene.
- The *Cyp1a2* gene of *Drosophila* is homolog to the human *CYP1A2* gene.

# METHODS

# SMART

- We evaluated the genotoxicity of ZEA (0, 100, 200, 400  $\mu\text{M}$ ), dissolved in phosphate buffer solution (PBS) pH 7, under and above  $\text{LC}_{<25}$ , with the standard (ST) and the high bioactivation (HB) crosses, with basal and high levels of P450s, respectively.

# ST & HB CROSSES



**Males**  
*mwh*



# ST & HB Crosses



- ST (*flare X mwh*)
- HB (*Oregon-flare X mwh*)

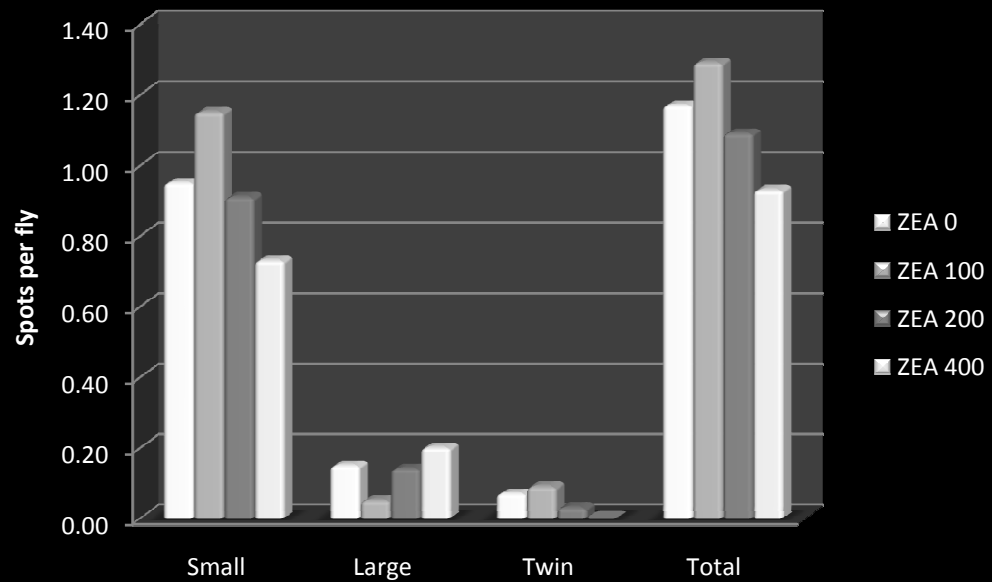
# ZEA CHRONIC TREATMENTS

TREATMENT	MilliQ WATER	PO <sub>4</sub> BUFFER (PBS) pH 7	ZEA μM	URE mM
MilliQ water	+			
PBS		+		
ZEA		+	100	
ZEA		+	200	
ZEA		+	400	
Urethane	+			20

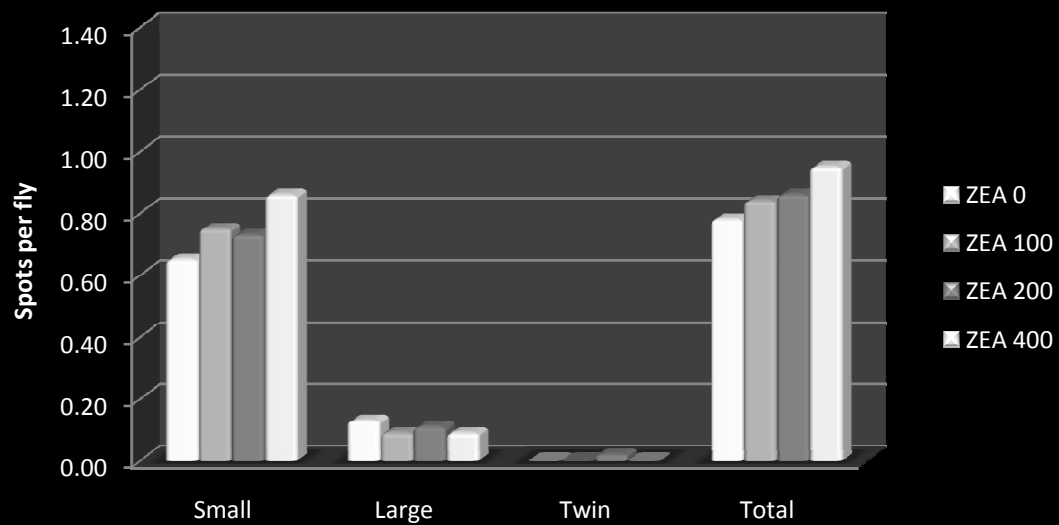
**SMART RESULTS**

<b>ST CROSS, <i>p</i>&lt;0,05</b>	<b>Number of flies</b>	<b>Small (1-2 cells)</b>	<b>Large (&gt; 2 cells)</b>	<b>Twin</b>	<b>Total</b>
<b>MilliQ Water</b>	<b>71</b>	<b>0,70</b>	<b>0,06</b>	<b>0.08</b>	<b>0.85</b>
<b>PBS</b>	<b>59</b>	<b>0.95</b>	<b>0,15</b>	<b>0.07</b>	<b>1,17</b>
<b>ZEA 100 <math>\mu</math>M</b>	<b>55</b>	<b>1,15 -</b>	<b>0,05 -</b>	<b>0,09 -</b>	<b>1,29 -</b>
<b>ZEA 200 <math>\mu</math>M</b>	<b>58</b>	<b>0,91 -</b>	<b>0,14 -</b>	<b>0,03 -</b>	<b>1,09 -</b>
<b>ZEA 400 <math>\mu</math>M</b>	<b>55</b>	<b>0,73 -</b>	<b>0,20 -</b>	<b>0,00 -</b>	<b>0,93 -</b>
<b>URE 20 mM</b>	<b>38</b>	<b>3,13 <math>\uparrow</math></b>	<b>0,24 <math>\uparrow</math></b>	<b>0,05 ns</b>	<b>3,42 <math>\uparrow</math></b>
<b>HB CROSS <i>p</i>&lt;0,05</b>	<b>Number of flies</b>	<b>Small (1-2 cells)</b>	<b>Large (&gt; 2 cells)</b>	<b>Twin</b>	<b>Total</b>
<b>MilliQ Water</b>	<b>60</b>	<b>0,75</b>	<b>0,10</b>	<b>0,02</b>	<b>0,87</b>
<b>PBS</b>	<b>55</b>	<b>0,65</b>	<b>0,13</b>	<b>0,00</b>	<b>0,78</b>
<b>ZEA 100 <math>\mu</math>M</b>	<b>55</b>	<b>0,75 -</b>	<b>0,09 -</b>	<b>0,00 -</b>	<b>0,84 -</b>
<b>ZEA 200 <math>\mu</math>M</b>	<b>56</b>	<b>0,73 -</b>	<b>0,11 -</b>	<b>0,02 -</b>	<b>0,86 -</b>
<b>ZEA 400 <math>\mu</math>M</b>	<b>56</b>	<b>0,86 -</b>	<b>0,09 -</b>	<b>0,00 -</b>	<b>0,95 -</b>
<b>URE 20 mM</b>	<b>39</b>	<b>6,31 <math>\uparrow</math></b>	<b>0,62 <math>\uparrow</math></b>	<b>0,26 <math>\uparrow</math></b>	<b>7,18 <math>\uparrow\uparrow</math></b>

## ST CROSS



## HB CROSS

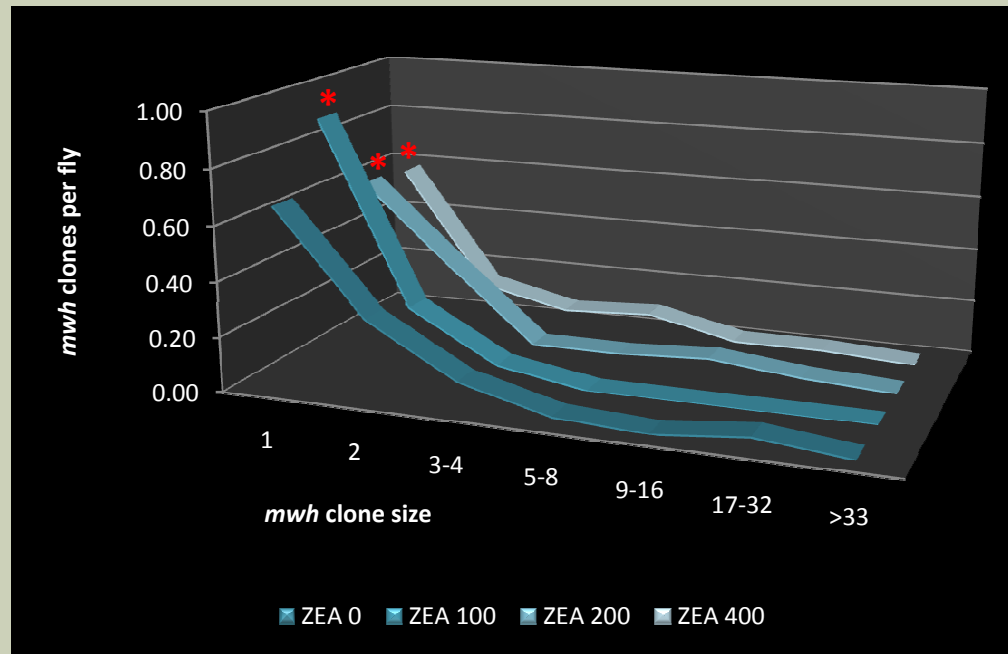


# HB VS ST

Statistical diagnoses according to Frei and Würgler (1988, 1995),  $p < 0.05$ .

NEGATIVE CONTROLS	TOTAL SPOTS/FLY
Distilled Water ST	0,85
Distilled Water HB	0,87 -
PBS ST	1,17
PBS HB	0,78 -
ZEA TREATMENTS	
100 $\mu$ M ST	1,29 $\uparrow$
100 $\mu$ M HB	0,84
200 $\mu$ M ST	1,09 -
200 $\mu$ M HB	0,86
400 $\mu$ M ST	0,93 -
400 $\mu$ M HB	0,95
URE POSITIVE CONTROL	
20 mM ST	3,42
20 mM HB	7,18 $\uparrow$

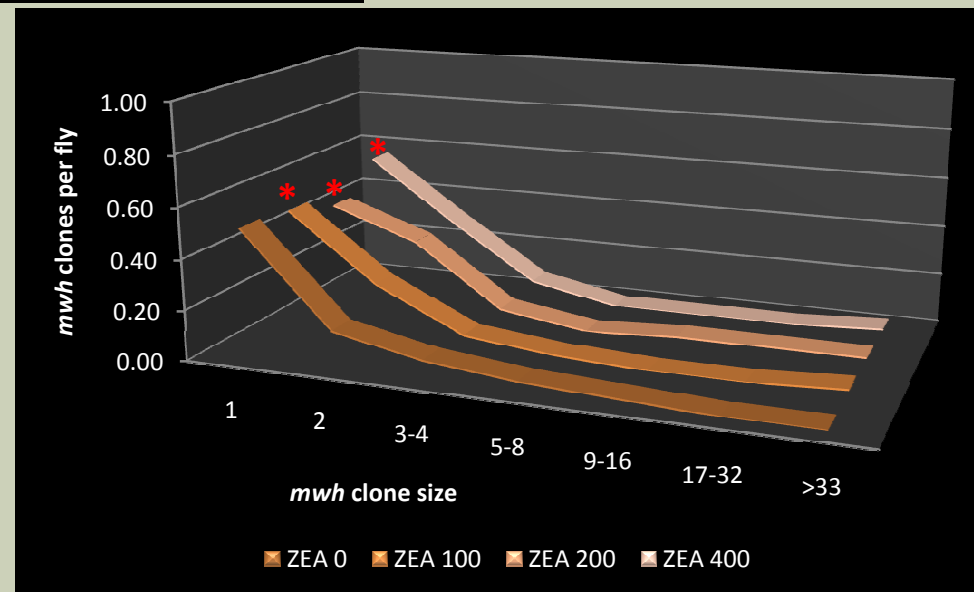
# Distribution of *mwh* clone size



HB CROSS

Clones size are produced by a number of cell division cycles after damage.

We did Kolmogorov-Smirnov analysis of distribution of the mean accumulated *mwh* clone size classes,  $p < 0.05$ .



# DISCUSSION

We have demonstrated that the *hsp60* gene is overexpressed in the flare strain (inducible P450s) treated with ZEA 260  $\mu\text{M}$  (unpublished data).

Because the Hsp60 family of chaperonins has anti and pro-apoptotic properties\*, we propose that in the ST cross (inducible P450s) the significant differences between small, twin and total spot frequencies from 100 vs. 400  $\mu\text{M}$ , could be related to ZEA apoptotic properties.

\*Arya *et al.* J Biosci 2007. 32:595–610



# DISCUSSION

## Toxicity (unpublished data)

- ✓ In  $LC_{50}$  experiments, ZEA was not toxic in the flare or Oregon-flare strains, but the flare strain was more sensitive ( $LC_{20} = 155.25 \mu\text{M}$ ) than the Oregon-flare strain ( $LC_{20} = 260 \mu\text{M}$ ).

These data can explain the low total spot frequency (0.84) of the HB cross fed on ZEA  $100 \mu\text{M}$ , since it presents high P450s levels, and the higher frequency in the ST cross (1.29) with inducible levels.

# DISCUSSION

- In the HB cross we did not find any significant differences between treatments even greater effect could be expected because the possible formation of more ZEA/ $\alpha$ -ZOL catechol metabolites & oxidative damage.
- We propose these results can be explained by a higher detoxifying metabolism due to the high P450s levels in this cross.

# CONCLUSIONS

- In contrast to previous genotoxicity reports in mammalian cells, we did not find any positive results in the *Drosophila* wing spot test, ST and HB crosses.
- Our results indicate that the ST cross is more sensitive to the tested ZEA treatments, than the HB cross. This could be explained by the difference between their CYPs levels.
- Because of the lack of an estrogen receptor in *Drosophila* results maybe due only to the effects of ZEA metabolites: alfa, beta-ZOL and OH-ZEA.

# CONCLUSIONS

- It is recommended to use apoptosis markers in order to demonstrate in both crosses if this event is induced in the ST cross fed with ZEA.
- Further studies are needed to assess levels of ZEA metabolites in third instar larvae to imago emergence of *Drosophila* flare and Oregon-flare mutant strains.

THANK YOU

