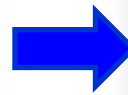


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What are the characteristics of white rot fungi that make them interesting candidates for solving some environmental problems?

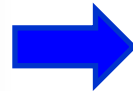
WHITE-ROT FUNGI



Ligninolytic enzymes
Secondary metabolites

WRF play important roles in ecosystems

WHITE-ROT FUNGI



ENVIRONMENTAL APPLICATIONS

Ligninolytic enzymes

Bioremediation

- Pesticide degradation
- Detoxification of industrial dyes

Fungal volatile organic compounds (FVOCs)

Chemical markers of fungal presence

Biopesticides



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Natural products of *Anthracophyllum discolor*: ligninolytic enzymes and antifungal volatile compounds

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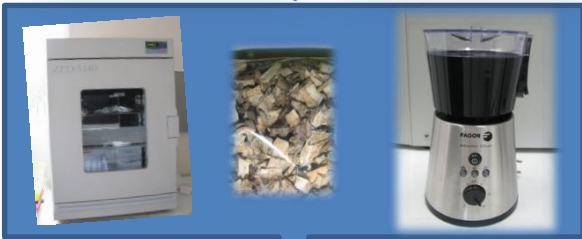
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Frankfurt, 2015

Objective

To evaluate the production of ligninolytic enzymes and antifungal volatile organic compounds (VOCs) by *A. discolor* Sp4 using potato peels (PP) and discarded potato (DP) as low cost nutritional supports.

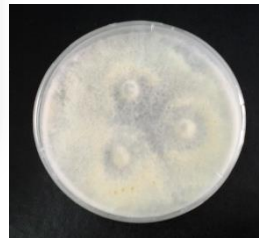
Processing potato wastes



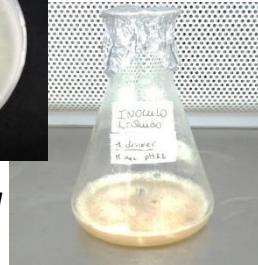
Inoculum



A. discolor



Agar media



Liquid media

1. Production of ligninolytic enzymes

- Laccase
- Manganese peroxidase
- Lignin peroxidase

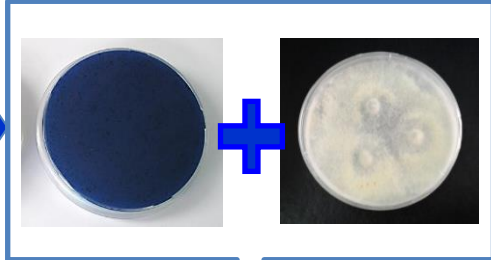
2. Production of antifungal volatile compounds

1. Production of ligninolytic enzymes

1.1. Dye decolorization test



Remazol Brilliant
Blue R (RBBR)
(0.05% w/v)



1. Kirk + agar
2. Discarded potato agar (DPA)
3. Potato peel agar (PPA)

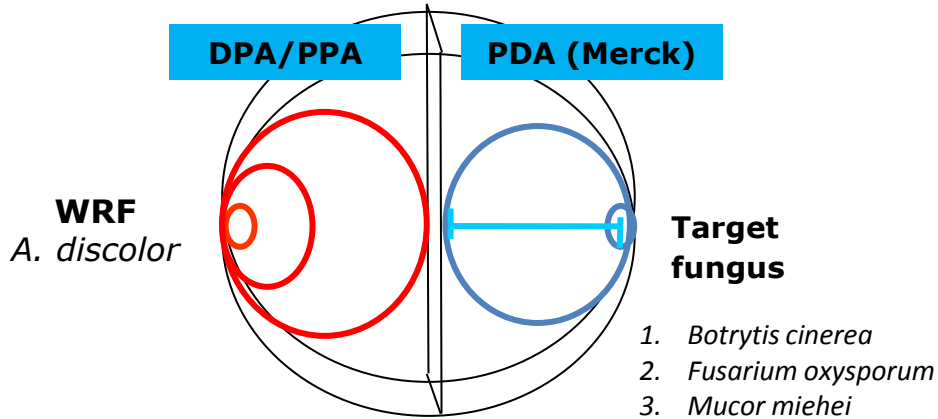


1.2. Ligninolytic enzyme activities



Lac a	MnP a	LIP activity
independe	oxidati	was measured by monitoring
(DMP)	adding	the oxidation of veratryl alcohol to
	veratraldehyde	at 310 nm

2. Production of antifungal volatile compounds

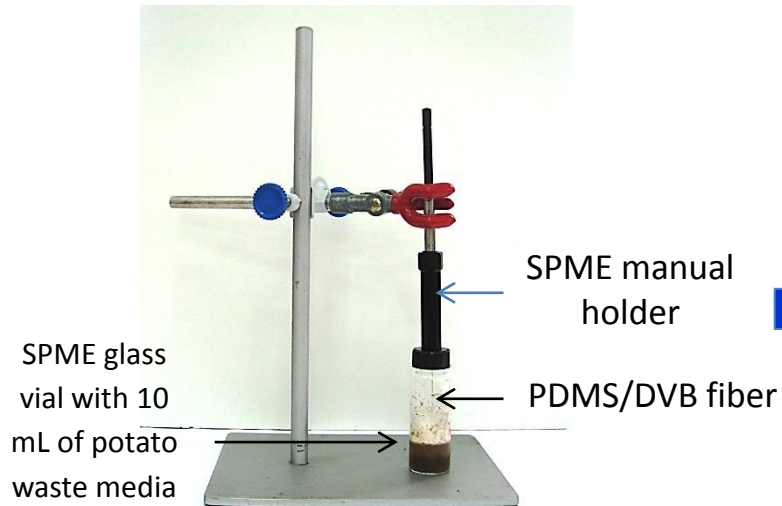


Bi-compartmented Petri dish assay



Analisis of VOCs effect by screening electron microscopy (SEM).

2. Production of volatile compounds



Solid phase micro-extraction (SPME)



Analisis of VOCs by gas chromatography coupled to mass spectrometry (GC-MS)

1. Production of ligninolytic enzymes

1.1. Dye decolorization test

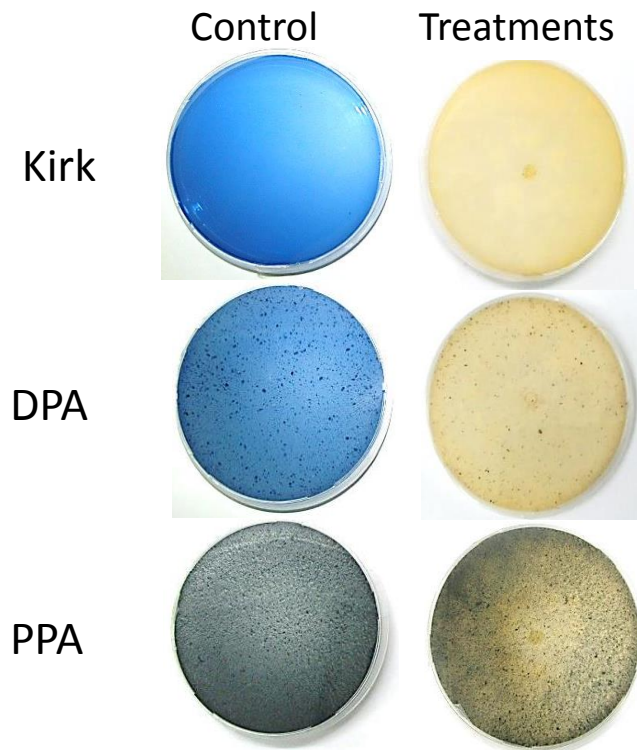


Figure 1. Decolorization RBBR (0.05% w/v) by *A. discolor* incubated during 21 days on discarded potato agar (DPA), potato peel agar (PPA) and modified Kirk medium.

1.2. Ligninolytic enzyme activities

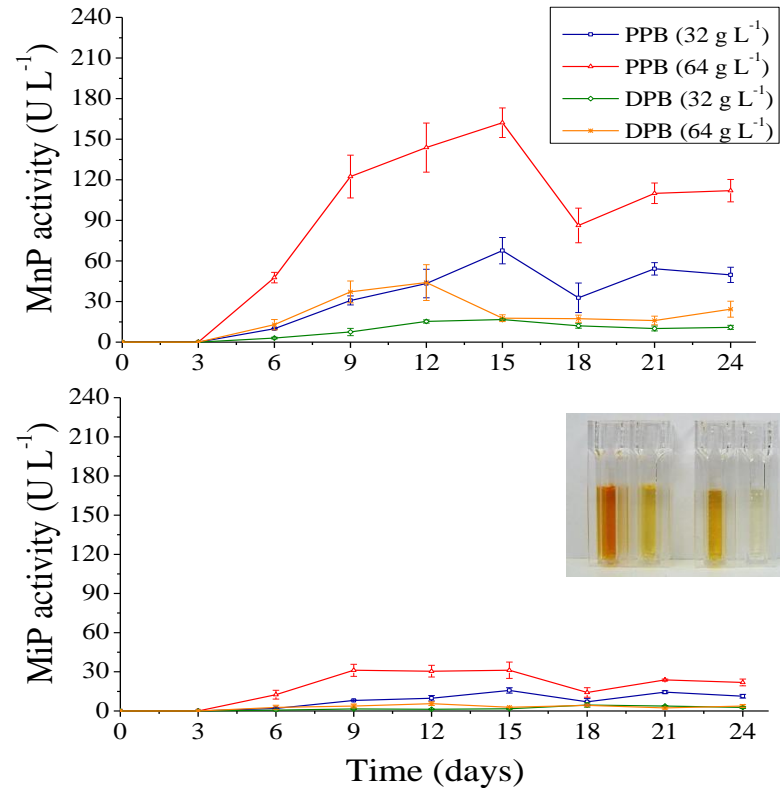


Figure 2. Manganese peroxidase (MnP) and manganese-independent peroxidase (MiP) activities by *A. discolor* Sp4 cultured in potato peel broth (PPB) and discarded potato broth (DPB) at two potato powder concentrations.

2. Production of volatile compounds

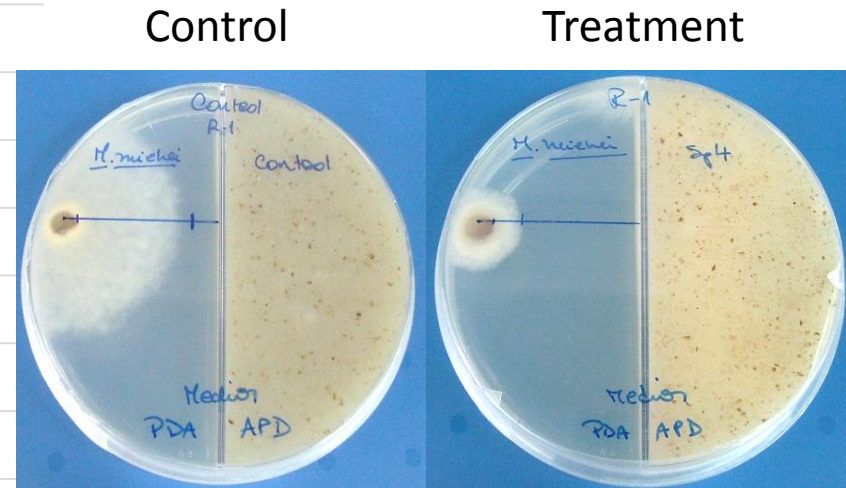
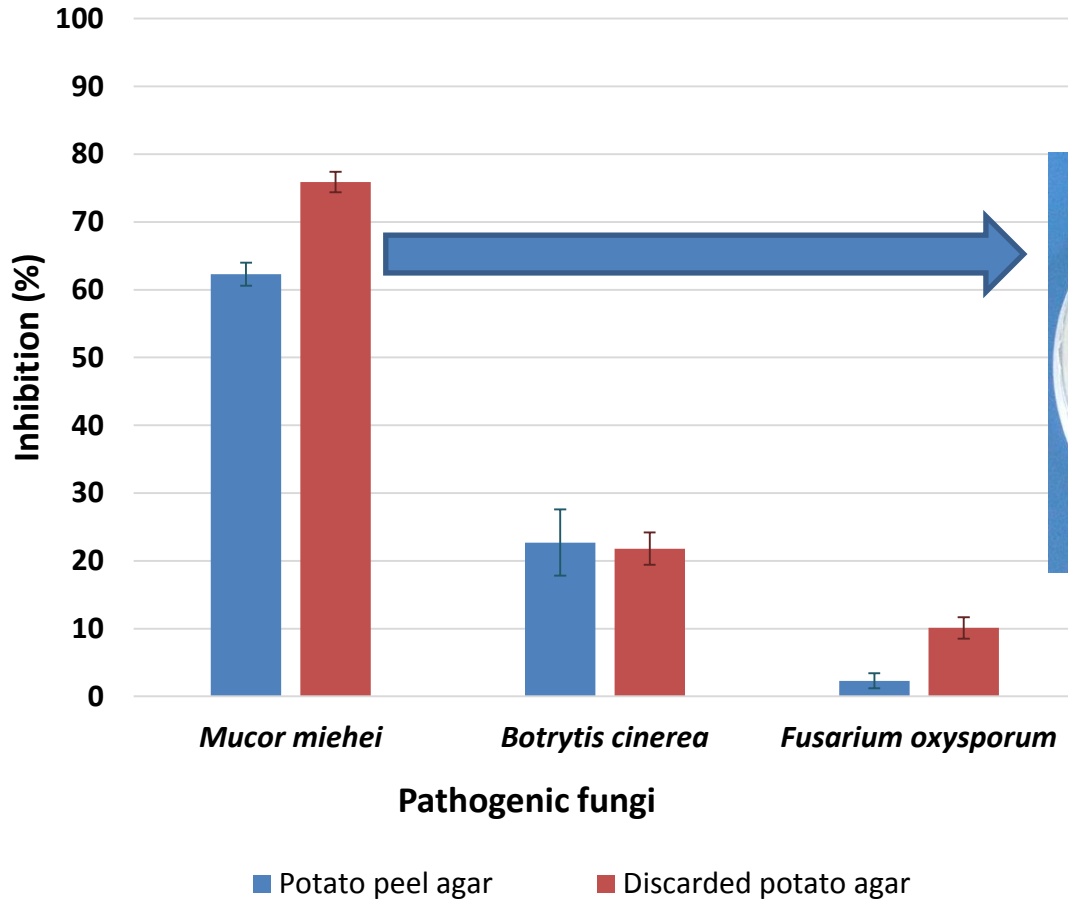


Figure 3. Antifungal activity of volatile compounds released from *A. dicolor* against *B. cinerea*, *F. oxysporum* and *M. miehei*.

2. Production of volatile compounds

CONTROL

ANTAGONIZED HYPHAE

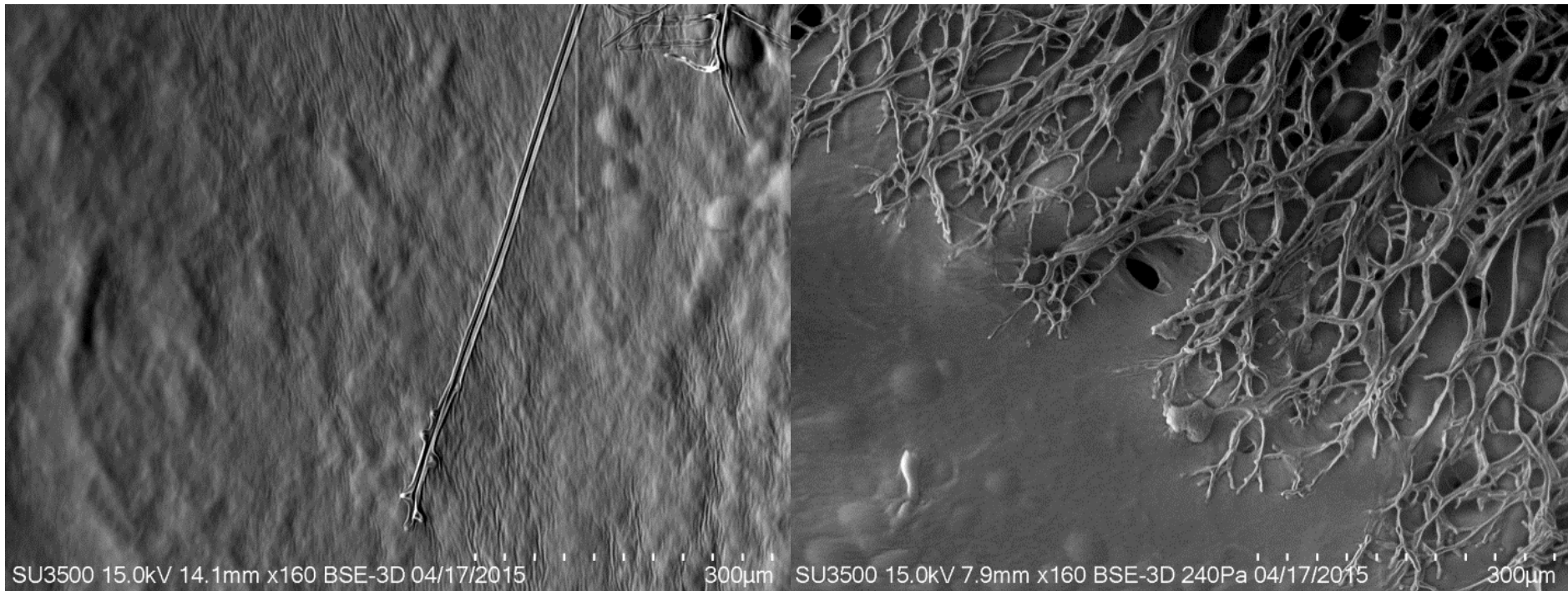
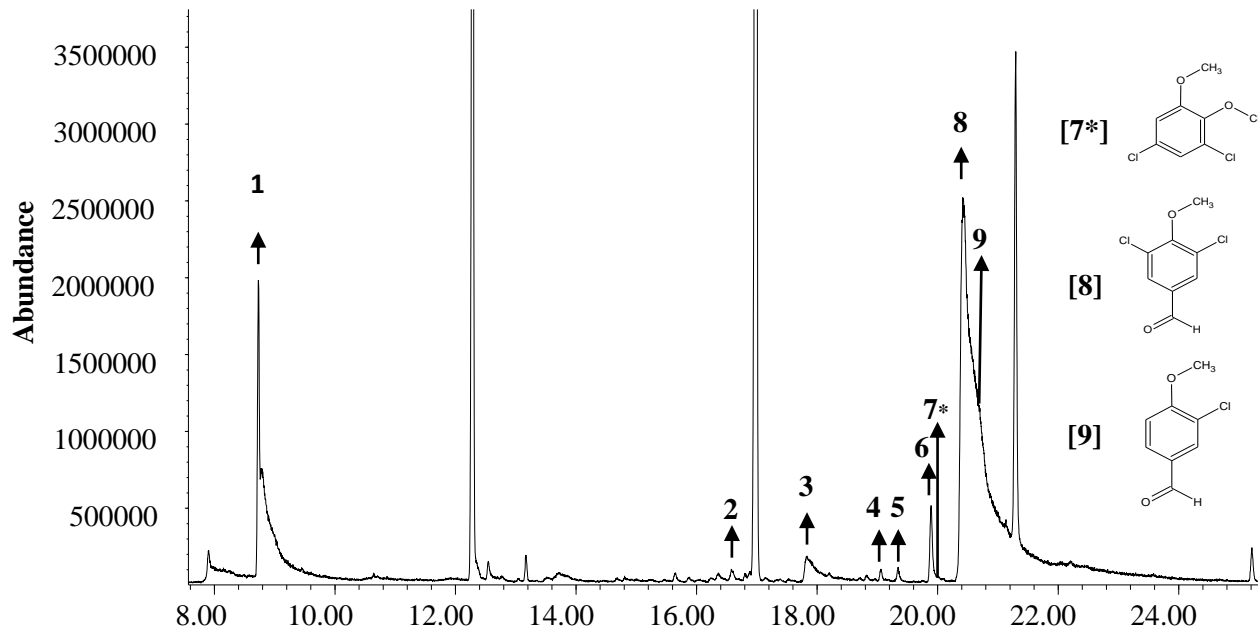


Figure 4. SEM images of untreated control hyphae of the target fungus *M. miehei* (left) and the antagonized hyphae of *M. miehei* exposed to the antifungal volatiles of *A. discolor* Sp4 (right).

2. Production of volatile compounds



The mass spectrums of peaks pointed to the structures of:

[1] 1-heptanol

[2] 2-butyl-1-octanol

[3] 3-ethyl-1,2-dihydroquinoxalin-2-one

[4] α -bisabolene

[5] 1,2,4a,5,6,8a-hexahydro-4,7-dimethyl-1-(1-methylethyl)-naphthalene

[6] bulnesene

[7] 1,5-dichloro-2,3-dimethoxybenzene*

[8] 3,5-dichloro-4-methoxybenzaldehyde

[9] 3-chloro-4-methoxybenzaldehyde.

Figure 5. Gas chromatogram of VOCs released from *A. discolor* Sp4 cultured on DPA medium (discarded potato agar). Peaks appearing in the control vial were omitted from the list (N=3).

Conclusions

The fungus *Anthracoephyllum discolor* Sp4 are able to produce ligninolytic enzymes using potato wastes as nutritional support.

VOCs released from *A. discolor* Sp4 showed antifungal activity against *Mucor miehei* and *Botrytis cinerea*. The antifungal activity seems to be related to the production of sesquiterpenes and chlorinated aromatic compounds.

ACKNOWLEDGEMENTS



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Acknowledgement

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