EFFECT OF ELECTROMAGNETIC FIELD INDUCED BY RADIO FREQUENCY WAVES AT 900 TO 1800 MHZ ON GROWTH PLATE IN GROWING RATS

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EMFs have biological effects on

- the behavior of bone cells,
- increase the maturation of bone trabecula,
- bone volume and formation.

(Rothman et al., 1996; Tabrah et al., 1998)

Some authors have observed that an external magnetic field accelerated the healing of bone fractures.

(Pickering and Scammell, 2002; Fini et al., 2008)

However, Yamada et al. (1985) did not observe any effects on bone tissue.

Although some papers have focused on the general effects of high frequency EMF on various organs and tissues, there is still no comprehensive study exploring the underlying mechanisms of the effects of high frequency EMF on bone tissue.

(Koyama et al., 2003; Diem et al., 2005; Koyu et al., 2005; Lixia et al., 2006; Zhang et al., 2006; Schwarz et al., 2008; Aydin and Akar, 2011; Dasdag et al., 2012, Kostoff and Lau, 2013)

The main goal:

to evaluate the effects of high frequency EMF exposure on the femoral epiphyseal growth plates in growing male rats.

MATERIALS AND METHODS

The animal study was approved by the Experimental Animal Studies Ethics Committee of Ondokuz Mayis University.



> Thirty-three (2 days old) male albino Wistar rats

> The rats were randomly divided into three independent groups:





900 MHz EMF group (n=11),



control group (n=11).

The rats were held in a piecage restrainer and then exposed 2 h/day for 90 days, at the same time.



Exposure system

to apply 900 MHz radio frequency signal to group 2, Everest GSM Simulator (900 CW4, Adapazarı, Türkiye);

to apply 1800 MHz radio frequency signal to group 3, Everest GSM Simulator (1800 CW2, Adapazarı, Türkiye) were used.



EMF measurements were performed with Portable Field Meter (PMM 8053, Costruzioni Elettroniche Centro Misure Radioelettriche Srl., Italy)

Equal distribution of electrical area was achieved by placing the dipol antenna in the middle point of restrainer.

> Antenna length was approximately 15 cm.



The power levels antenna output power values of the RF generator exposure system were kept as value of which shows the same effect to the cellular and digital communication handsets commonly used by the general public.

SAR calculation

The SAR is expressed in watt per kilogram (W/kg). The SAR given;

$$SAR(x, y, z) = \sigma(x, y, z) \cdot \frac{[E_{x, y, z}]_{rms}^2}{\rho} \quad (W/kg)$$

These SAR calculations were obtained after 500 000 iterations in MATLAB codes by using the finite-difference time-domain (FDTD) method.

The SAR values for 900 MHz were found to between 1.2-3.00 mW/kg.
 The SAR values for 1800 MHz were found to be 0.011-0.053 mW/kg.

I would like to thank Assist. Prof. Dr. Mesut Kahraman from Suleyman Demirel University, Electronic and Communication Engineering Department because of supplying us the software used in the calculation of SAR value.

Clinical evaluation

The weight (by using precision scales) and height (using a ruler) of the rats were measured weekly from 2 days to 90 days of age.

Clinical observations such as:

Joint and bone development,

Joint movements,

Presence or absence of lameness and mobility values

were performed during periods of growth of rats.

Biochemical evaluation

Blood samples (1 ml) were drawn from the right ventricle of the heart under ether anesthesia from all animals at the end of the third month.

Plasma samples were obtained by centrifugation at 3000 rpm for 10 min and stored at –20 °C.



• Estradiol					
• Testosterone					
$m \cdot$ Growth hormone (GH)					
	• calcium (Ca)				
	• phosphorus (P)				

Radiographic evaluation



The animals were premedicated using xylazine and anesthetized with ketamine

Radiographs of the distal femoral growth plates of rats from each group were taken at the end of the first, second, and third months.



Radiographs were obtained in the antero-posterior and mediolateral projections.

Histopathological evaluation

- The right femurs of the rats were ablated, including epiphyseal growth plates.
- The bones were kept in a formaldehyde solution of 10% buffer for 24 h,
- Decalcified with formic acid-paraformaldehyde solution for 4 weeks.
 - The tissues were processed with alcohol and xylol series
 - They were blocked in paraffin

• The 5 mm-cross-sections provided by microtome were stained with hematoxylin eosin, Safranin O and toluidine blue.

These cross-sections were scored with Modified Mankin Method

Results Clinical evaluation

There were no apparent differences between the rats with regard to joint functions, and general mobility.



Weeks Length (cm) Groups Weight (g) Week_1 Control 12.27 ± 0.82^{a} 6.53 ± 0.09 $16.00 + 0.77^{b}$ 1800 MHz 6.62 ± 0.07 900 MHz 15.73 ± 0.62^{b} 6.70 ± 0.16 Week 2 Control 24.36 ± 1.74^{a} 7.78 ± 0.17^{a} 1800 MHz 30.27 ± 0.97^{b} 8.32 ± 0.12^{b} 900 MHz 26.45 ± 1.32^{a} 8.14 ± 0.21^{a} $39.09 + 2.54^{a}$ Week 3 Control 10.01 ± 0.25^{a} 1800 MHz 47.64 ± 1.65^{b} 19.78 ± 9.02^{b} 900 MHz $44.00 + 2.10^{b}$ 10.57 ± 0.19^{a} Week 4 Control 69.73 ± 3.41 11.54 ± 0.22^{a} 1800 MHz 76.00 ± 2.28 12.43 ± 0.19^{b} 74.82 ± 2.72 12.02 ± 0.22^{b} 900 MHz Week 5 Control 12.80 ± 0.26^{a} 94.91 + 4.561800 MHz 13.62 ± 0.31^{b} 106.00 ± 2.90 900 MHz 13.27 ± 0.22^{b} 105.09 ± 3.48 Week 6 Control 103.64 ± 6.90 $13.30 + 0.23^{a}$ 1800 MHz 111.55 ± 3.68 14.13 ± 0.17^{b} 900 MHz 111.36 ± 5.52 14.01 ± 0.24^{b} Week 7 Control 126.18 ± 4.94^{a} 14.95 ± 0.15^{a} 1800 MHz 161.55 ± 5.39^b 15.49 ± 0.12^{b} 900 MHz 162.09 ± 5.13^{b} 15.86 ± 0.16^{b} Week 8 Control 149.64 ± 7.18^{a} 15.15 ± 0.24^{a} $196.73 + 6.82^{b}$ 16.14 ± 0.25^{b} 1800 MHz 900 MHz 201.73 ± 7.28^{b} 16.45 ± 0.24^{b} Week 9 Control 169.45 ± 7.80^{a} 15.23 ± 0.25^{a} 213.18 ± 6.27^{b} 1800 MHz 16.90 ± 0.18^{b} 16.93 ± 0.25^{b} 900 MHz 217.36 ± 8.14^{b} Week 10 Control 191.00 ± 6.62^{a} 16.59 ± 0.18^{a} 1800 MHz 234.73 ± 5.20^{b} 17.88 ± 0.19^{b} 900 MHz 236.73 ± 7.73^b $18.39 \pm 0.18^{\circ}$ Week 11 Control 182.64 ± 5.68^{a} 17.33 ± 0.26^{a} 1800 MHz $241.09 + 5.25^{b}$ 18.65 ± 0.15^{b} 900 MHz 241.82 ± 7.36^b 18.65 ± 0.15^{b} Week_12 Control 143.55 ± 5.06^{a} 17.23 ± 0.28^{a} $228.55 + 7.21^{b}$ 19.10 ± 0.19^{b} 1800 MHz 900 MHz 228.18 ± 7.66^b 19.47 ± 0.20^{b}

Weekly increase in length and weight of the EMF and control groups.

^{a,b,c}Significantly different at the same time point (p < 0.05).



Rats in the EMF group experienced a faster increase in weight and length.

Biochemical evaluation

Calcium, phosphorus, growth hormone, estradiol and testosterone hormone levels were higher in the EMF groups than in the control group (p < 0.05).

However,

there were no differences between the EMF groups

Groups	Ca (mg/dl)	P (mg/dl)	Growth H (ng/ml)	Estradiol (ng/ml)	Testosterone (ng/ml
900 MHz	23.98 ± 2.14^{a}	9.66 ± 1.68	0.54 ± 0.01^{a}	36.19 ± 0.03^{a}	6.01 ± 0.03^{a}
1800 MHz	24.31 ± 2.24^{a}	8.98 ± 1.21	0.53 ± 0.00^{a}	36.22 ± 0.04^{a}	5.99 ± 0.00^{a}
Control	21.78 ± 0.68^{b}	8.38 ± 1.69	0.49 ± 0.03^{b}	36.14 ± 0.03^{b}	5.73 ± 0.14^{b}

Radiological evaluation

Radiological examinations conducted at the end of the third month revealed that distal femoral growth plates were no longer visible in all three groups.



Radiographic view of the left femurs of rats after 90 days of EMF exposure, ML position. A. 900 MHz group, B. 1800 MHz group, C. control group.

Histopathological evaluation

Modified Mankin scoring scale showed that there were no significant differences between the groups with respect to cellularity in the reserve, proliferative and hypertrophic zones of the femoral growth plate.

Cellularity and sta	aining propert	ies of femora	growth p	olate.
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Groups	Cellularity-reserve zone	Cellularity-proliferative zone	Cellularity-hypertrophic zone	Staining-reserve zone	Staining-proliferative zone	Staining-hypertrophic zone
900 MHz 1800 MHz Control	$\begin{array}{c} 2.00 \pm 0.00 \\ 1.81 \pm 0.40 \\ 2.18 \pm 0.40 \end{array}$	$\begin{array}{c} 2.00 \pm 0.00 \\ 1.81 \pm 0.40 \\ 2.18 \pm 0.40 \end{array}$	$\begin{array}{c} 2.00 \pm 0.00 \\ 2.00 \pm 0.44 \\ 2.36 \pm 0.50 \end{array}$	$\begin{array}{c} 2.54 \pm 0.82^{a} \\ 1.54 \pm 0.68^{b} \\ 2.63 \pm 0.50^{a} \end{array}$	$\begin{array}{l} 2.54 \pm 0.68^a \\ 2.00 \pm 0.77^a \\ 2.72 \pm 0.46^b \end{array}$	$\begin{array}{c} 2.63 \pm 0.67 \\ 2.27 \pm 0.78 \\ 2.72 \pm 0.46 \end{array}$

Mean \pm SD, p < 0.05.

In Safranin O staining for mature proteoglycans of growth plate, the cartilage matrix density, was lowest in the reserve zone at 1800 MHz (p < 0.05) and was significantly increased in the proliferative zone of the control group (Fig. a).



Fig. a) Microscopical appearance of femoral growth plate at 1800MHz group. There are partial losses in matrix staining of the reserve zone (arrows) with an increase in thickness of the reserve and proliferative zones. Note a decrease of thickness of the trabecular zone with the thin and irregular spicules (arrowheads). Safranin O stain. Bar=50 µm.

Fig. b) The femoral growth plate of control group. The intense staining areas for cartilage matrix in the proliferative zone (arrows). Note thick and regular bone spicules in trabecular zone (arrowheads) compared to 1800 MHz group (a). Safranin O stain. Bar = 50 μ m.

Experimental studies have shown that high frequency EMF has an effect on endocrine and nervous systems (Hardell and Sage, 2008).

This study evaluated the long-term effects of 1800 MHz and 900 MHz EMF on growing healthy rat bones, using clinical, radiological, histopathological and biochemical analyses.







 Clinical observations revealed no significant difference

in joint and bone growth nor in joint and
other mobilities of the rats.

However, it was found that rats in the
EMF groups
more rapidly gained weight and increased in length compared with the control group. In light of these results, it is believed that 1800 MHz and 900 MHz EMF may cause prolong the growing process in growing rats.





Considering the results of the study, further studies by including before- and after-puberty periods at even longer time intervals are necessary to clarify the mechanisms of EMF. This work has been supported by the grants from the Ondokuz Mayis University Scientific Projects Unit (BAP.PYO.VET.1901.005).

