In The Name of God

Fecal carriage of carbapenem resistance Enterobacteriaceae among inpatients in university hospital in Iran

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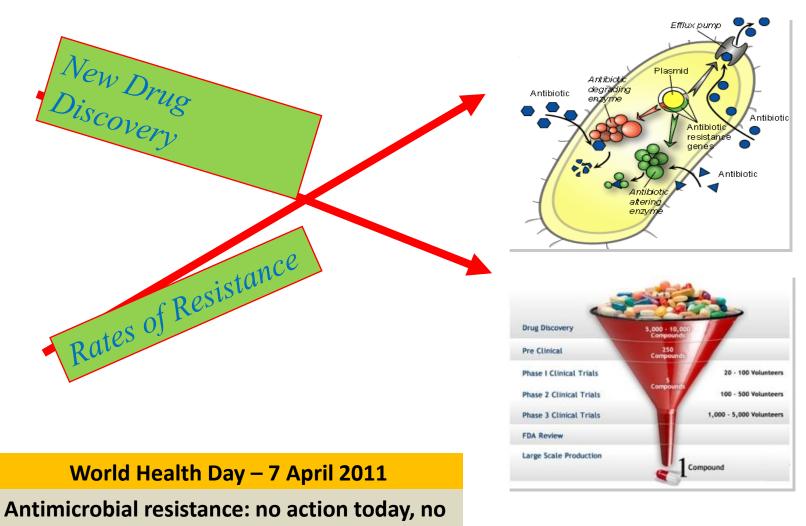
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Antibiotic resistance is a global problem, but the solutions are at the national and regional level. The benefits of conservation efforts accrue locally while contributing to antibiotic effectiveness at the global scale.

We Have a Basic Problem



cure tomorrow

1. Reduce the need for antibiotics through improved water, sanitation, and immunization

2. Improve hospital infection control and antibiotic stewardship

3. Change incentives that encourage antibiotic overuse and misuse to incentives that encourage antibiotic stewardship

4. Reduce and eventually phase out subtherapeutic antibiotic use in agriculture

5. Educate health professionals, policy makers, and the public on sustainable antibiotic use

6. Ensure political commitment to meet the threat of antibiotic resistance

FIGURE 5-1: Six strategies needed in national antibiotic policies











WHO priority pathogens list for R&D of new antibiotics (2017)

Priority 1: CRITICAL

- Carbapenem-resistant Enterobacteriaceae
- Acinetobacter baumannii, carbapenem-resistant
- Pseudomonas aeruginosa, carbapenem-resistant

Priority 2: HIGH

- Enterococcus faecium, vancomycin-resistant
- Staphylococcus aureus, methicillin-resistant, vancomycin-intermediate and resistant
- Helicobacter pylori, clarithromycin-resistant
- Campylobacter spp., fluoroquinolone-resistant
- Salmonellae, fluoroquinolone-resistant
- Neisseria gonorrhoeae, cephalosporin-resistant, fluoroquinolone-resistant

Priority 3: MEDIUM

- Streptococcus pneumoniae, penicillin-non-susceptible
- Haemophilus influenzae, ampicillin-resistant
- Shigella spp., fluoroquinolone-resistant



Our projects:

 Evaluation of phenotypic and molecular epidemiology of carbapenemase producing Enterobacteriaceae isolated from carriers and clinical samples.

 Study of phenotypic, genotypic characteristics and molecular epidemiology of predominant isolates of ESBL producing Enterobacteriaceae family (EPE) isolated from carriers and clinical samples.

Our projects:

 Molecular typing of carbapnemase producing Enterobacteriaceae isolates and determine the genomic structure of plasmids carrying carbapenemase genes.

Our projects:

• The aim of this study was to evaluate the

prevalence and risk factors associated with

CRE fecal colonization among inpatients.

Two main approaches to prevent the spread of CRE

- Detection of infected patients
- Detection of carriers

How does a CRE patient present?

□ Infection versus Colonization

□ A patient with CRE can be colonized and/or infected.

Why detecting rectal colonized patients with CPE is so important?

- Fecal carriage with CPE is a marker for infection with CPE
- nearly 10% of patients with positive CPE rectal carriage are latter positive in a clinical sample (85% being true infection) - 11 days median time interval (range 3-27 days) between positive rectal screening and positive clinical specimen
- **prevent transmission** of the resistant bacteria in community and hospital
- **prevent** the development of **nosocomial** outbreaks due to the multidrug-resistant bacteria
- Screening test for surveillance program

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		ESBL Ø
The human gut microbiome as a transporter of antibiotic resistance genes between continents	This Article Accepted manuscript posted online 10 August 2015, doi: 10.1128/AAC.00933-15 AAC.00933-15	Current Issue September 2015, Volume 9
Johan Bengtsson–Palmeª, Martin Angelinb, Mikael Huss¢, Sanela Kjellqvist¢, Erik Kristiansson ^d , Helena Palmgrenb, D.G. Joakim Larssonª and Anders Johansson ^{#,} e	 » Abstract PDF Article Usage Stats 	ANTIMICROBIAL AGENTS AND CHEMOTHERAP
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ABSTRACT Previous studies of antibiotic resistance dissemination by travel have, by targeting only a select number of cultivable bacterial species, omitted most of the	Email this article to a colleague Similar articles in ASM journals Alert me when this article is cited Alert me if a correction is posted Similar articles in this journal	AAC
human microbiome. Here, we used explorative shotgun metagenomic sequencing to address the abundance of >300 antibiotic resistance genes in fecal specimens	Similar articles in PubMed Alert me to new issues of AAC	About AAC
from 35 Swedish students taken before and after exchange programs on the Indian peninsula or in central Africa. All specimens were additionally cultured for	Download to citation manager Reprints and Permissions Copyright Information	Subscribers
extended-spectrum beta-lactamase (ESBL) producing enterobacteria and the	Books from ASM Press Microbe/Vorld	Authors
isolates obtained genome sequenced. The overall taxonomic diversity and composition of the gut microbiome remained stable comparing before and after	+ Google Scholar	Reviewers
travel, but with increasing abundance of Proteobacteria in 25/35 students. The relative abundance of antibiotic resistance genes increased, most prominently for	+ PubMed + Social Bookmarking	Advertisers
genes encoding resistance to sulfonamide (2.6–fold increase), trimethoprim (7.7– fold) and beta–lactams (2.6–fold). Importantly, the increase observed occurred		Inquiries from the Press
without any antibiotic intake. Of 18 students visiting the Indian peninsula, 12		Permissions & Commer

What specimens?

• stools and rectal swabs are the most suitable specimens for performing this screening process.

• It is imperative to point out that the screening process on admission still requires the patients to be kept in strict isolation prior to results being obtained (at least for 48 h).

Who must be screened?

- Screening should include at least **'at-risk'** patients, such as those in **intensive care units**, and transplantation and immunocompromised patients.
- If a patient is confirmed as being infected or colonized by a carbapenemase producer, the screening program should be extended to **neighboring patients** on the hospital ward.
- Screening shall be done at least to patients transferred from a **foreign hospital** on addition to any hospital.

Does everyone who gets the bug become sick?

- No, some people are colonized.
- Patients with unrecognized colonization with CPE have served as reservoirs for transmission

during outbreaks.



Screening of Carriers

• The prevention of spread of carbapenemase producers relies on **early** and **accurate** detection of carriers in hospital units or on admission/discharge either to the hospital or to a specific unit.

Why Enterobacteriaceae?

- Source of community- and hospital-acquired infections.
- ≻ Normal gut flora
- They have the propensity to spread easily between humans (hand carriage, contaminated food and water, medical equipment)
- Acquire genetic material through horizontal gene transfer, mediated mostly by plasmids and transposons.



Why Carbapenemases:

- Carbapenemases are a source of concern because :
- **✓** They confer resistance to essentially all β -lactams
- Strains producing carbapenemases often possess resistance mechanisms to a wide-range of antimicrobial agents
- Carbapenemases increasingly have been reported in Enterobacteriaceae in the past 10 years
- > Infections with **CPE** are associated with **high mortality rates**
- No vaccines are readily available for preventing infections with carbapenemase producers



Carbapenemases: the triple difficulty

Difficulty of detection in the clinic and in the laboratory

- Difficulty of treatment (owing to their MDR, XDR, PDR character)
- Difficulty to limit transmission and spread and to control outbreak (local, regional, national, pandemic)

Methods Phenoty Suscept Conjugat pic Detectio Identifi ibility n of detectio ion Data cation testing Sampli n of carbape experim PFGE collecti of CRE and carbape nemase ents ng colonie MIC on nemase genes determ S producti (PCR) ination on

Sampling

- Hospital based cross-sectional study
- July to November 2015
- Rectal swab specimens were collected from 95 inpatients
- Informed consent was obtained from all participants
- Swab was inserted 2 to 3 cm into the rectum
- Specimens were inoculated immediately in TSB (5mL) containing a 10 µg ETP disk

Data collection

Clinical and epidemiologic data were collected from all inpatients

> The following data were recorded:

✓ Age

✓ Sex

- ✓ Unit of hospitalization
- ✓ Invasive medical device utilization
- ✓ History of surgery, presence of wounds
- ✓ Transfer from another hospital
- ✓ Transfer between hospital units
- \checkmark Exposure to antibiotics

Data Collection

Case details	فت ۵	sing swabs: Al-Zahra	Contraction of the second
Number of file $\sim 5^{<}$	20199	Age	7.8
Male sex	M	Ward	Jack 1
Prior antimicrobial use	An stir	Antimicrobial treatment	المريب الولاع كم مترزار ال - كموكر اليلم - مقلور م
Previous surgery	Y 25 - 1 - 1 - 1 - 1 - 1 - 1 - 1 - 1 - 1 -	Presence of wounds	~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~
Type of infection	- 21 . 	Mechanical ventilation	tus
Long (>2 weeks) acute-care hospital stay in past 6 months		Prior hospital stay < 1 year prior	
Transfer between hospital units	2 (1):1' - Us (4) -	Cross-border transmission by direct hospital-to-hospital transfer	المناه در در ما میلات
Direct transfer of patients from a foreign hospital	-	Positive clinical samples	
History of Urinary tract infection in past 6 months		History of Invasive device usage in the past 3 month	21×- in -
Date of admission	25,0,00	Date of discharge or death	97,0,71
T'ALS -	الرسارى مزمز تنقسى سقع		+ AIQ YT, NOLV

CRE screening

- We used two different phenotypic methods for detection of CRE in rectal swab specimens
- Method 1, MacConkey agar (CDC)
- Method 2, CHROMagar KPC medium
- Isolates were confirmed by using standard biochemical tests and API 20E



CRE screening in rectal swab specimens

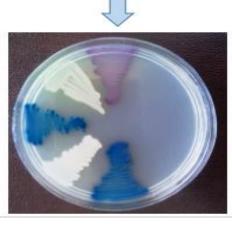


MacCon key agar+ MEM

Sample

Select TSB + carbapenem disk Differentiate

CHROMa gar KPC



Susceptibility testing and MIC determination (CLSI)

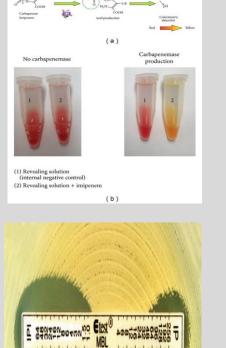
- Disk diffusion method on Muller-Hinton agar plates
- ✓ Imipenem, Meropenem, Ertapenem, ceftazidime, cefotaxime, cefepime, ciprofloxacin, amikacin, gentamicin, aztreonam and tigecycline (MAST)
- Minimal inhibitory concentrations (MICs) of IMP, MEM and ETP were determined by gradient test strips
- ✓ *E. coli* ATCC 25922 was used as control strains

Phenotypic detection of carbapenemase production

- □ Modified Hodge test (MHT)
- Combined disk test (CDT)

✓ EDTA → MBLs

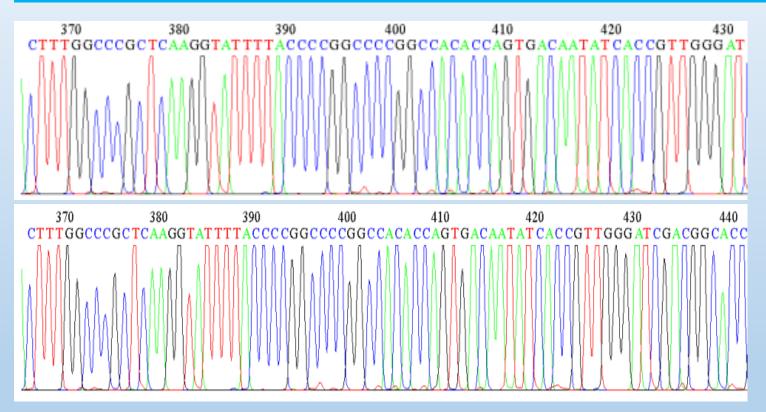
- □ MBL-E-test
- 🗖 Carba NP







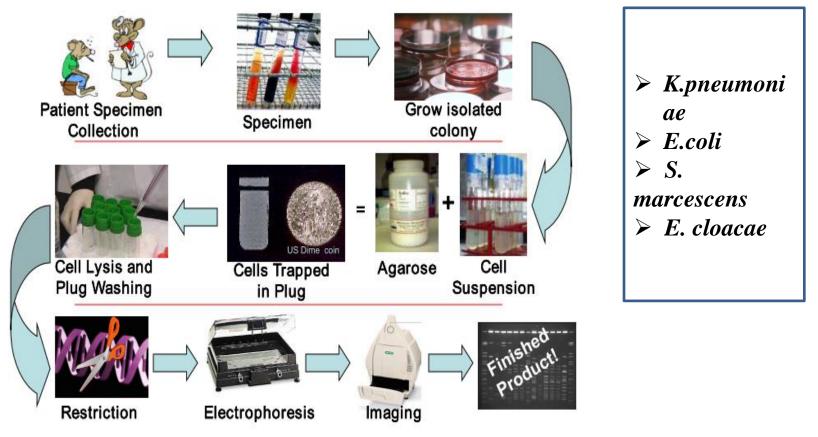
Nucleotide sequences and GenBank accession numbers



≻ KX467530, KX467529 and KX671151

Pulsed-field gel electrophoresis (PFGE)

PFGE Process



MLST

✓ K. pneumoniae

• *gapA*, *infB*, *mdh*, *pgi*, *phoE*, *rpoB* and *tonB*

http://www.pasteur.fr/recherche/genopole/PF8/mlst/Kpneumoniae.html

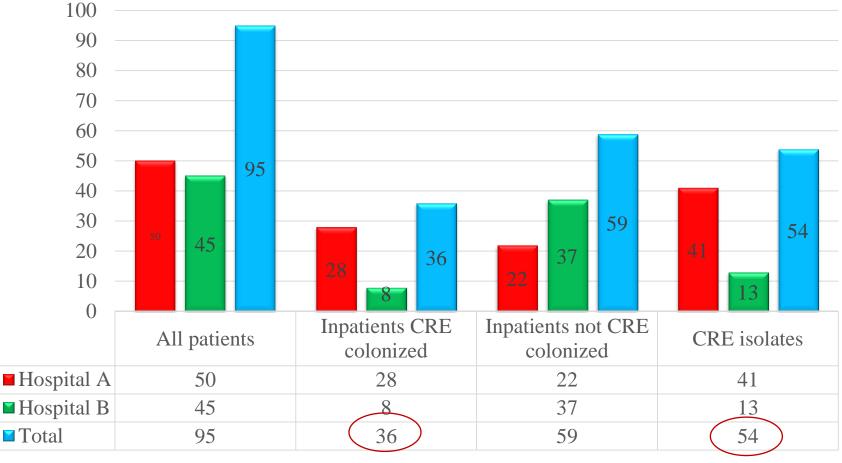
✓E. coli

• *adk*, *fumC*, *gyrB*, *icd*, *mdh*, *purA*, and *recA*

http://mlst.warwick.ac.uk/mlst/dbs/Ecoli



Rate of colonization of inpatients by CRE in two Al-Zahra and Loghman hospitals



Hospital A Hospital B Total

The isolates recovered from 95 rectal samples using 2 different methods

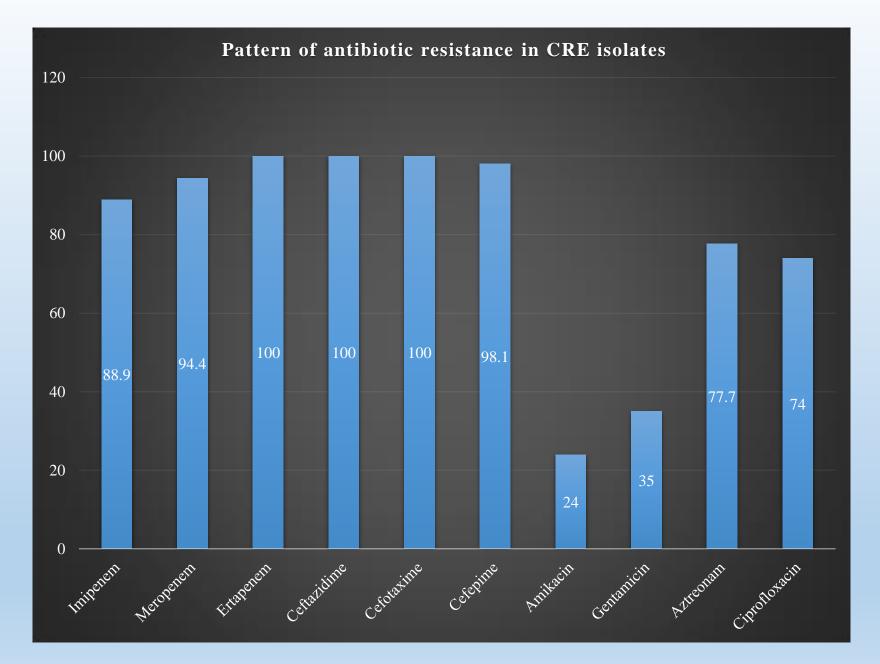
CRE isolates	CHROMagar KPC		MacConkey agar	
	Total	Carbapenemase producing	Total	Carbapenemase producing
K. pneumoniae	33	29	33	29
E.coli	19	16	18	15
E.cloacae	1	1	1	1
P.mirabilis	1	1	-	-
Total isolates	54	47	52	45

 \triangleright rate of carriage was high (37.9%)

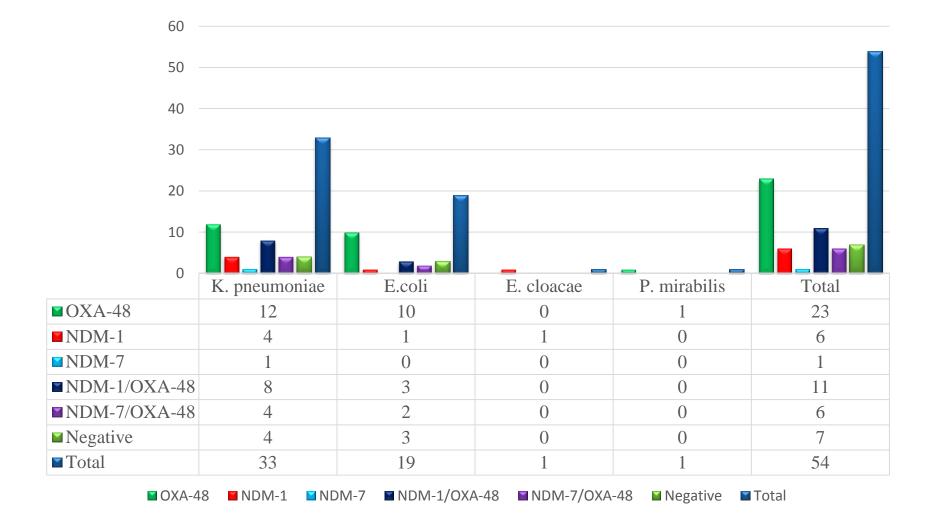
 \triangleright A higher proportion (28/36; 77.8%) of colonization with CRE isolates was identified among admitted patients in the hospital A compared to hospital B (8) colonized patients)

 \succ CRE multiple colonization \longrightarrow 15 patients





Distribution of carbapenemase genes in CRE isolates recovered from carriers in both hospitals.



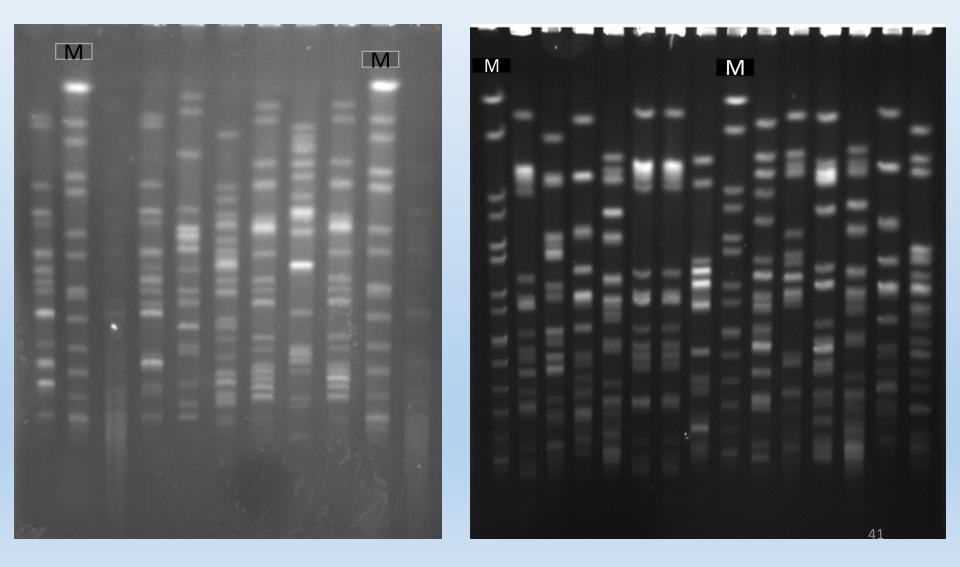
Characteristics of the 54 CRE isolated from intestinal carriage

Patient	Species	Ward/Hospital	Data of isolation (day/month)	MHT	<u>Etest</u> - MBL	CDT		MICs (µg/ml)			Carbapenemases	PFGE
						MRP/BO	MRP/EDTA	ETP	MRP	IMP	-	<u>clasters</u>
P1	K. pneumoniae	ICU-2/ HA	12/10/2015	+	+	-	+	8	8	12	NDM-7, OXA-48	Cluster-I
P2	K. pneumoniae	ICU-2/ HA	12/10/2015	+	+	-	+	8	8	12	NDM-1	Cluster-III
P3	K. pneumoniae	ICU-2/ HA	12/10/2015	+	+	-	+	8	8	16	NDM-7, OXA-48	Cluster-I
	E.coli	ICU-2/ HA	12/10/2015	+	+	-	+	8	8	8	NDM-7, OXA-48	singleton
P4	K. pneumoniae	ICU-2/ HA	12/10/2015	+	+		+	8	8	12	NDM-7, OXA-48	
P5	K. pneumoniae	ICU-2/ HA	12/10/2015	-	+	-	+	8	8	64	NDM-7	singleton
	E.coli	ICU-2/ HA	12/10/2015	+	+	-	+	1	0.5	<4	OXA-48	singleton
P6	K. pneumoniae	ICU-2/ HA	12/10/2015	+	+	-	+	8	1.5	<4	OXA-48	Cluster-I
P7	E.coli	ICU-2/ HA	12/10/2015	+	-	-	-	ND	ND	ND	-	-
P8	K. pneumoniae	ICU-2/HA	12/10/2015	+	ND	+	+	ND	ND	ND	-	-
P9	K. pneumoniae	ICU-2/HA	12/10/2015	+	ND	+	+	ND	ND	ND	-	-
	E.coli	ICU-2/ HA	12/10/2015	+	+	-	+	8	8	12	NDM-1, OXA-48	singleton
P10	K. pneumoniae	ICU-2/ HA	15/10/2015	+	+	+	+	8	8	8	OXA-48	Cluster-I
P11	E.coli	ICU-2/ HA	15/10/2015	+	+	-	+	8	8	12	NDM-7, OXA-48	Cluster-I
P12	K. pneumoniae	ICU-2/ HA	15/10/2015	+	-	+	-	8	8	8	OXA-48	Cluster-I
	K. pneumoniae	ICU-2/ HA	15/10/2015	+	+	-	+	8	8	8	OXA-48	Cluster-I
P13	K. pneumoniae	ICU-2/HA	12/10/2015	+	+	-	+	8	8	256	NDM-1, OXA-48	Cluster-IV
	E.coli	ICU-2/ HA	15/10/2015	+	-			1	0.25	<4	OXA-48	singleton
P14	K. pneumoniae	ICU-2/ HA	15/10/2015	+	+	-	+	8	8	8	OXA-48	Cluster-I
	E.coli	ICU-2/ HA	15/10/2015	+	+	-	+	8	8	32	OXA-48	Cluster-II
P15	K. pneumoniae	ICU-1/ HA	17/11/2015	+	+	+	+	8	8	16	NDM-1	Cluster-III
P16	K. pneumoniae	ICU-1/ HA	17/11/2015	+	+	-	+	8	8	12	NDM-7, OXA-48	singleton
P17	K. pneumoniae	ICU-1/ HA	17/11/2015	+	+	-	+	8	8	256	NDM-1, OXA-48	Cluster-I

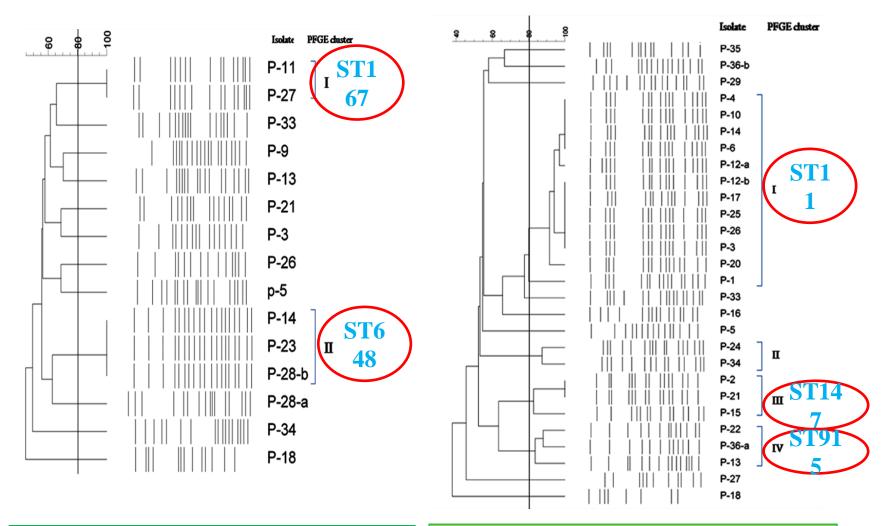
		,										
P18	K. pneumoniae	ICU-1/ HA	17/11/2015	+	+	-	-	8	8	12	NDM-1, OXA-48	singleton
	E.coli	ICU-1/ HA	17/11/2015	+	+	-	-	1	0.25	<4	OXA-48	singleton
P19	E.coli	ICU-1/ HA	17/11/2015	+	+	-	+	8	8	12	NDM-1, OXA-48	
P20	K. pneumoniae	ICU-1/ HA	17/11/2015	+	+	-	+	8	8	8	OXA-48	Cluster-l
P21	K. pneumoniae	ICU-3/ HA	17/11/2015	+	+	-	+	8	8	12	NDM-1, OXA-48	Cluster-III
	E.coli	ICU-3/ HA	17/11/2015	+	+	-		8	8	16	NDM-1, OXA-48	singleton
P22	K. pneumoniae	ICU-2/ HA	17/11/2015	+	+	-	+	8	8	256	NDM-1, OXA-48	Cluster-IV
P23	E.coli	ICU-3/ HA	17/11/2015	+	+	-	+	8	8	16	OXA-48	Cluster-II
P24	K. pneumoniae	ICU-3/ HA	17/11/2015	+	+	+	-	8	8	8	NDM-1, OXA-48	Cluster-II
P25	K. pneumoniae	ICU-3/ HA	17/11/2015	+	+		+	8	8	8	OXA-48	Cluster-I
P26	K. pneumoniae	ICU-3/ HA	17/11/2015	+	+	-	+	8	8	8	OXA-48	Cluster-l
	E.coli	ICU-3/ HA	17/11/2015	+	-	-	-	0.25	0.25	<4	OXA-48	singleton
	P.mirabilis	ICU-3/ HA	17/11/2015	+	+	-	+	8	8	48	OXA-48	
P27	K. pneumoniae	ICU-2/ HA	17/11/2015	+	+	+	-	8	2	<4	OXA-48	singleton
	K. pneumoniae	ICU-2/ HA	17/11/2015	+	+	+	-	8	8	6	OXA-48	•
	E.coli	ICU-2/ HA	17/11/2015	+	+			8	1.5	<4	OXA-48	singleton
P28	E.coli	ICU-2/ HA	17/11/2015	+	+	+	-	8	8	6	OXA-48	Cluster-II
	E.coli	ICU-2/ HA	17/11/2015	+	+		+	8	8	6	OXA-48	singleton
P29	K. pneumoniae	ID/HB	1/7/2015	+	+	-	+	8	8	16	NDM-1, OXA-48	singleton
P30	E.coli	ID/ HB	1/7/2015	-	ND	+	-	ND	ND	ND	-	•
P31	K. pneumoniae	G- ICU/ HB	18/6/2015	-	ND	+	-	ND	ND	ND	•	•
	K. pneumoniae	G- ICU/ HB	18/6/2015	•	ND	+	-	ND	ND	ND	•	•
P32	E.coli	E- ICU/ HB	31/7/2015	-	ND	+	-	ND	ND	ND	-	•
P33	K. pneumoniae	G- ICU/ HB	1/8/2015	•	+	-	+	8	8	8	NDM-1	singleton
	E.coli	G- ICU/ HB	1/8/2015	+	+	-	+	8	8	8	NDM-1	singleton
	E.cloacae	G- ICU/ HB	1/8/2015	+	+	-	+	8	3	4	NDM-1	
P34	K. pneumoniae	G- ICU/ HB	20/9/2015	+	+	-	+	8	8	16	NDM-1, OXA-48	Cluster-II
	E.coli	G- ICU/ HB	20/9/2015	+	+	+	-	8	1.5	<4	OXA-48	singleton
P35	K. pneumoniae	ID/ HB	18/9/2015	+	•	+		8	8	6	OXA-48	singleton
P36	K. pneumoniae	E- ICU/ HB	19/10/2015	+		+		8	8	<4	OXA-48	Cluster-IV
	K. pneumoniae	E- ICU/ HB	19/10/2015	-	+		+	8	8	256	NDM-1	singleton

PFGE *E.coli*

PFGE K. pneumoniae



The clonal relatedness among *E.coli* and *K. pneumoniae* isolates recoverd from carriages

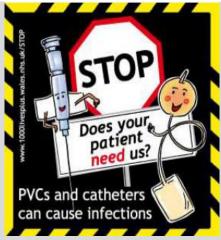


- PFGE dendrogram in 15 CP *E.coli*
- PFGE dendrogram in 28 CP K. pneumoniae

Discussion

Risk Factors for CRE colonization

- ✤ Admission to ICU
- ▶ mainly ICU-2 (p=0.00004) and general ICU (p=0.007)
- **♦** Surgery (p=0.03)
- ♦ Ventilation (p=0.0004)
- ✤ Urinary catheter (p=0.04)
- Antibiotic exposure
- mainly third-generation cephalosporins (p=0.00001)
- ✤ Transfer between hospital units (p=0.008)
- ✓ Zhao et.al in 2014: China
- ✓ Torres et al in 2015: Mexico





Discussion

- The predominant species found in our study was *K. pneumoniae*, followed by *E. coli*
- The bla_{OXA-48} was the most frequently detected carbapenemase and bla_{NDM-1} was the second rank
- Both carbapenemase producing *K. pneumoniae* and *E.coli* cluster I strains were isolated among inpatients who shared a room hence PFGE profile of the strains was identical. Therefore, it is possible that a spread of CPE from patient to patient occurred.

Discussion: Carriage

- Mainly hospitalized patients in the ICUs were screened
- Rate of carriage was high (37.9%)
- ✓ Higher than in a 2012 report from Greece (12.8) as well as one from Korea (0.3)
- Hand hygiene non-compliance, breaches in environmental sanitation in all hospital areas (especially in the ICU), increased duration of hospitalization and extensive use of broad-spectrum antimicrobial agents
- CRE multiple colonization in the same patient (15 patients)
- ✓ Intra- and inter-species transmission of *bla*_{NDM} (IncA/C, IncFII) and *bla*_{OXA-48} (IncL/M) genes within the gut microbiome.
- Rapid identification of CRE carriages could be an important strategy to control the transmission of these organisms in healthcare facilities

Conclusion

- Clonal dissemination and outbreak of OXA-48 and NDM-1 producing *K*. pneumoniae has been observed.
- ≻ ST11, ST893, ST147
- ★ Detection of high risk clones (ST11 *K. pneumoniae* and ST131 E. coli)
- ➢ high capacity to colonize and persist over time
- Co-expression of carbapenemase genes together reduces the options of treatment
- * Alert on the large dissemination of these genes to other hospitals and community
- □ So, we must therefore focus on rapid identification of CPE colonized patients and implementation effective infection control measures.

Conclusion: Other CRE isolates

Serratia marcescens

- \checkmark 5 isolates collected from Tehran
- \checkmark Probably the routes of transmission from patient to patient are either by:
- 1. direct contact through carriage of CRE on the hands of HCWs
- 2. indirectly via contaminated environmental surfaces or shared equipm

> Proteus mirabilis

✓ 4 isolates collected from Esfahan

Enterobacter cloacae

- ✓ 2 isolates collected from Tehran
- The potential dissemination of *P. mirabilis* and *S. marcescens* OXA-48 producer isolates is a major problem, because this organism is intrinsically resistant to colistin

Conclusion

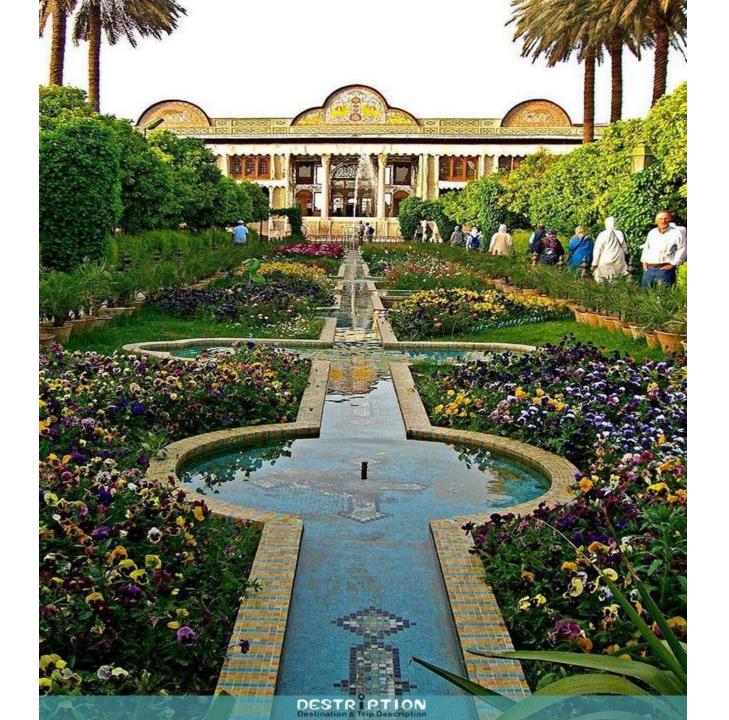
- Our study, provides novel information about the presence and distribution of carbapenemases among the CRE isolates in Iran
- ✤ High rate of CRE intestinal colonization and CRE infection among inpatients:
- 1. Might be related to the close relationship between Iran and neighbor countries such as Pakistan, Turkey, Afghanistan and Iraq in terms of population exchanges
- 2. The lack of knowledge of hospital personnel about these bacteria

Prevention

• An aggressive infection control and prevention strategy is recommended, including reinforcement of hand hygiene, using contact precautions and early detection of ESBL-E and CPE through use of ⁵⁰targeted surveillance







Thank you for your attention