Mechanisms of cell death in eukaryotic microorganism: An in-depth overview

Fatimah Binti Hashim,

School Of Fundamental Science, Universiti Malaysia Terengganu, Terengganu, MALAYSIA

- Eukaryotic microorganisms- shared similar characteristics with mammalian cells,
- typical plasma membrane,
- organelles,
- life cycle progression
- death pathway mechanisms,
- single living cell- life cycle is shorter than mammalian cells.
- Pathogenic species of eukaryotic microorganism such as Leishmania, Plasmodium, Trypanosome, Toxoplasma have been studied for many years for their potential antimicrobial therapy.
 - The mode of drug action and cell death crucial as treatment for this disease wouldn't harm the host.











Infections inflicted in man by Acanthamoeba

- Systemic infection (brain, skin, CNS)
- Ocular infection Acanthamoeba keratitis (AK)
 - "... 85 % of AK cases is from contact lens wearers.." (Murdoch et al., 1998)
 - ".....Often misdiagnosed as bacteria, fungi keratitis..." (Yun et al., 2006)
 - ".....Delayed treatment can cause blindness...." (Khan, 2003)



Current treatment for Acanthamoeba keratitis

- Chlorhexidine gluconate (CHX) and Polyhexamethelene biguanide (PHMB), effective in killing *Acanthamoeba* spp. (Khan, 2003)
- CHX
 - ".... more effective than PHMB..." (Noble, 2002)
 - "....has been recommended for many Acanthamoeba keratitis
 - treatment..." (Hay et al., 1994)
 - ".... induced membrane disruption in bacteria and yeast...." (Russel and Hugo, 1988)
 - ".... able to irritate to the eye, ulcerative keratitis, swollen cornea nuclei, alteration to the cell membrane and cytolysis... " (Masamichi et al., 2005)
- Alternative treatment surgical excision of the infected cornea
- Search for drug with low toxicity is essential!
- Apoptosis and autophagic cell death mechanisms the safest way for the diseased cell or parasite should die.



Cytotoxicity in drug discovery

- Acanthamoeba Euchromatin, mitochondria and cell membrane
- Cytotoxicity and mode of cell death determination is important in cancerous cell
- The important of cytotoxicity assay-
 - -to determine the cell killing property of chemical compound or mediator cell (Doris et al., 1988), confirmed that the drugs can be used or not
- IC₅₀ value- allows the comparison of chemical necessary to inhibit any measurable parameter (cell proliferation, protein synthesis, DNA synthesis etc.)
- Membrane integrity, chromatin pattern, DNA cleavage
- Cell death : Apoptosis (Programmed cell death) and Necrosis
- Cytotoxicity in this study -Drug search for pathogenic amoeba:
- This study focused only on cytoxicity and genotoxicity effects of the MH, GR and CHX on trophozoite of Acanthamoeba spp.

Cultivation and maintenance of the test Organisms
Determination of <i>Acanthamoeba</i> Population Doubling Time (Freshney et al. 1987)
Preparation of MH, GR and CHX stock solution
Cell viability and IC ₅₀ values determination by Eosin Dye Technique Assay (Wright et al., 1988) after treatment with MH and GR.
Observation on Morphology of <i>Acanthamoeba</i> spp: after treatment with MH and GR. - By Light Microscopy - By SEM - By TEM

Results and Discussion

MATERIALS AND METHODS

Test organisms and cultivation

No.	Species	Isolate	Source	Pathogenicity
1	Acanthamoeba castellanii	IMR	Unknown	Pathogen
2	Acanthamoeba polyphaga (Puschkarew) Volkonsky 1931	CCAP 1501 /3A	Freshwater, Winsconsin USA	Potential pathogen
3	Acanthamoeba sp.	HKL	Corneal scrapping from a keratitis patient (Hospital Kuala Lumpur)	Pathogen
4	Acanthamoeba sp.	SW	Freshwater, Setiu Wetland	Not known

(Sub-cultured every 3 to 5 days and were maintained in 30 °C incubator)

Fifty percent Inhibition Concentration (IC₅₀) Values for MH and GR Determined by Eosin Dye Technique Assay



Fifty percent Inhibition Concentration (IC₅₀) Values for CHX Determined by Eosin Dye Technique Assay



Light Microscopy Observation.

Untreated Acanthamoeba spp.



GR-treated *Acanthamoeba* spp.



MH-treated *Acanthamoeba* spp.



CHX-treated Acanthamoeba spp.



The effects of MH, GR and CHX on Acanthamoeba spp. size

Significant reduction of cell size compared to untreated *Acanthamoeba* (P<0.001)

Implies a significant loss of cytoplasmic constituents

Dense cytoplasms observeddehydration

	Treatment	Size		
<i>Acanthamoeba</i> isolates		Length ± S.D (µm) (n=30)	Width ± S.D (µm) (n=30)	
Acanthamoeba	Control	23.55 ± 4.36	13.75 ± 2.50	
castellanii	MH	17.07 ± 3.69*	13.81 ± 3.42	
(IMR isolate)	GR	14.39 ± 3.61*	13.20 ± 3.42	
	CHX	15.96 ± 2.36*	13.00 ± 3.13	
Acanthamoeba	Control	25.62 ± 5.80	16.23 ± 4.26	
polyphaga	MH	20.16 ± 3.17*	12.36 ± 3.48*	
(CCAP 1501/3A)	GR	23.25 ± 5.84	16.23 ± 5.12	
	CHX	19.82 ± 3.60*	12.65 ± 6.52*	
Acanthamoeba sp.	Control	23.82 ± 5.24	16.98 ± 4.67	
(HKL isolate)	MH	17.92 ± 3.23*	13.29 ± 2.84*	
	GR	18.32 ± 3.20*	14.19 ± 3.15*	
	CHX	19.54 ± 3.86*	14.56 ± 3.88*	
Acanthamoeba sp.	Control	23.21 ± 3.09	15.78 ± 3.87	
(SW isolate)	MH	21.51 ± 3.75	19.32 ± 3.84	
	GR	17.78 ± 2.47*	15.02 ± 2.16	
	CHX	16.95 ± 3.11*	14.37 ± 3.91	

SEM Observation



Untreated *Acanthamoeba* spp.

 Irregular Acanthamoeba shape

Numerous acanthapodia (spine-like structure) on cell surface











Loss of spine-like appearance (acanthapodia)

Uneven cell surface

Cystic shape of *Acanthamoeba* cell

Sunken food cup

Elongated acanthapodia attached to the substratum

GR-treated Acanthamoeba spp.









- Loss of spine-like appearance (acanthapodia)
- Uneven cell surface
- Cystic shape of

Acanthamoeba cell

- Sunken food cup
- Elongated acanthapodia attached to the substratum

CHX-treated Acanthamoeba spp.



- Typical acanthapodia still
 occurred after treatment
 with CHX on the cell
 surface
- Wrinkle cell surface
- Cystic shape of *Acanthamoeba* cell
- Sunken food cup
- Loss of acanthapodia
- Abnormal appearance of CHX-treated *Acanthapodia* (compressed).

The amoebae were undergo encystment process

Loss of acanthapodia and pseudopodia structure, rounded shape of amoebae

"...unable to bind to the host cells which leads to phagocytosis (engulfing host cells).." (Khan, 2003)

Cyst are survival forms of Acanthamoeba under harsh conditions.

Cystic shape cells- reduce surface area for absorption "....cyst are non-infective stage of Acanthamoeba.." (Dudley et al., 2005, Garate et al., 2006)

TEM Observation

Rounded shape of lipid droplets



Distin mitoch cris



Distinctive mitochondria cristae

Untreated A. polyphaga (CCAP 1501/3A)







MH-treated Acanthamoeba sp. (SW isolate)









- Fragmented and condensed chromatin after treatment with MH and GR – similar type of chromatin condensation in apoptotic intestinal parasite, *Blastocyst hominis* (Tan and Nasirudin., 2005)
- Chromatin condensation pattern different from mammalian cells activation of different protease
- Pyknotic nucleus volume of nucleus of a damaged cell became decreased & darker due to the condensation of nuclear chromatin.
- Alteration to the mitochondria cristae pathway of mitochondria damage, attempt to maintain the energy generating in mitochondria of amoeba cell
- Mitochondria adjacent to the lipid droplets (LD)- reported in higher eukaryotic cells, suggested lipid metabolism by mitochondria after treatment with MH, GR and CHX
- Ruptured LD metabolized in the encystment process
- Autophagy damaged organelle surrounded by endoplasmic reticulum

CONCLUSION...

MH and GR having potent anti amoebic activity against all species of Acanthamoeba used in this study.



".....By Light Microscopy and SEM, the alterations and changes observed were mainly on the surface of cell membrane and acanthapodia structure..."

".....TEM revealed the changes occurred at Acanthamoeba ultrastructural level. The noticeable changes were on the nuclear structure, mitochondria, lipid droplets and endoplasmic reticulum complex....."

MODE OF CELL DEATH DETERMINATION IN *ACANTHAMOEBA* SPP. TREATED WITH MAHANIMBINE AND GIRINIMBINE

- Cell viability determination in amoeba cell by AO/PI staining method.
- **Fragmentation of DNA (apoptosis) in amoeba cell by DNA laddering assay.**
- Mode of cell death determination (by apoptosis)-

>potent mechanism to remove parasitized cell from host (Bruchhaus et al., 2007).

Results and discussion



Fluorescence Microscopy by AO/PI Staining

Treatment with the compounds and CHX at their IC_{50} value in 6-well plate, 72 hours

Amoeba cells were harvested by centrifugation (3000rpm, 15 mins)

> Pellet obtained resuspended in 100uL AO/PI solution

Washed once with PBS

Incubated for 10 mins

Amoeba cells were placed onto slides

Viewed under fluorescence microscopy, blue filter Recentrifuged

DNA Laddering Assay by Agarose Gel Electrophoresis





Evaluation of Mode of Cell Death in Individual Acanthamoeba Cell Using AO/PI Staining Acanthan and spr





A. polyphaga

Acanthamoeba sp. (SW isolate)



Green fluoresce Acanthamoeba cytoplasma, nucleus

viable Acanthamoeba cells intact membrane intact nucleus







Denaturation and condensation of nuclear structure

Evidences: Yellow-orange precipitates -ssb of DNA

Green precipitates- dsb of DNA (based on Darzynkewicz et al., 1983)

Yellow-orange granules -active lysosomes during autophagy (based on Darzynkiewicz et al.,)



GR-treated Acanthamoeba spp.

Denaturation and condensation of nucleus structure

Yellow-orange precipitates -ssb of DNA

Green precipitates -dsb of DNA

Yellow-orange granules -active lysosomes during autophagy

Yellow-orange amoeba cells
 membrane leakage



Yellow-orange amoeba cells -membrane leakage

Acanthaman Acantha

Yellow-orange precipitates -ssb of DNA (apoptosis)

Green precipitates -dsb of DNA (apoptosis)

Yellow-orange granules
 -active lysosomes during autophagy

Green and red precipitates:

Single and Double strand of Fragmented DNA

Green precipitates : Double strand break of DNA



AO: Binds to dA & dG moieties of DNA
 Intercalate between the DNA base pairing
 No H⁺ (intact hydrogen bond), no acidification and AO fluoresce green

Green and red precipitates:

Single and Double strand of Fragmented DNA

Red DNA precipitates: Single strand break of DNA



 Proton (H⁺) due to the breakage of the H bond between the DNA strand
 Acidification to the DNA moieties
 AO fluoresce red

Yellow-orange vesicular structure: Protonated Lysosomes





Inside lysosome : pH ~ 4.5-5.0
 Cytosol : pH ~ 7.0-7.4
 AO:

- Accumulate in lysosome
- Quenching of its fluorescence correlated with Δ pH across the lysosomal membrane
- Sensitive to the acidification (H⁺)
- Support the existence of lysosomal H⁺ pump in amoeba lysosomes

Yellow-orange Amoeba Cells: Disrupted Amoeba Cell Membrane





Propidium Iodide

- charged molecule, excluded from a cell's lipid membrane
- passes freely through the permeabilized membrane of dead and dying cells (Stork and Li., 2006)
- reaction with Zn²⁺ (high in dying or dead cells) (Frederickson et al., 2005)

Analysis of DNA Fragmentation in *Acanthamoeba* spp.





DNA fragmentation in

A. polyphaga (CCAP 1501/3A)

DNA fragmentation in treated *Acanthamoeba* (apoptosis)

Smeary pattern and fragmentation of DNA in – combination of apoptosis and necrosis *Acanthamoeba* cells after treatment

Intact DNA of untreated Acanthamoeba

	-ve GR MH CHX	
		 1500bp
*		 1000bp
>		 500bp
>		

DNA fragmentation in

Acanthamoeba sp. (HKL isolate)

^{500bp} Acanthamoeba (apoptosis)

Smeary pattern and fragmentation of DNA in – combination of apoptosis and necrosis *Acanthamoeba* cells after treatment

Intact DNA of untreated *Acanthamoeba*



DNA fragmentation in Acanthamoeba sp. (SW isolate)

DNA fragmentation in treated *Acanthamoeba* (apoptosis)

Smeary pattern and fragmentation of DNA in – combination of apoptosis and necrosis *Acanthamoeba* cells after treatment

□Intact DNA of untreated Acanthamoeba



Endogenous Nuclease Enzyme

Confirmation of apoptosis and necrosis by Annexin V-FITC after 24 hours treatment confirm the absent of apoptosis event at early stage of Acanthamoeba cell death



The apoptosis was measured by Annexin V-FITC and flow cytometry analysis (a) untreated cells as negative control with 0.06% of apoptosis (b) chloramphenicol-treated cells 0.89% of apoptosis (c) EO_3 . Pr.Pic treated cells with 0.10% of apoptosis (d) EO_3 (Nd)H₂O.Pic treated cells with 0.14% of apoptosis (e) and EO_3 . Gd.Pic treated cells with 0.08% of apoptosis.

APOPTOSIS

Chloramphenicol-treated cells EO₃.Pr.Pic treated cells EO₃(Nd)H₂O.Pic treated cells EO₃.Gd.Pic treated cells

- 0.89% of apoptosis
- 0.10% of apoptosis
- 0.14% of apoptosis
- 0.08% of apoptosis.

Apoptosis

- Indicated externalization of PS.
- Annexin V-FITC bind to PS.

Autophagy – lanthanide complexes treated cells

- Intercellular cell death
- No externalization of PS occurred.
- Annexin V-FITC unable to detect due to no exposure of PS.

Necrosis and programmed cell death occurred in Acanthamoeba cells after treatment with MH and GR Fragmentation of DNA, typical apoptosis criteria were also observed in amoeba cells after treatment with both compounds

Necrosis and programmed cell death (autophagy and apoptosis) were recognized and quantified in a population of amoebae when double fluorochromes staining AO/PI, an express, fast, easy, sensitive and reproducible method were utilized.

DNA fragmentation after treatment with MH and GR, a characteristics feature of apoptosis in *Acanthamoeba* cells were confirmed by DNA laddering assay in agarose gel.

- Evidences MH and GR exhibited similar ladder pattern (250 internucleosomal cleavage) in all treated *Acanthamoeabae*
 - Staining by AO/PI also indicated that autophagic type of cell death also occurred in Acanthamoeba cells after treatment with both compounds.

- MH and GR exhibited very low IC₅₀ values on isolates of Acanthamoeba used in this study
- Significant changes on treated Acanthamoeba observed by LM, SEM, TEM.
- □ Modification in SEM technique.
- Type of Acanthamoeba cell after exposed to the compounds were confirmed by combination of AO and PI (apoptosis, autophagic cell death and necrosis)
- Activation of lysosomal activity in treated Acanthamoeba observed by AO/Pl staining
- DNA laddering confirmed the apoptosis process in treated Acanthamoeba cells
- Genus Acanthamoeba, model for mitochondriate eukaryotic cells
- No similar death cascade has been reported in Acanthamoeba to mammalian cells. Metacaspase in Acanthamoeba (Trzyna et al., 2008) - Arg/Lys specific protein



