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OMICS Group International is an amalgamation of Open Access publications and worldwide international science conferences and events. Established in the year 2007 with the sole aim of making the information on Sciences and technology 'Open Access', OMICS Group publishes 400 online open access scholarly journals in all aspects of Science, Engineering, Management and Technology journals. OMICS Group has been instrumental in taking the knowledge on Science & technology to the doorsteps of ordinary men and women. Research Scholars, Students, Libraries, Educational Institutions, Research centers and the industry are main stakeholders that benefitted greatly from this knowledge dissemination. OMICS Group also organizes 300 International conferences annually across the globe, where knowledge transfer takes place through debates, round table discussions, poster presentations, workshops, symposia and exhibitions.

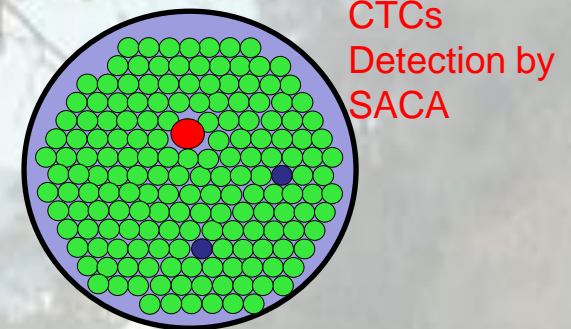
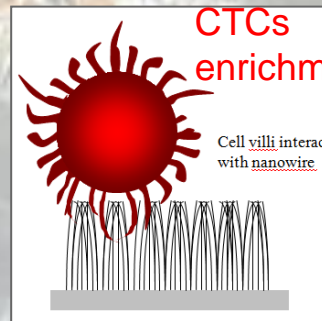
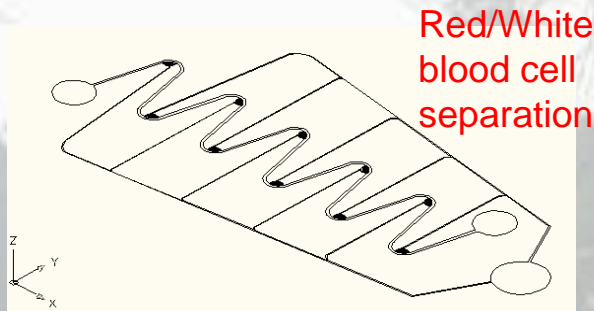
About OMICS Group Conferences

OMICS Group International is a pioneer and leading science event organizer, which publishes around 400 open access journals and conducts over 300 Medical, Clinical, Engineering, Life Sciences, Pharma scientific conferences all over the globe annually with the support of more than 1000 scientific associations and 30,000 editorial board members and 3.5 million followers to its credit.

OMICS Group has organized 500 conferences, workshops and national symposiums across the major cities including San Francisco, Las Vegas, San Antonio, Omaha, Orlando, Raleigh, Santa Clara, Chicago, Philadelphia, Baltimore, United Kingdom, Valencia, Dubai, Beijing, Hyderabad, Bengaluru and Mumbai.



A Nano/Micro Fluidic System for Circulating Tumor Cells (CTCs) Rapid Detection and Diagnosis



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National Health Research Institutes, Zhunan, Taiwan 350**

**^cDivision of Mechanics, Research Center for Applied Sciences,
Academia Sinica, Taipei, Taiwan 115**

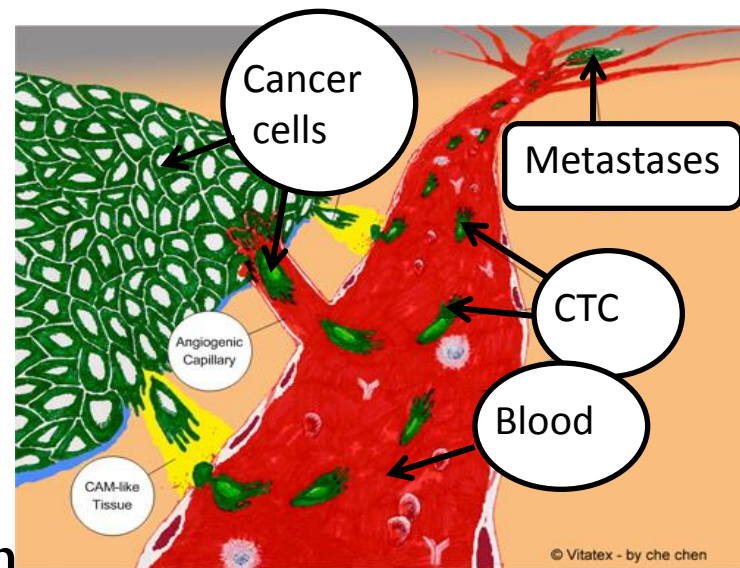
The main technical challenge of CTCs detection

- Main spread way of metastases
 1. nearby tissues spread
 2. blood system spread
 3. lymphatic system spread
- The importance of CTCs amount.
- Challenge:

rare amount of CTC: **1-10 CTCs/1ml**

low separation efficiency: **1 CTCs/10⁶⁻⁷ WBCs**

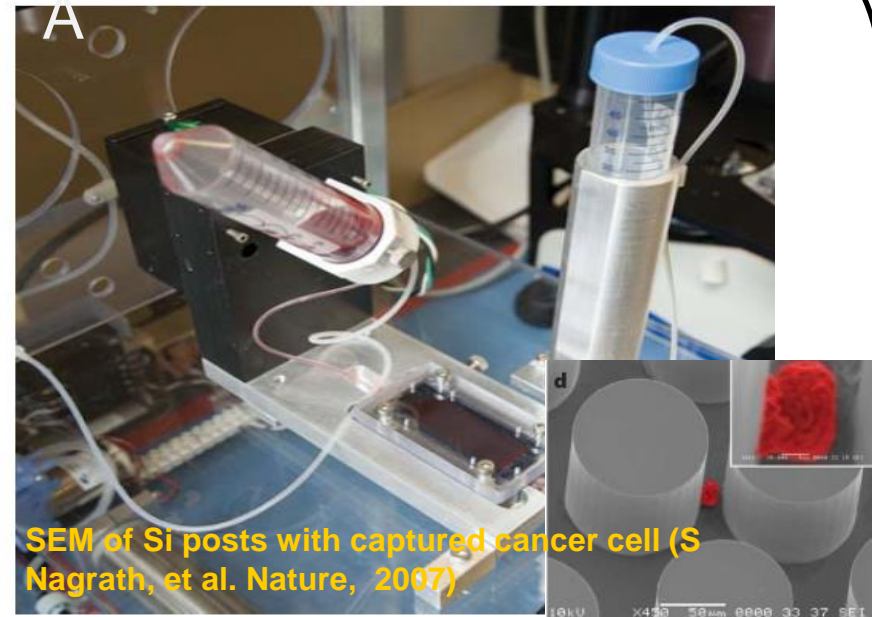
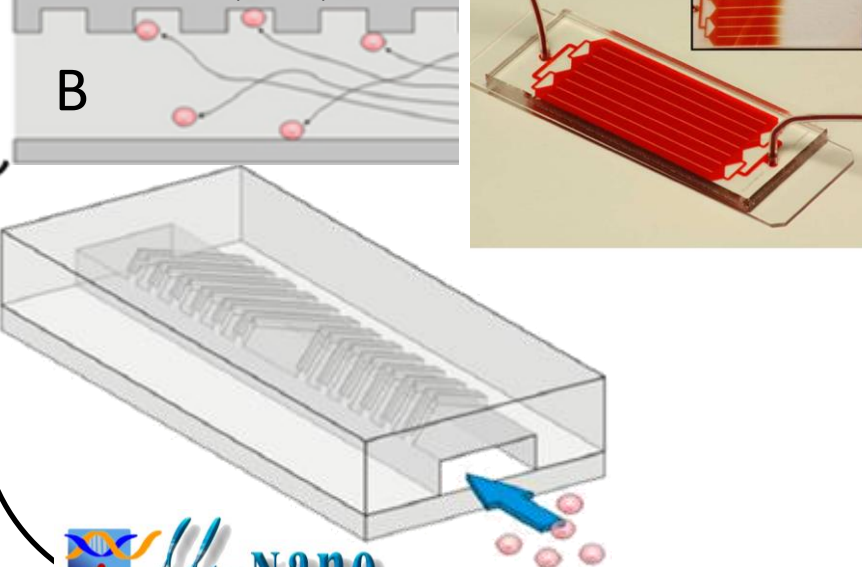
low sensitivity: **40-50%** (specific antibody)



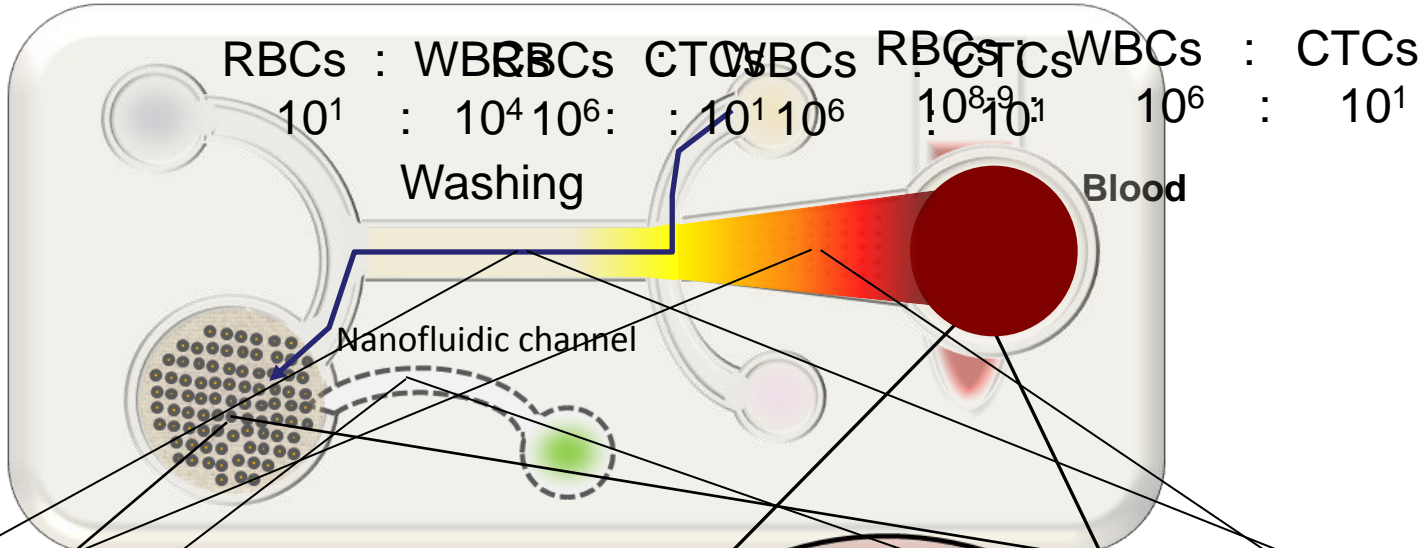
CTCs separation and capturing in whole blood

- Posts fabricated from Si wafer
 - 100 μm diameter
 - 100 μm tall
- Posts coated with anti-EpCAM
- Whole blood flowed through device by pressure source
- mL-scale volumes

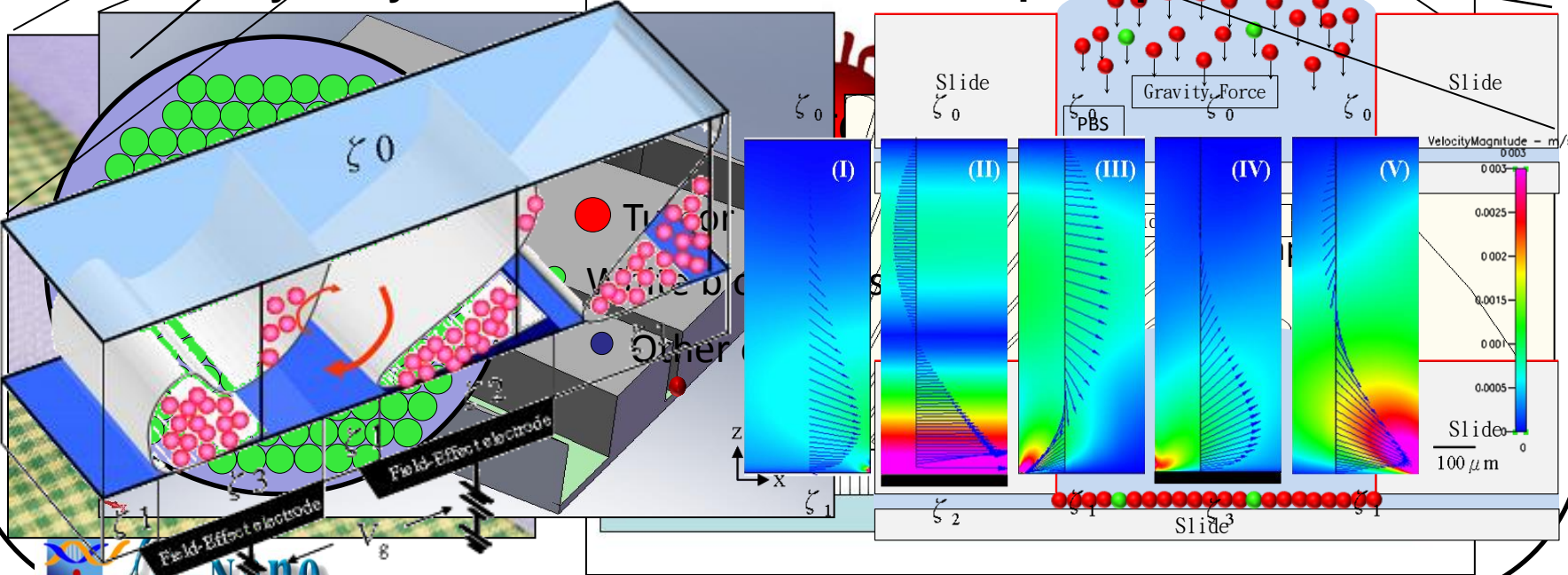
Shannon L. Stotta, PNAS, 2010



- High sensitivity (1 target cell in 1 billion blood cells)
- Selectivity (47%(A), 63%(B) capture efficiency)
- High yield (99(A)% ,95%(B)).
- Max flow of ~ 1 mL/hr to 2.5 mL/hr
- **4-12 hours to run sample**
- Non-specific binding and clogged
- Without sample preconcentration

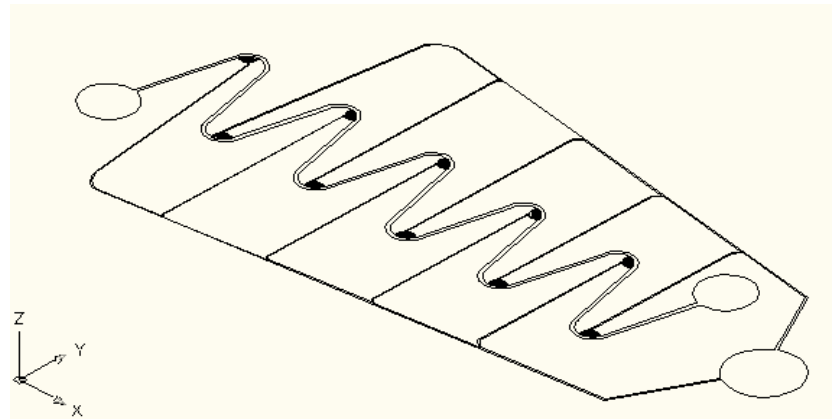


Dual-Channel Dielectrophoretic (DEP) Focusing

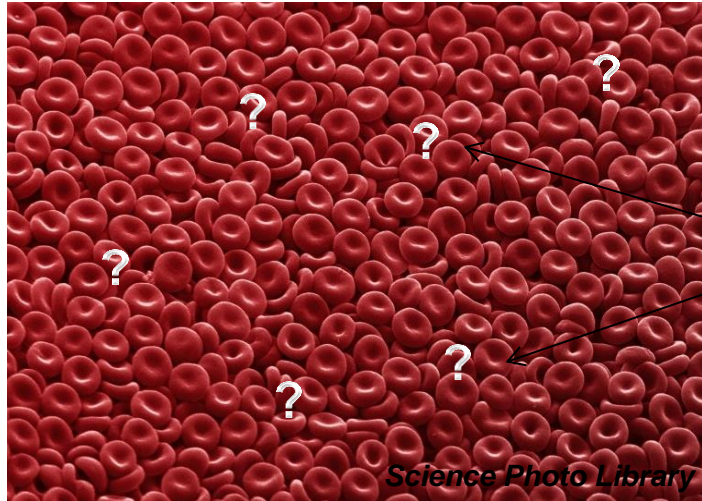




Part I: High-Throughput Blood Cells Separation



The Challenge in early CTCs detection



Red Blood Cell Count (RBCs, 7-9 μm)
 $4.3-6.2 \times 10^9/\text{mL}$ (Male)
 $3.8-5.5 \times 10^9/\text{mL}$ (Female)
 $3.8-5.5 \times 10^9/\text{mL}$ (Infant/Child)

White Blood Cell Count (WBCs, 8-12 μm)
 $4.1-10.9 \times 10^6/\text{mL}$

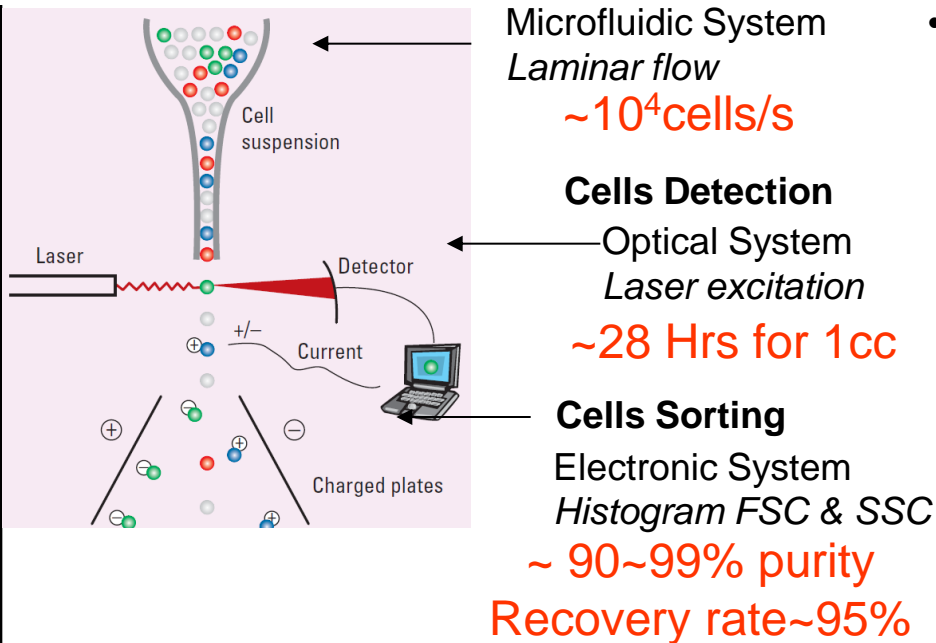
Cancer cell (CTCs) count
Stage O/I: 5-20/mL
Stage II : 20-50/ml
Stage III : 50-100/ml
Stage IV: >100/ml

Limits of current detection:

- Low sensitivity and reliability **>20cells/mL (possibility: 20~60%)**
- Take a long processing time and low recovery rate **>1hr & 80~%**

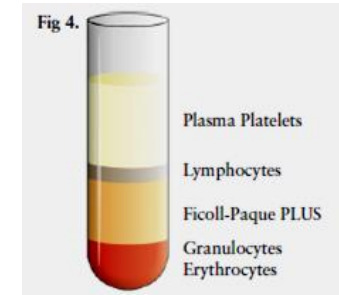
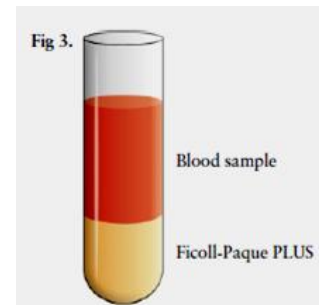
Current technologies for cells separation

1. Flow Cytometry



3. Lysis Buffer (細胞裂解溶劑)

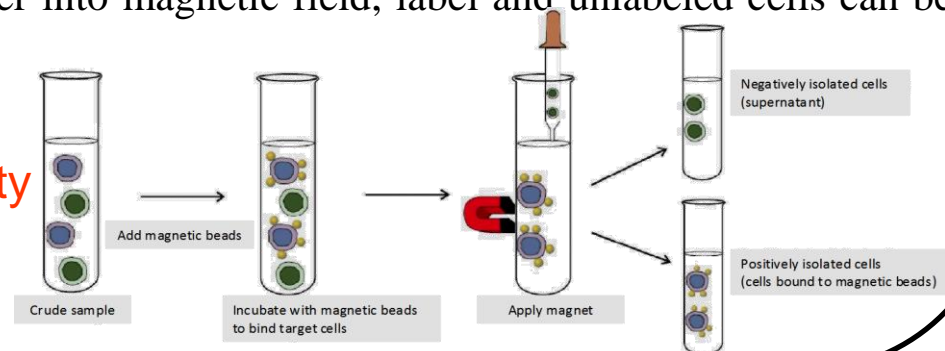
- By using chemical buffer to create osmotic pressure difference between the cell membrane and environmental, specifically use to cleavage and removal of mammalian red blood cells or without nuclei cells in whole blood samples $\sim 10^4/s$, 1hr for 1cc, 90% purity
Recover rate $\sim 40\sim 50\%$



2. Magnetic-Activated Cells Sorting (磁性細胞分選儀)

- By using magnetic-beads combine with high specific monoclonal antibody bind onto the surface antigen of target cells. When the cell enter into magnetic field, label and unlabeled cells can be separated.
- The magnetic-beads diameter is 50nm.

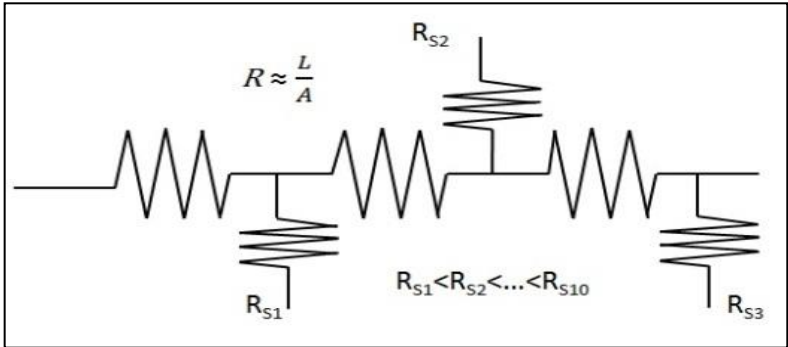
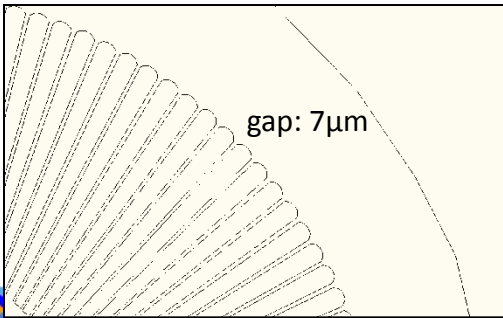
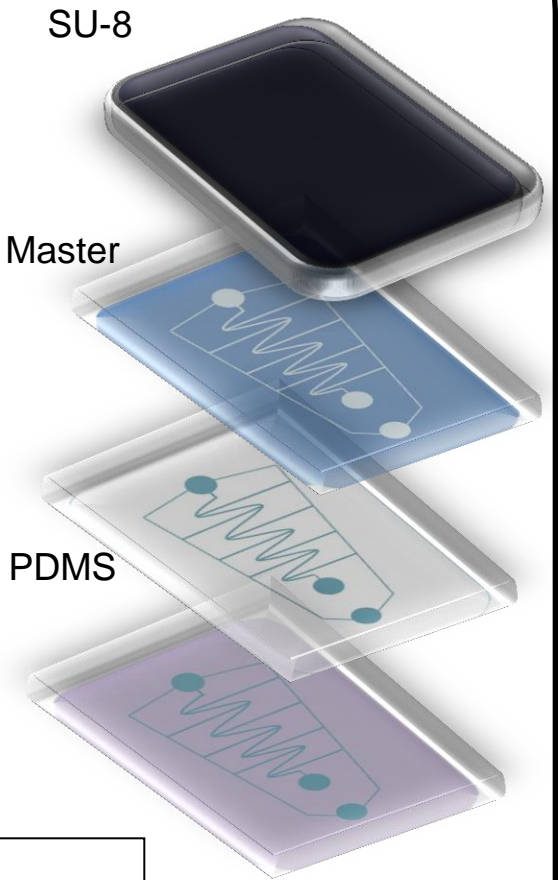
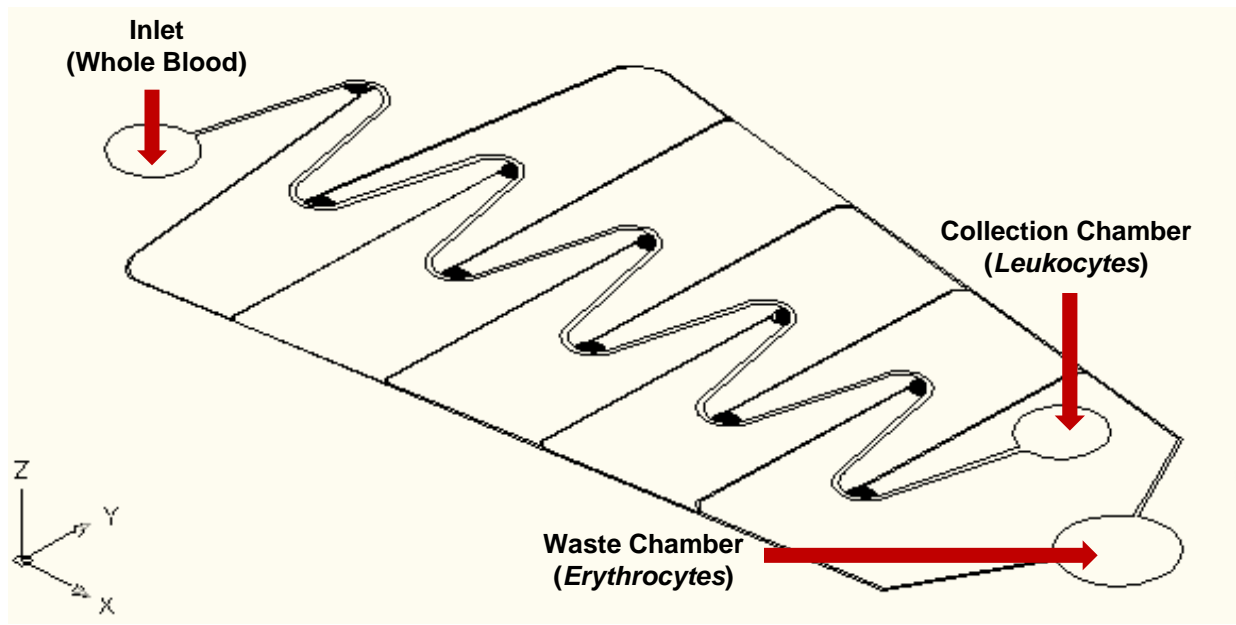
$\sim 10^7$ cells/s, ~ 10 min for 1cc, 99% purity
Recovery rate $\sim 70\sim 90\%$



High Efficient Blood Cells Separation



Chip Size	Sample load	Process time
Main channel (L) 28 cm (WxH) 200 x 580 μm^2	10X dilution 4.3×10^8 cells/mL	(~400 $\mu\text{L}/\text{min}$) 25min.



Theory analysis of Hydrodynamic and Inertial Force

1. Straight Channel

Lift Force (F_L)

$$F_L = \rho G^2 C_L a_p^4$$

Shear Rate (G)

Stokes Drag (F_D)

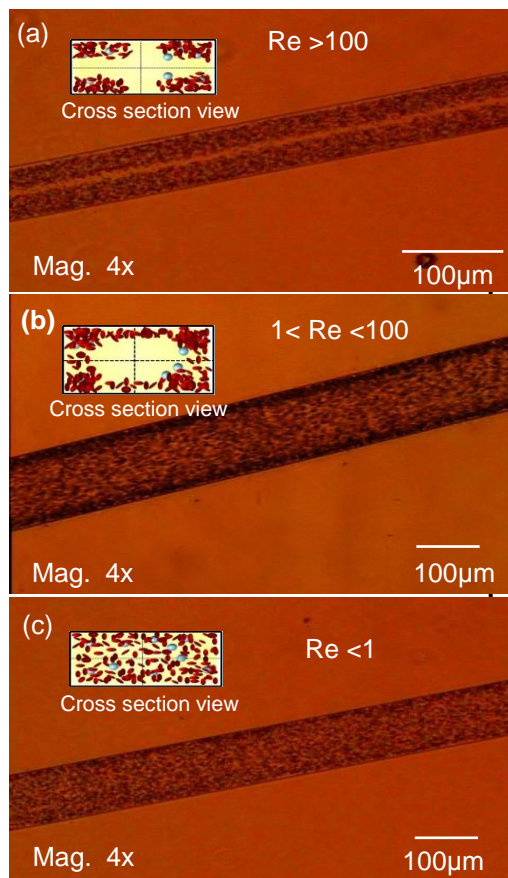
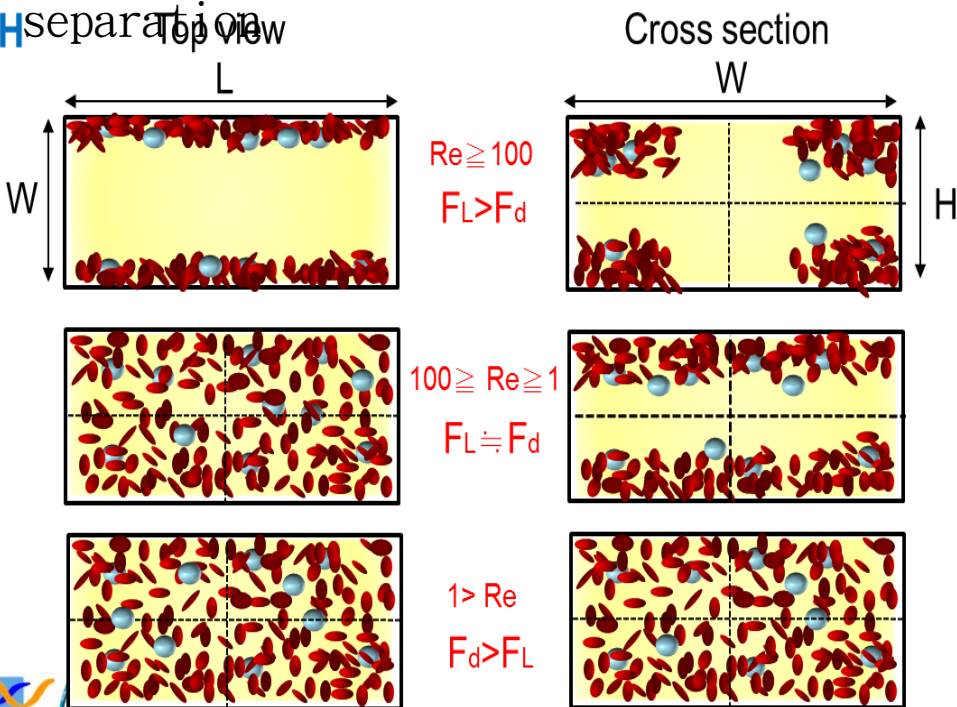
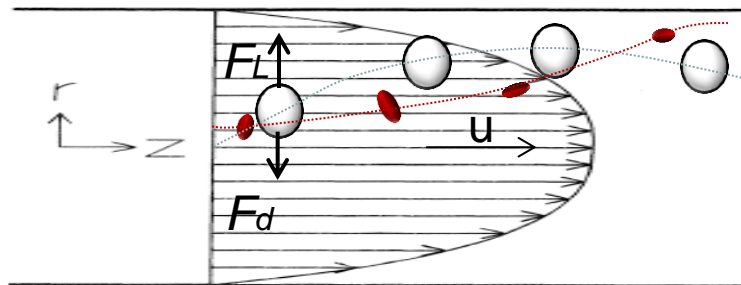
$$F_d = 3\pi\mu a_p U_L$$

Viscosity (μ)

$$L = \frac{2WH}{3(W+H)Re} \left(\frac{W}{a_p} \right)^2$$

L: Length for enough

$W \gg H$ separation

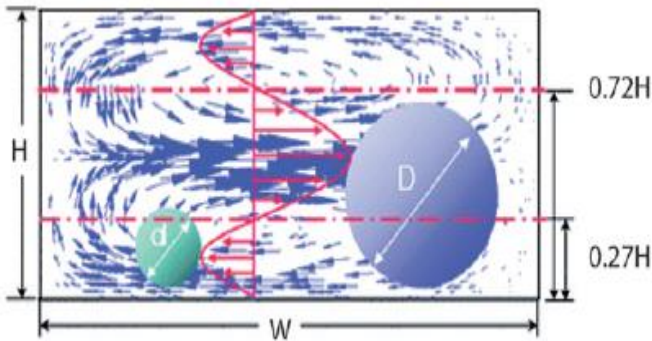


Theory analysis of Hydrodynamic and Inertial Force

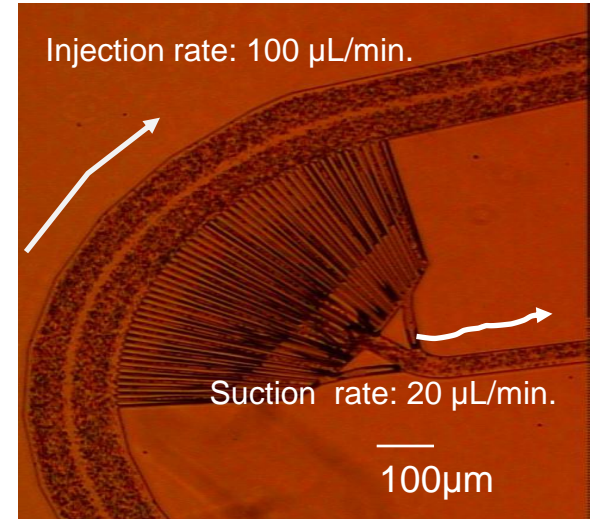
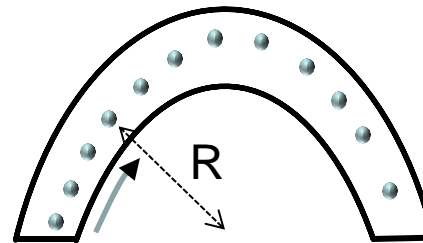
2. Curved Channel

$$\frac{a^2 R}{H^3} > 0.04 \longrightarrow De = \frac{\rho U_f D_h}{\mu} \sqrt{\frac{D_h}{2R}} = Re \sqrt{\frac{D_h}{2R}} > 15$$

Cross section

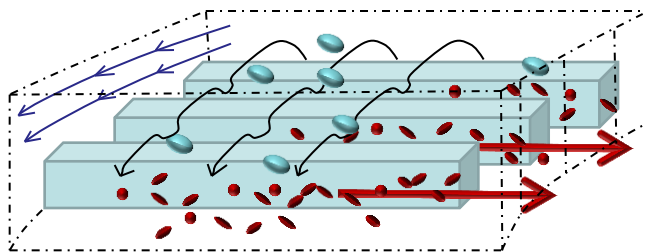


Top view

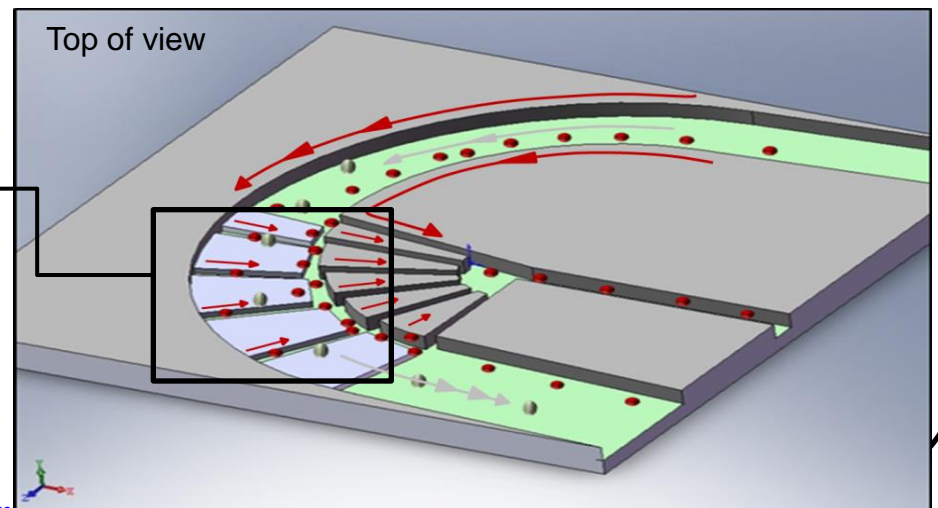


3. Obstacle Channel

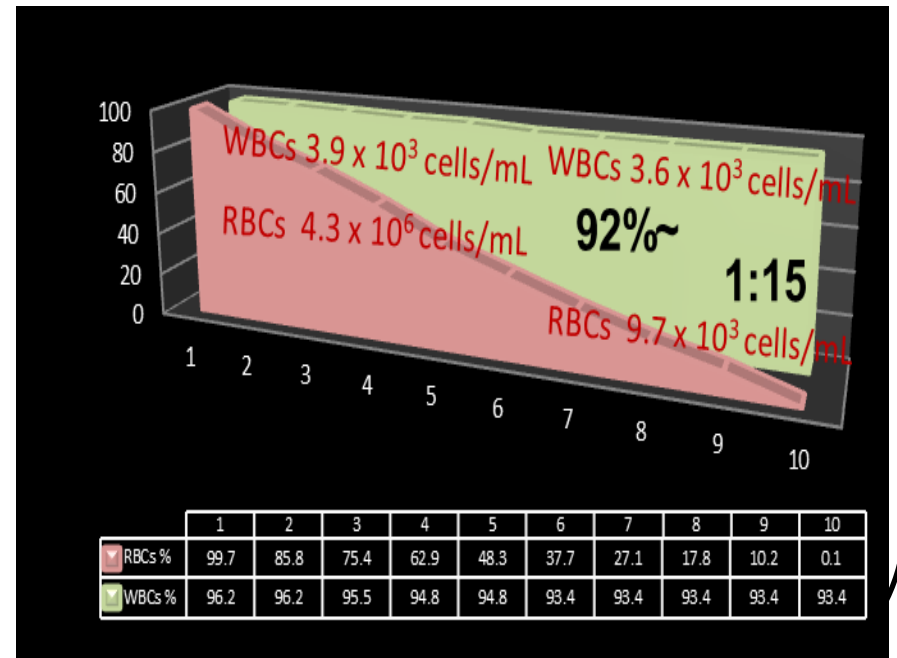
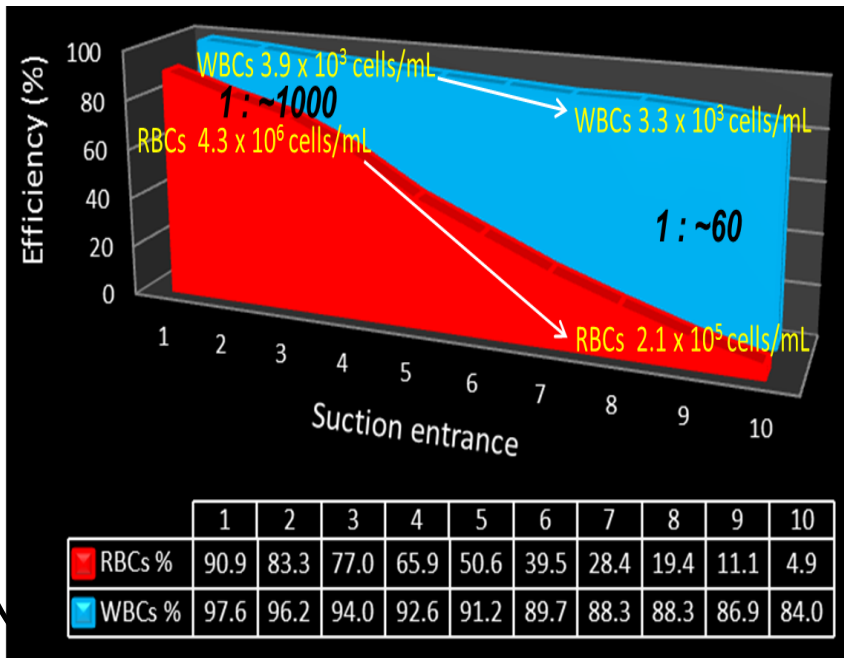
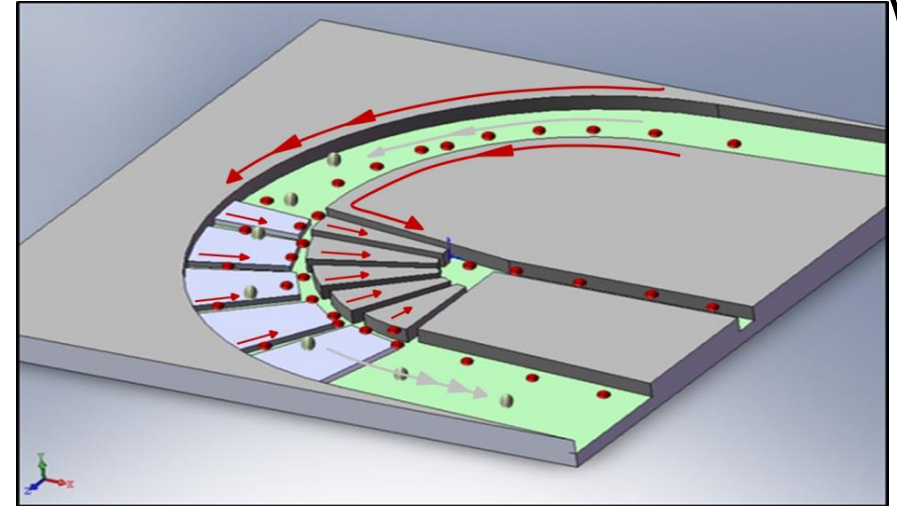
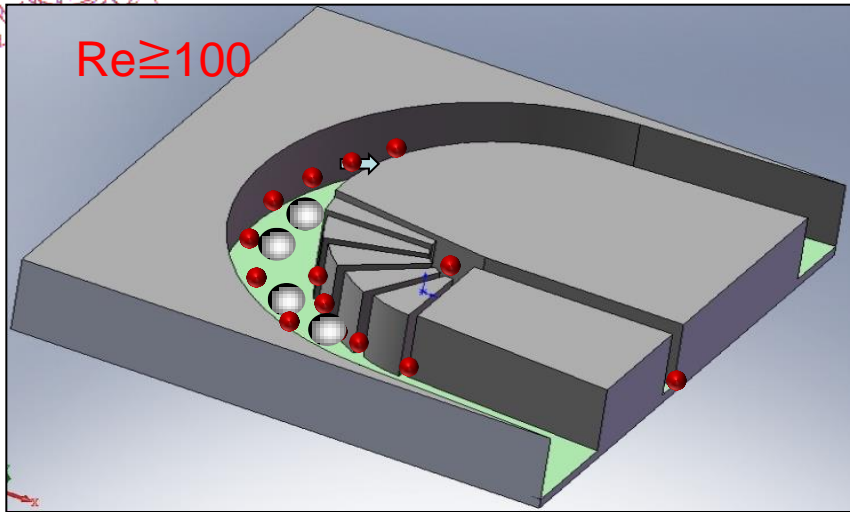
Side of view



Top of view



Results

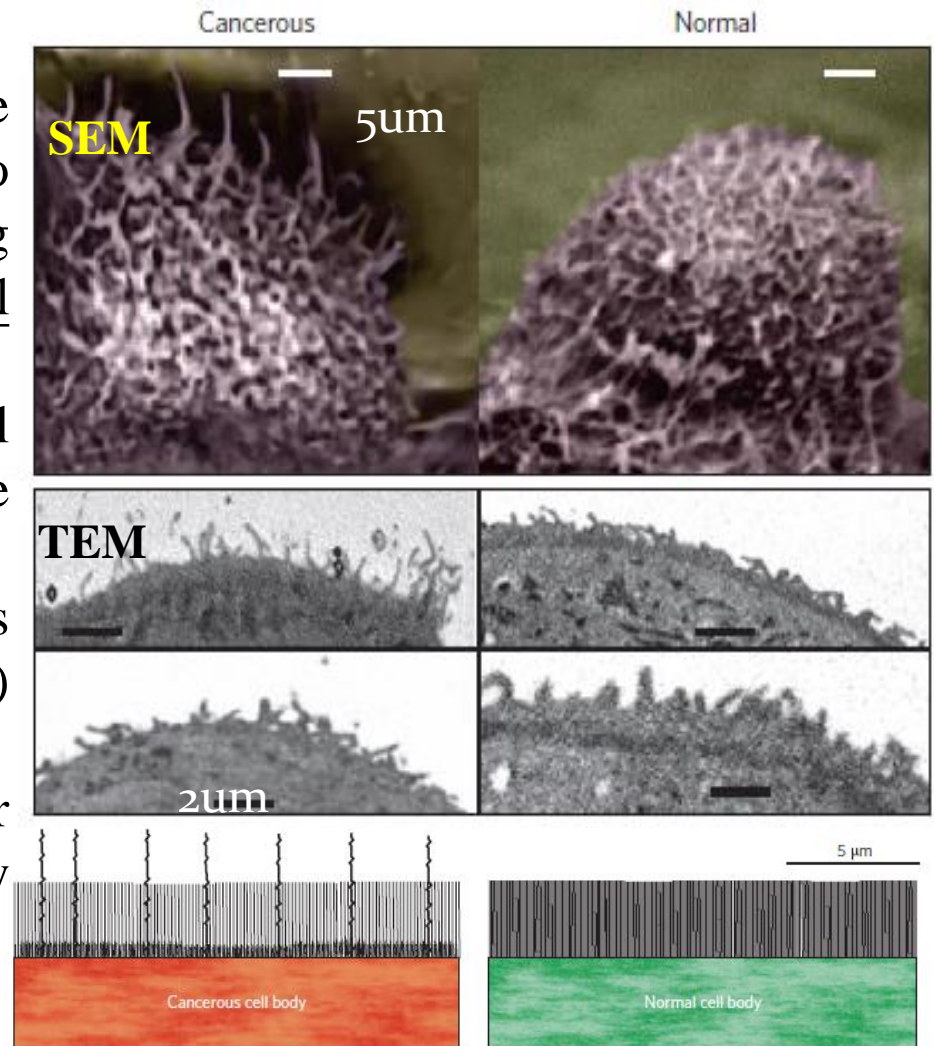




Part II: Silicon nanowire biochip applied on circulating tumor cells detection

Nano structure enhance capture yield

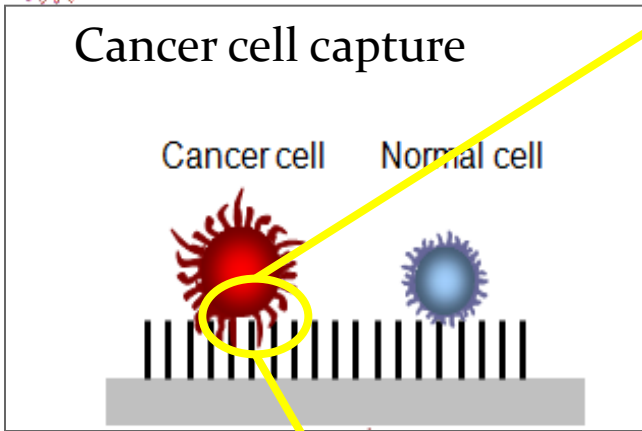
- Microvilli are cellular membrane protrusions, important for of cells to interact with environment, forming brush border, common in epithelial cells
- Microvilli also appears on the cell surface of white blood cells, help the migration of white blood cells.
- The microvilli on the surface appears longer on cancer cell(left, 5&2.6 μm) than normal cell(right, 2.4 μm).
- our research plan to enhance cancer cell capture rate from WBCs by utilizing the length of microvilli.



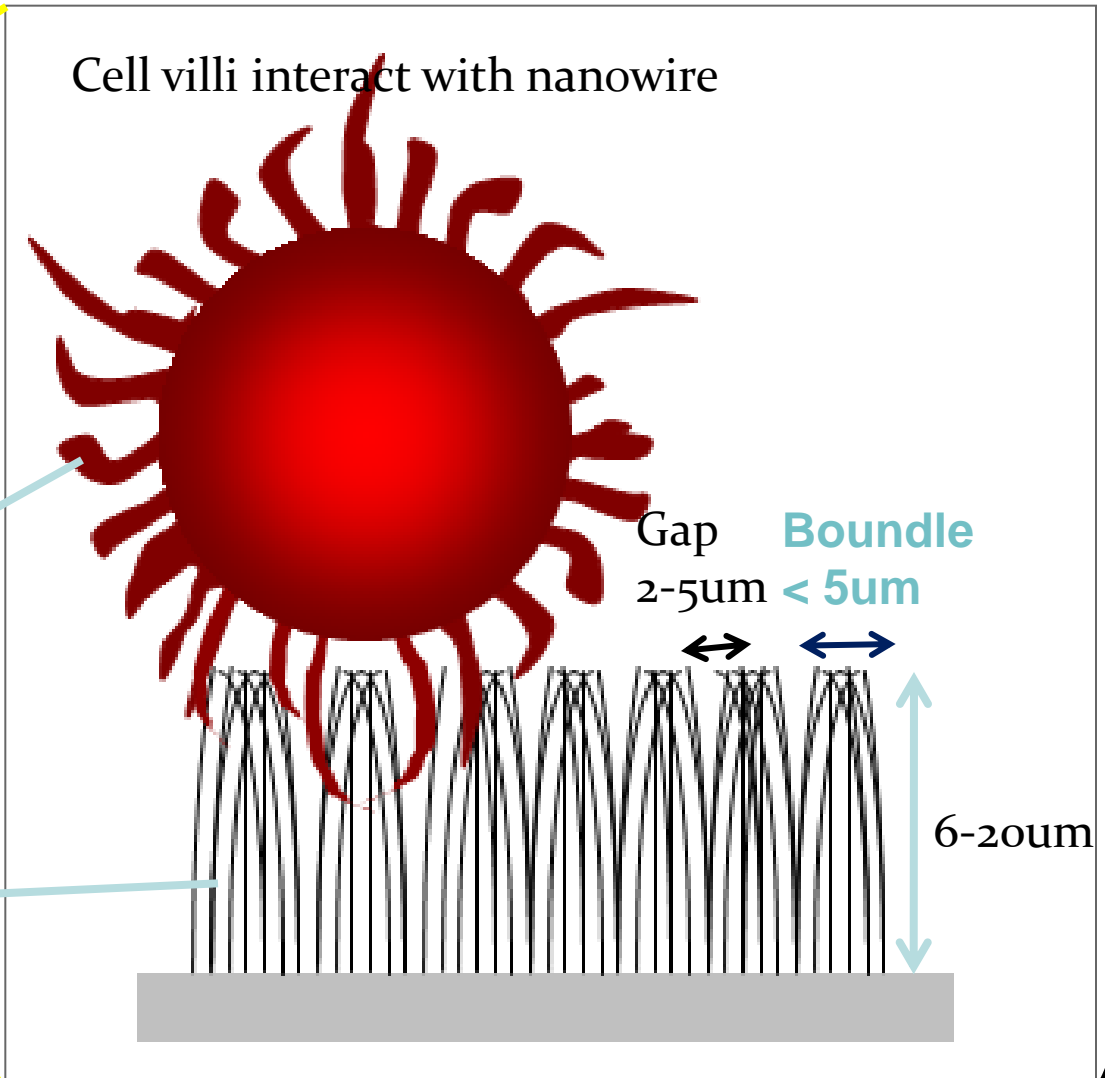
Igor Sokolov et al, 2009,
Nature nanotechnology

Prof. Fan-Gang Tseng

Principle of nanowire-cell villi interaction



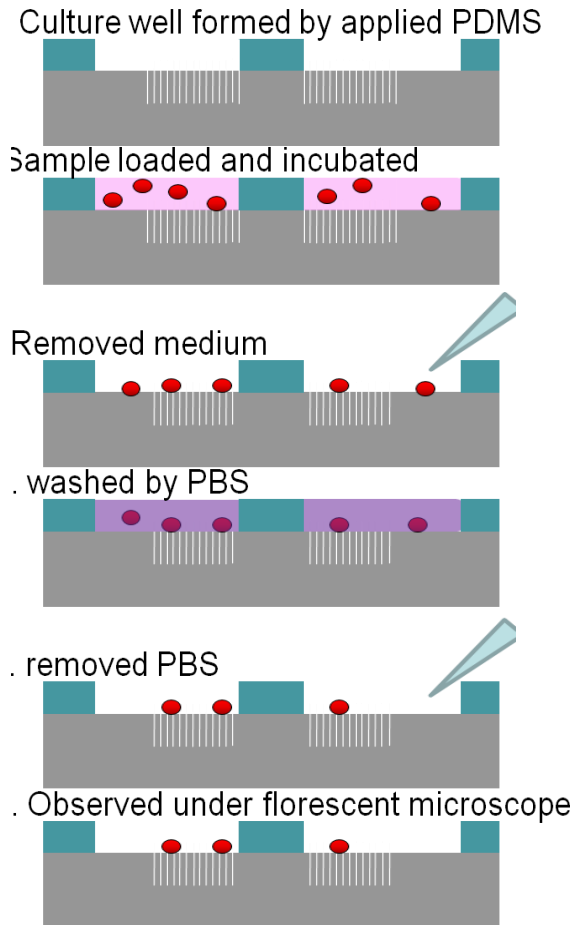
Velcro



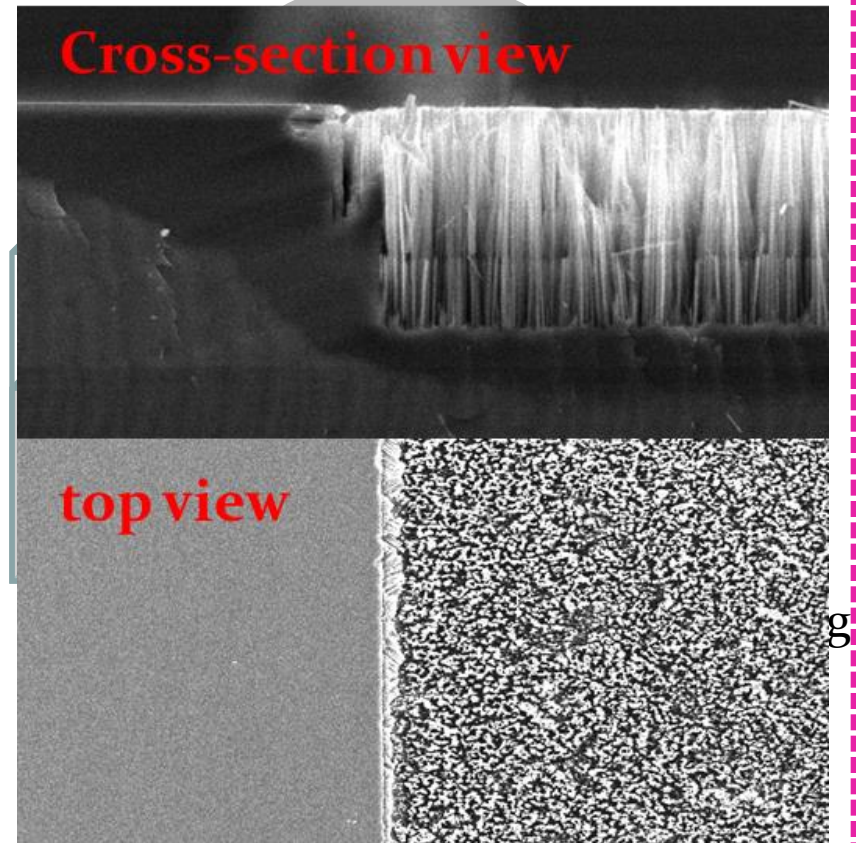
Experimental process and chip fabrication

Conditions:

1. Incubated time
2. Nanowire length

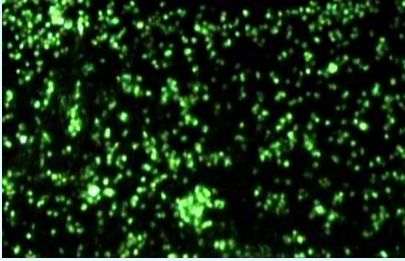
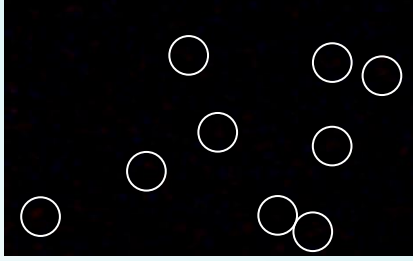
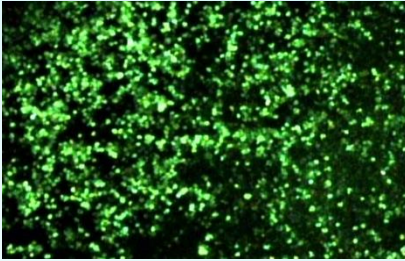
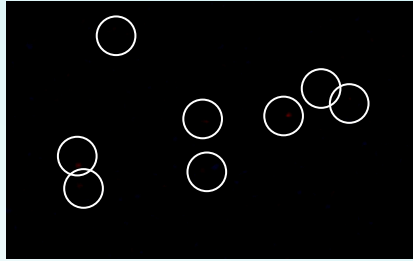

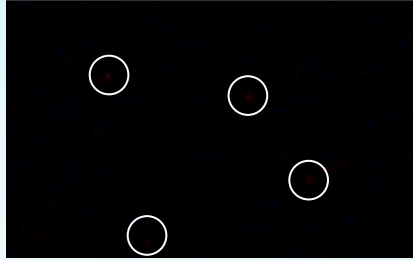


Si nanowire forming



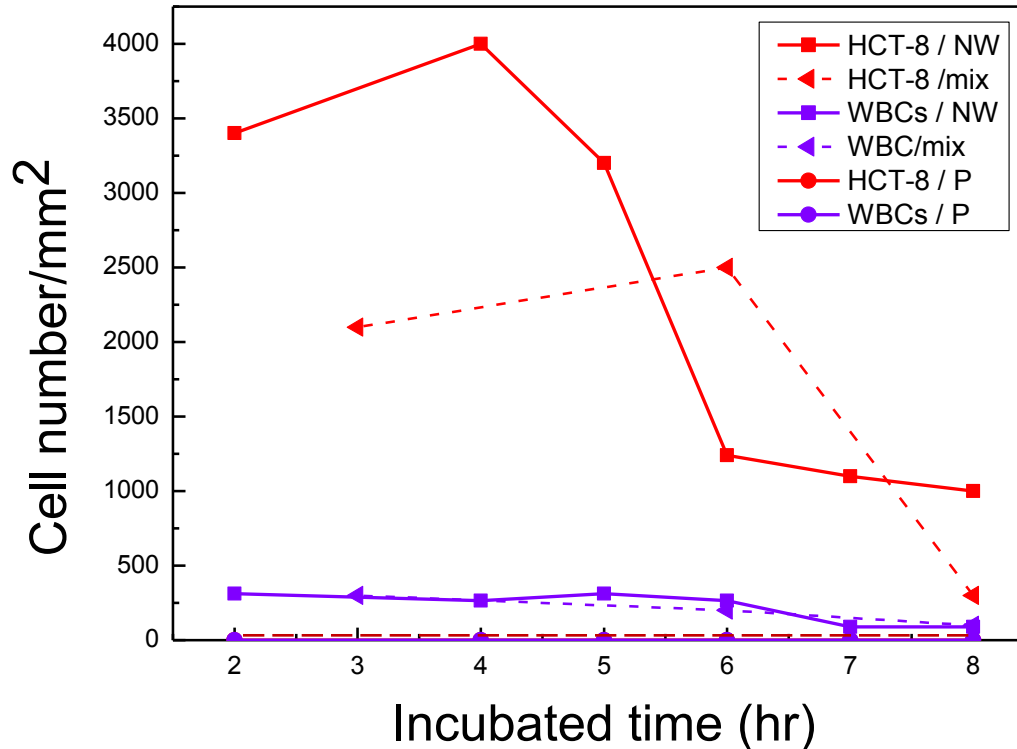
Gap: 1.5~2 μm
aggregation: 1~1.5 μm

Cancer cell (HCT-8) with WBCs(1:1) capture ratio with variable Incubated time

Incubated time	Green emission	Red emission
3hr		
6hr		
8hr		

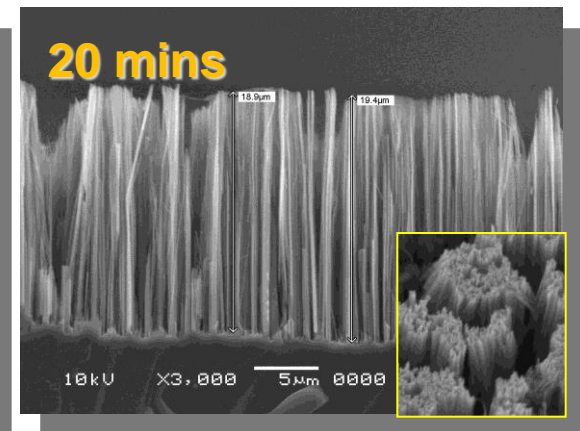
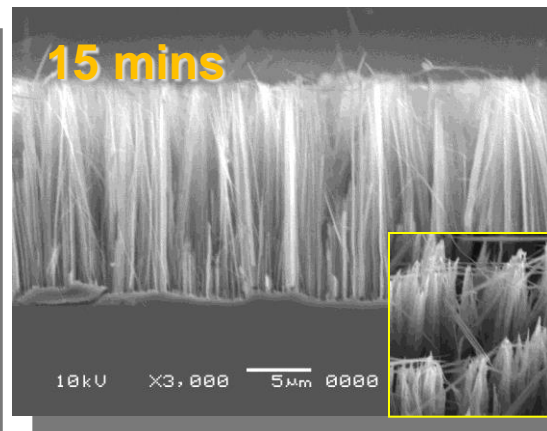
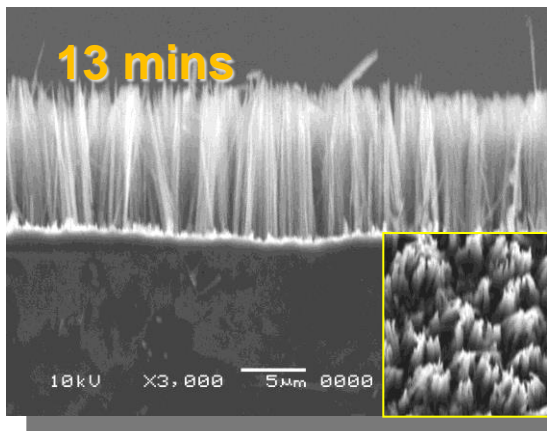
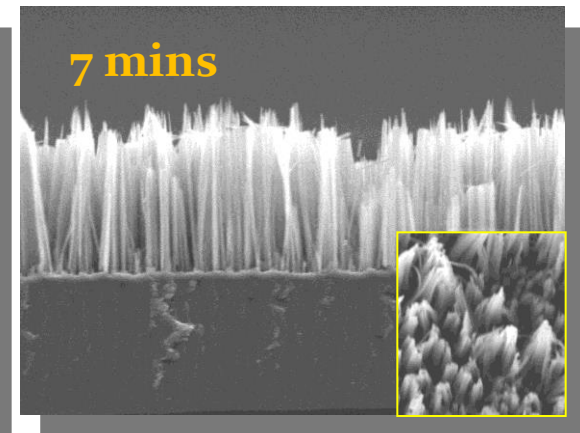
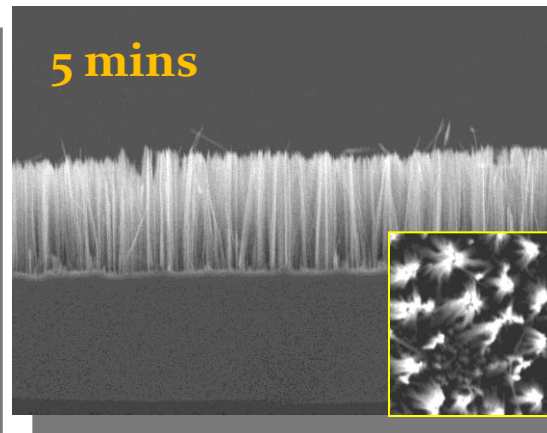
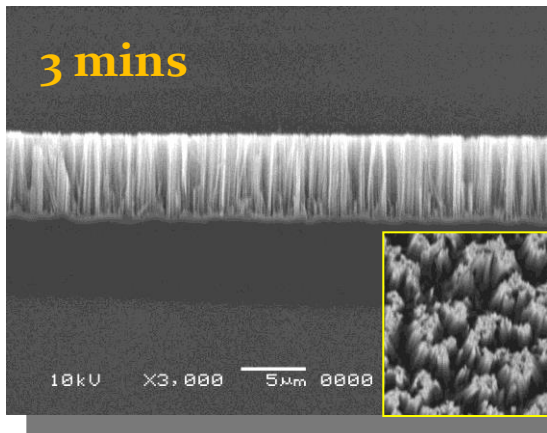
The capture result of different incubated time

Retention rate of two phase chip



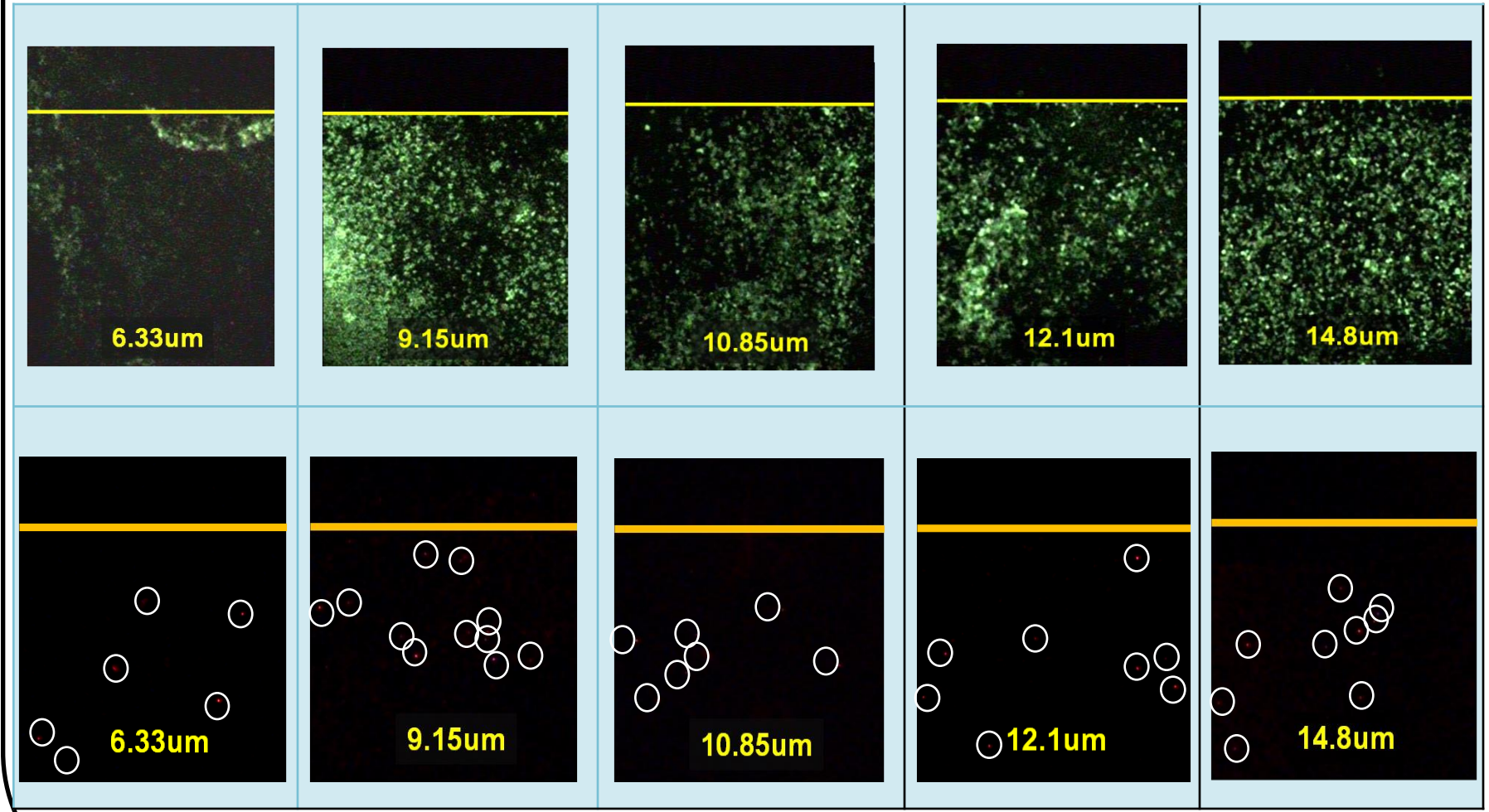
1. Maximum of cancer cell number appears at 4 hours
2. Mixed two cell result in lower capture cell number

Etching time V.S. nanowire length



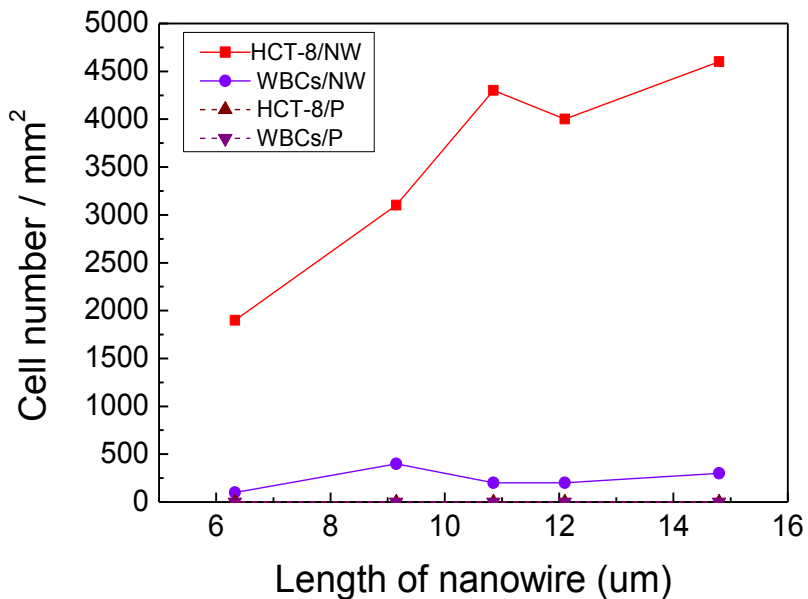


Cancer cell (HCT-8)/WBCs(1:1) capture ratio with variable length



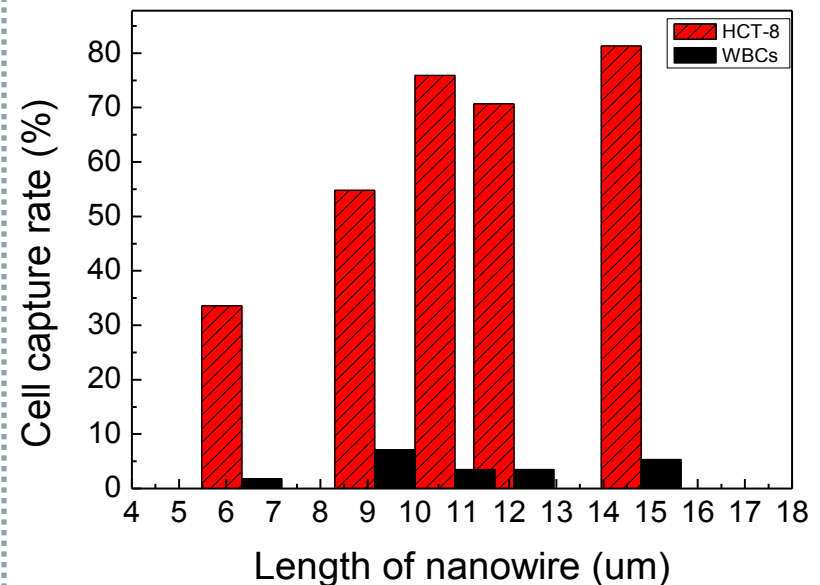
The capture result of different length

Retention rate of two phase chip



When incubated under the same time, the highest retention rate appeared at 14.8 um length nanowire.

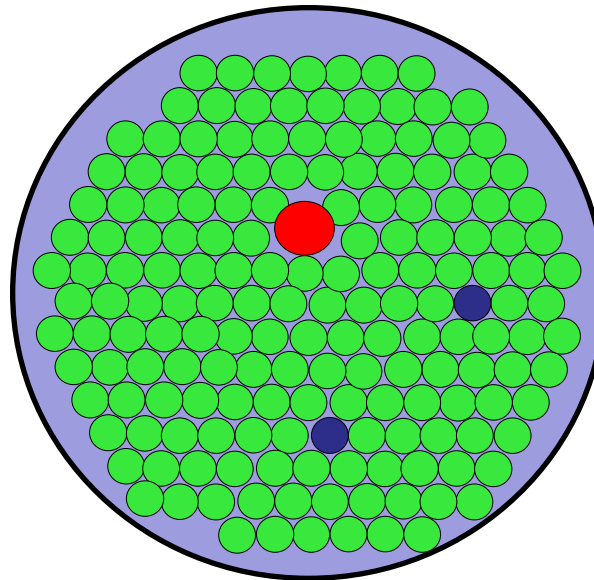
capture rate of two phase chip



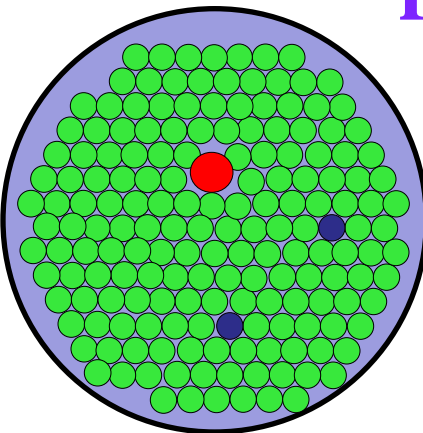
The 14.8 um length nanowire corresponds to 80% capture rate.



Part II: High Density Cell Array Chip for In-Parallel Detection of CTCs



High Density Cell Monolayer Array for in-parallel Detection

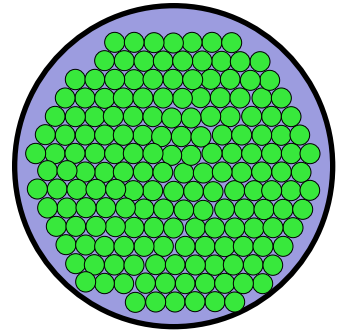


Step 1 ▶

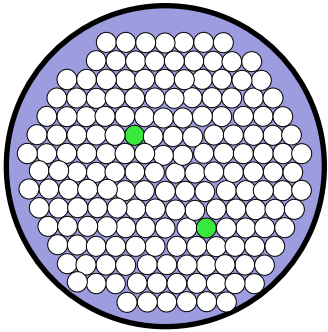
- Tumor cells
- White blood cells
- Other cells

Distinguishing ratio > 100,000:1
 Assembly time < 10 min

High throughput: (10⁶ cells)/cm²)



Step 2 ▶



Step 3 ▶

● Cells stained with dye

○ Cells with GTA fixation

● 647nm : Background cells

● Cells stained with dye

● 488nm : Target cells

• Single type cells

• Two type cells with one stained

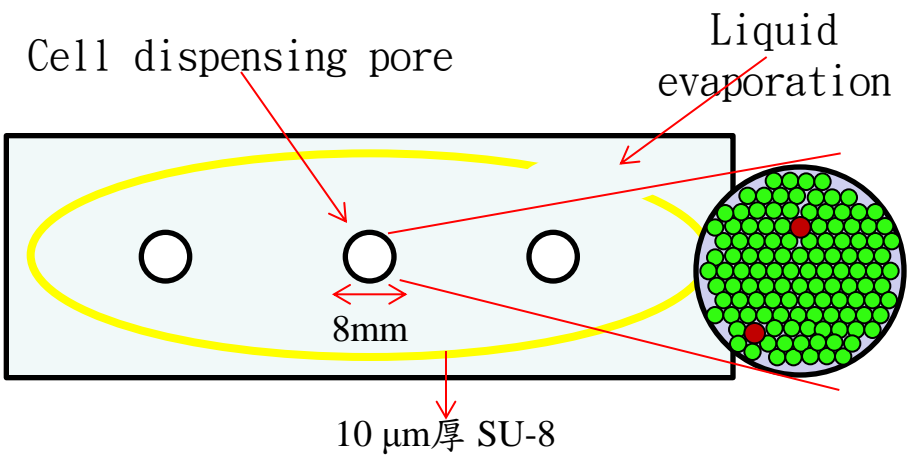
• Two type cells stained differently



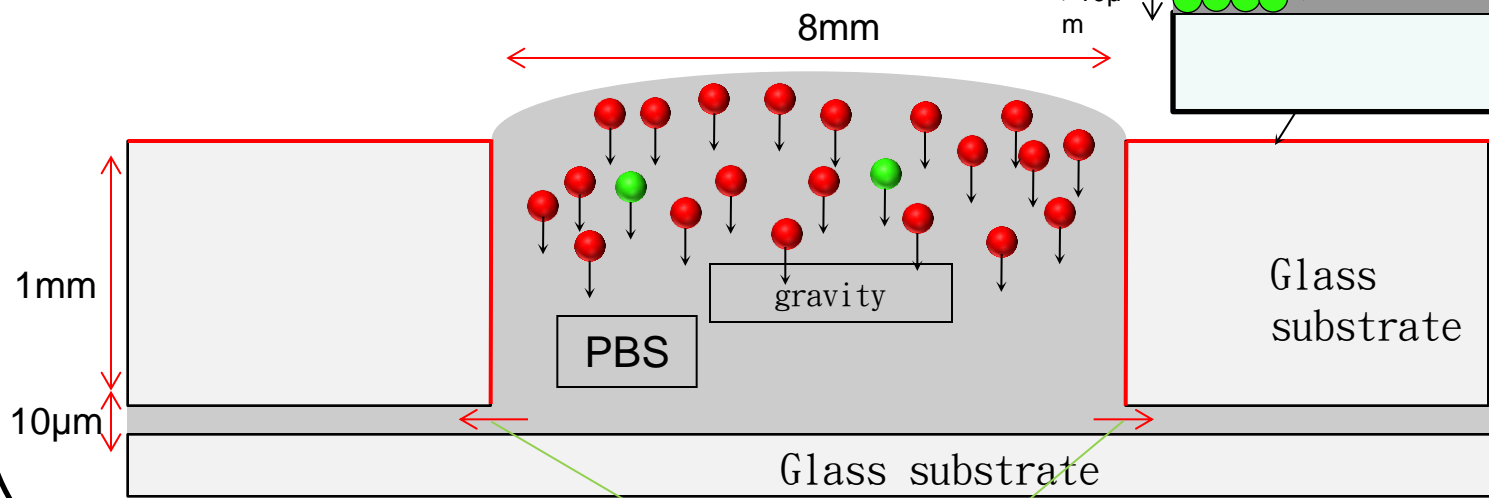
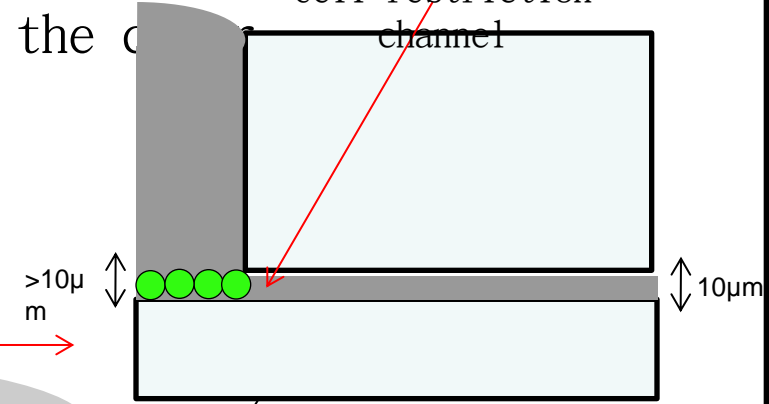
Principle of high density Self-Assembly Cell Array (SACA)



- Major mechanisms:
1. Gravitation force
 2. lateral flow by evaporation

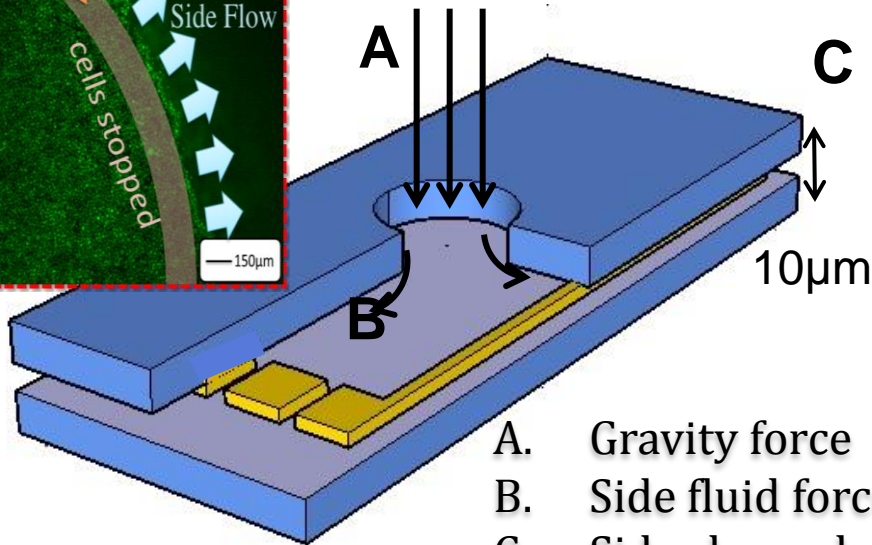
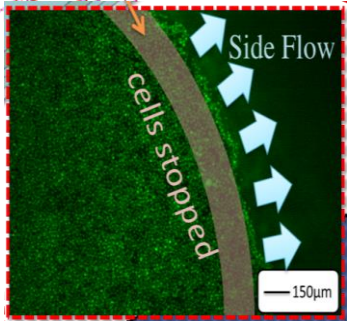


3. Cell restriction into the channel



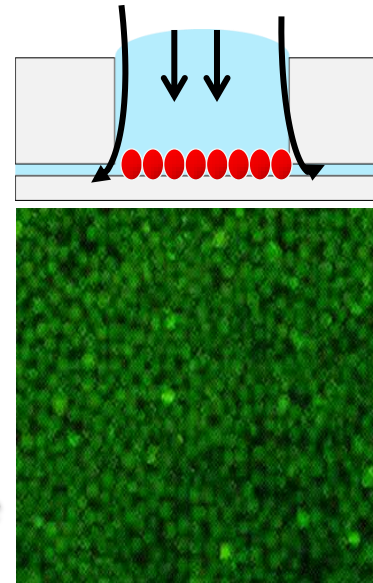
Self-Assembled Cell Array (SACA) Chip

NTHU/ESS/NEMS Institute

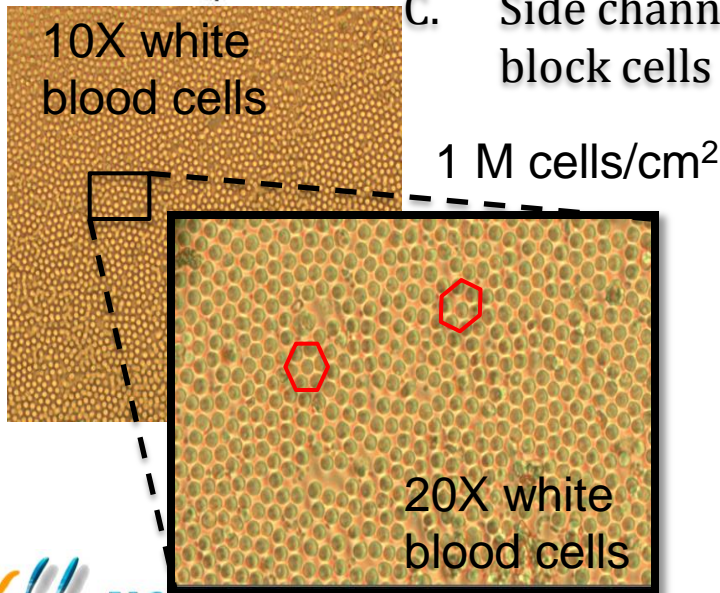
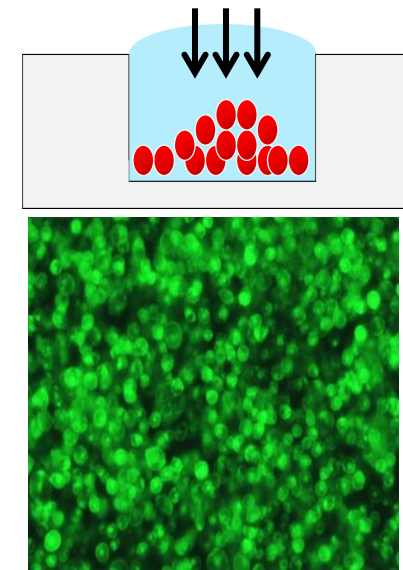


- A. Gravity force
- B. Side fluid force
- C. Side channel block cells

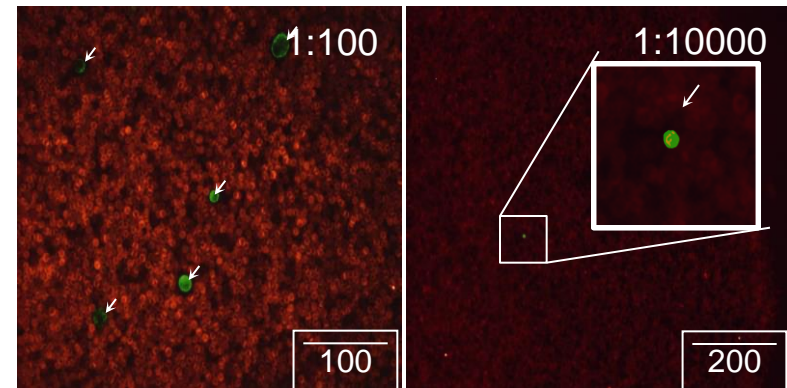
With side channel



NO side channel



Hela cells in white blood cells

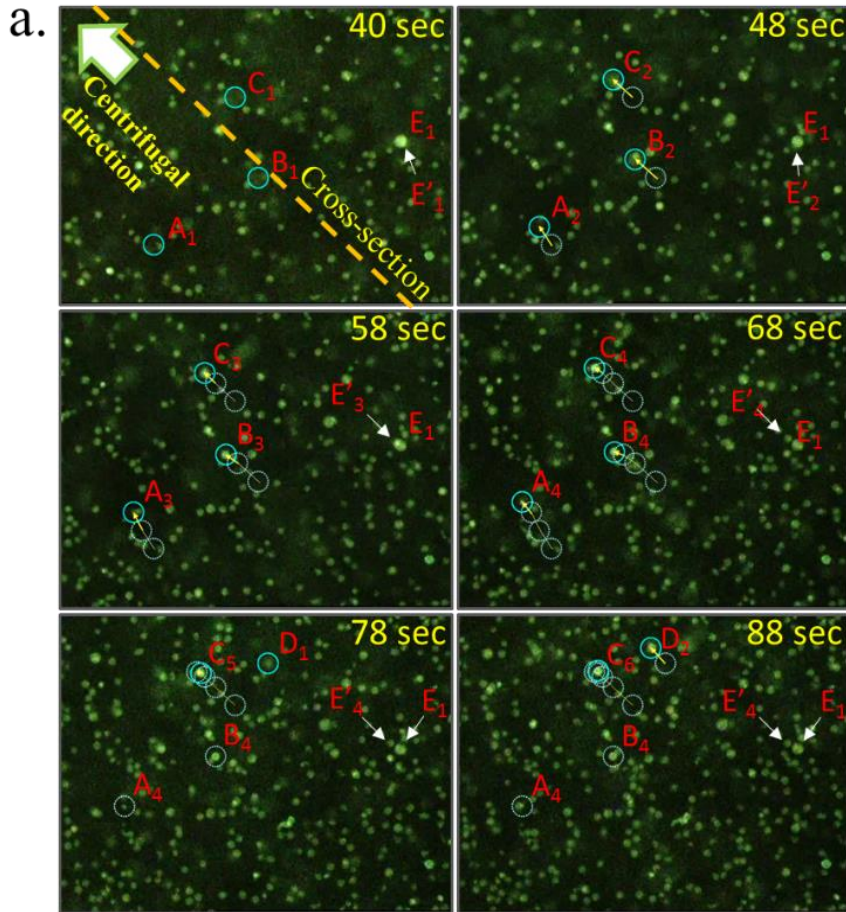


T.J Chang, and FG Tseng, *Biomicrofluidics*, 2014

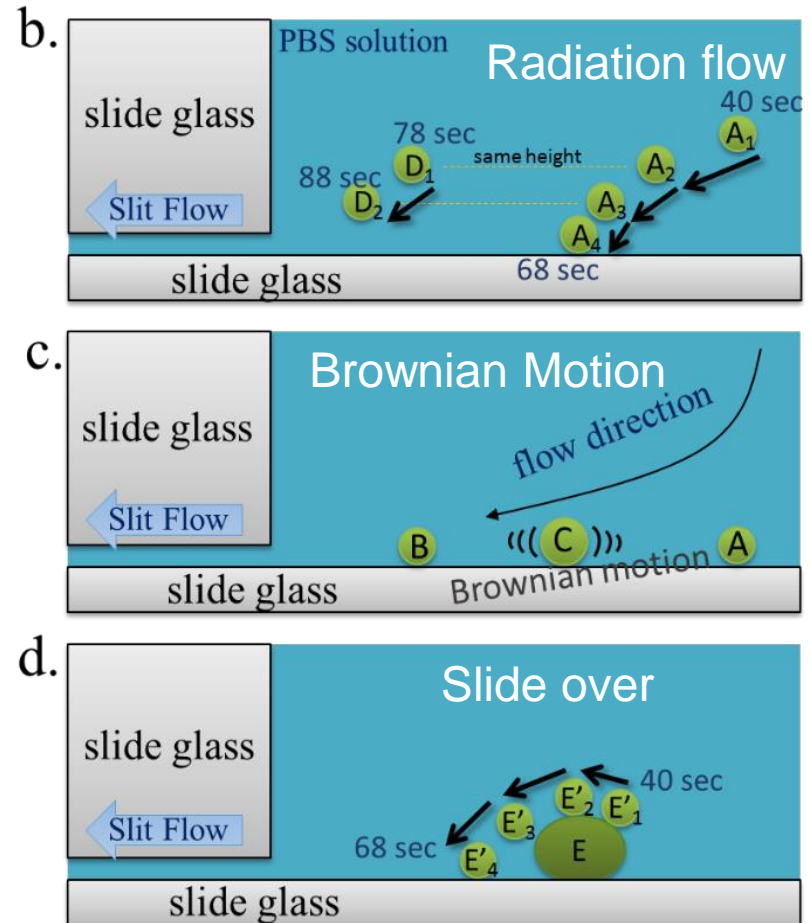
Prof. Fan-Gang Tseng

Mechanism for Cell Self-Assembly

Bottom view



Cross-section side view

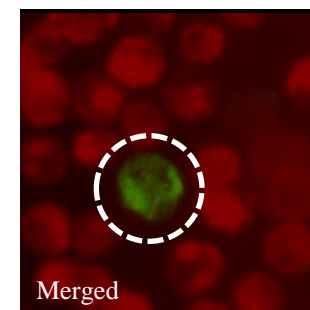
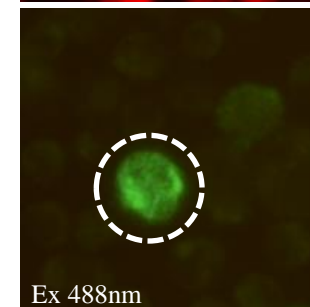
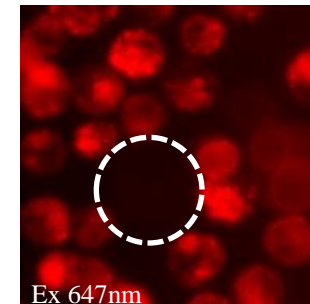
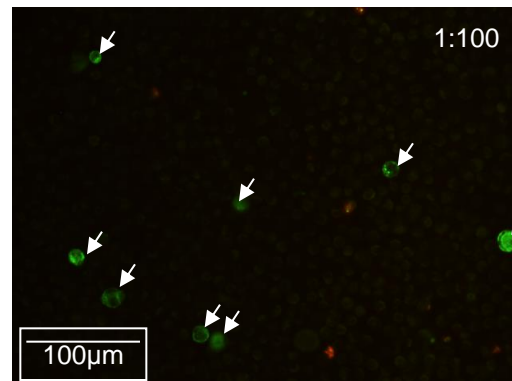
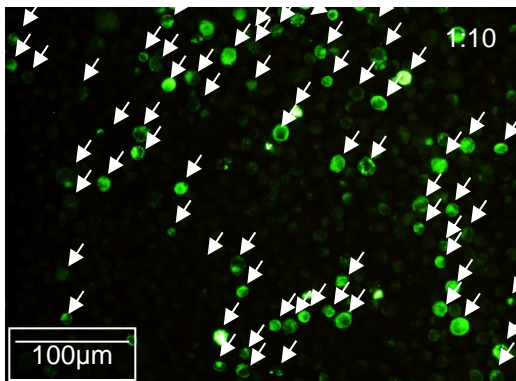
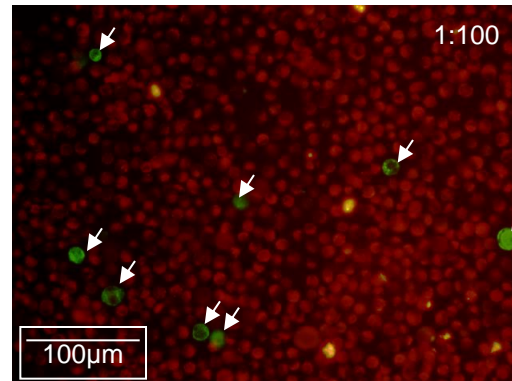
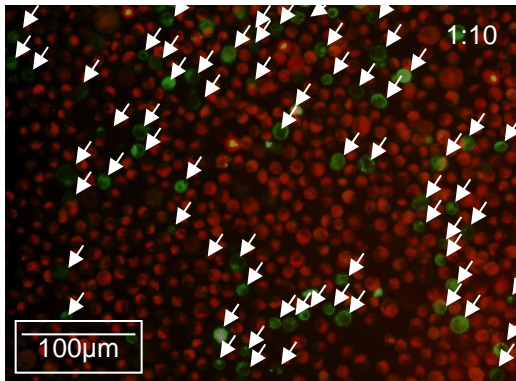




Two different cells

Target cells: HeLa with Alexa Fluor 488nm (green)

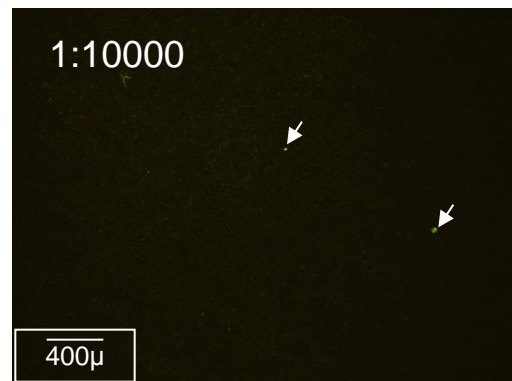
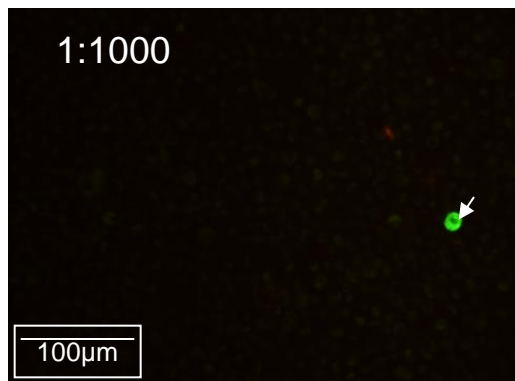
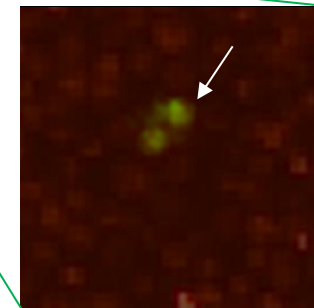
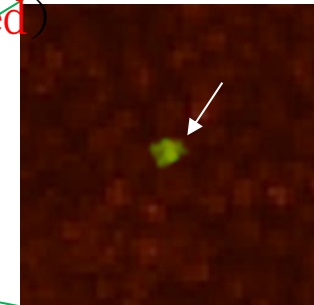
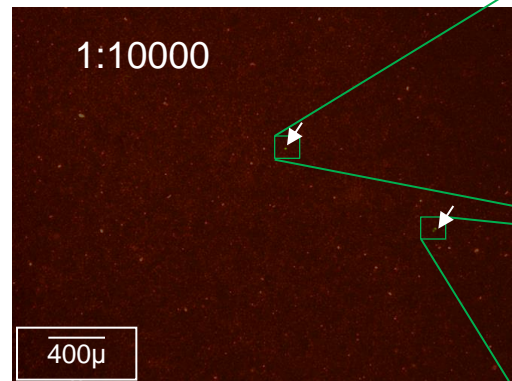
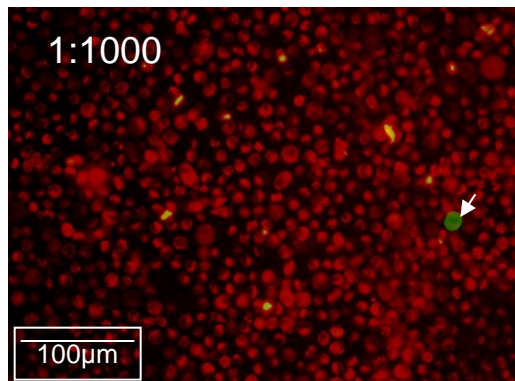
Background cells: MCF-7 with Alexa Fluor 647nm (red)



Two different cells (cont.)

Target cells: HeLa with Alexa Fluor 488nm (green)

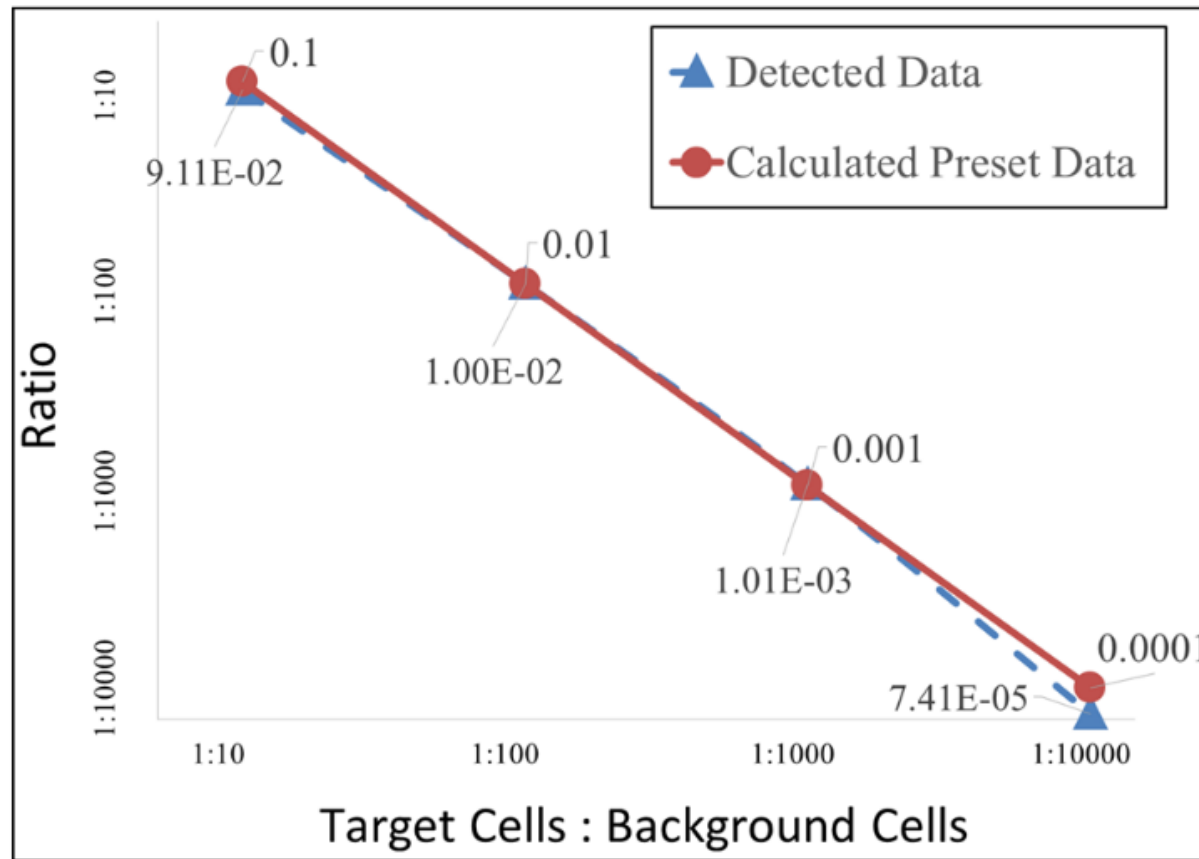
Background cells: MCF-7 with Alexa Fluor 647nm (red)



- Easy to identify target cells

Distinguish ratio reaches: 10000:1

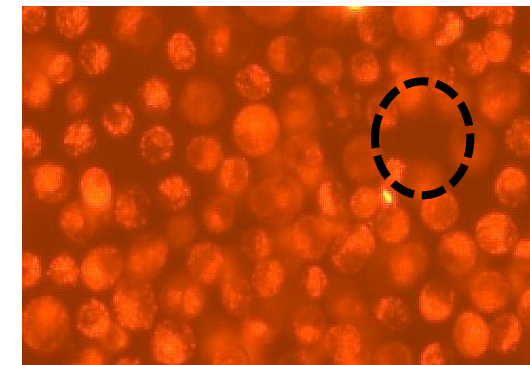
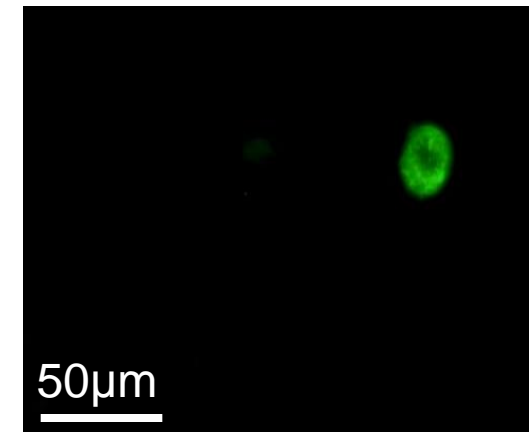
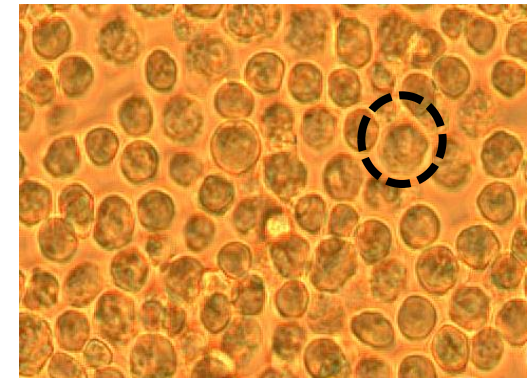
Distinguishing Ratio



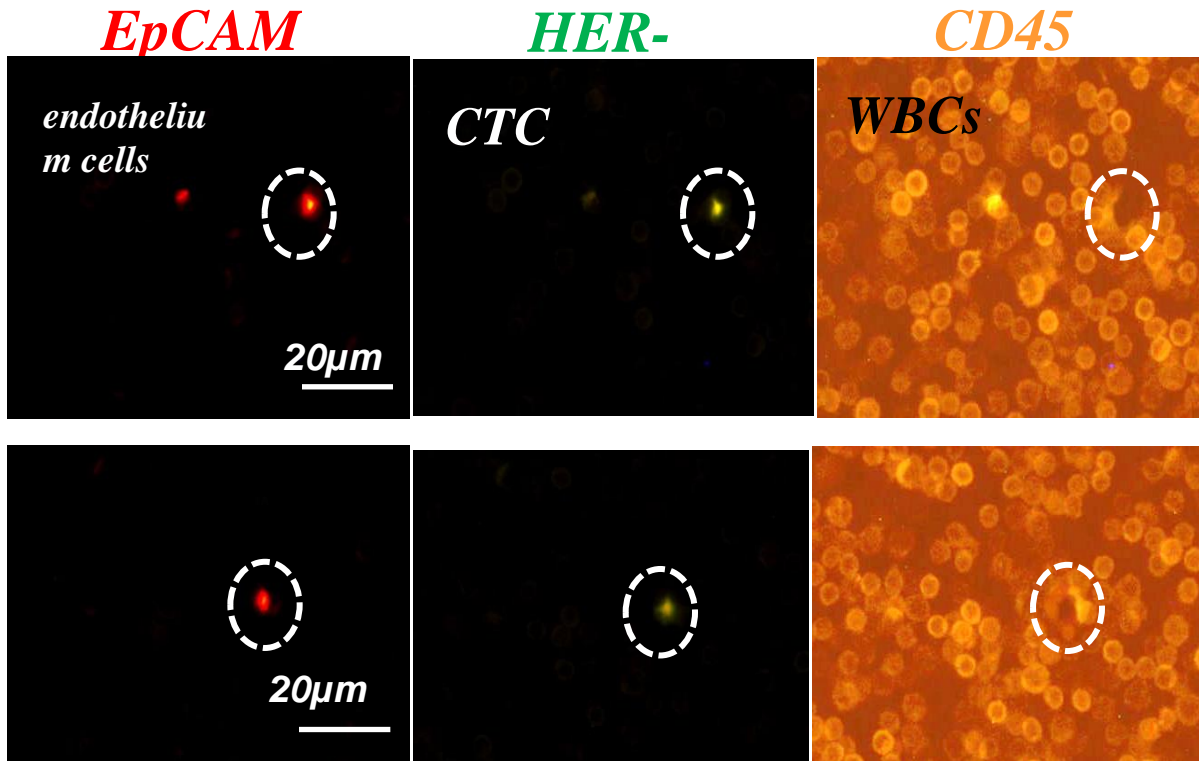
More than 10,000 to 1

Sensitivity Test

HeLa: WBC	Mixed HeLa cells number	Detected HeLa cells number	Average deviation value
1:10,000	29	27	8%
	34	30	
	41	38	
1:100,000	35	33	9%
	39	34	
	42	38	
1:1,000,000	34	30	12.7%
	36	31	
	39	34	
1:10,000,000	33	29	15.2%
	37	30	
	38	32	



CTCs Detection from Real Patient Samples (VGH, Taiwan)



Stage VI breast cancer patient (under treatment)

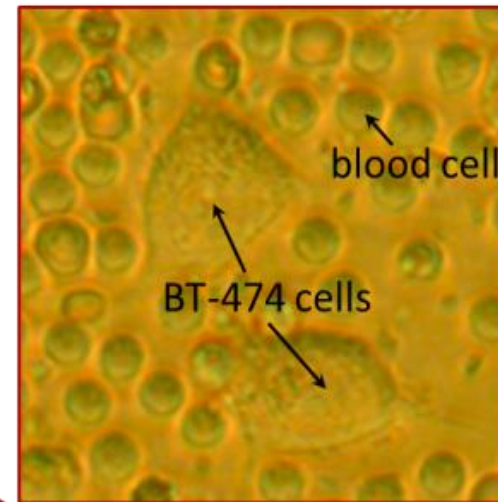
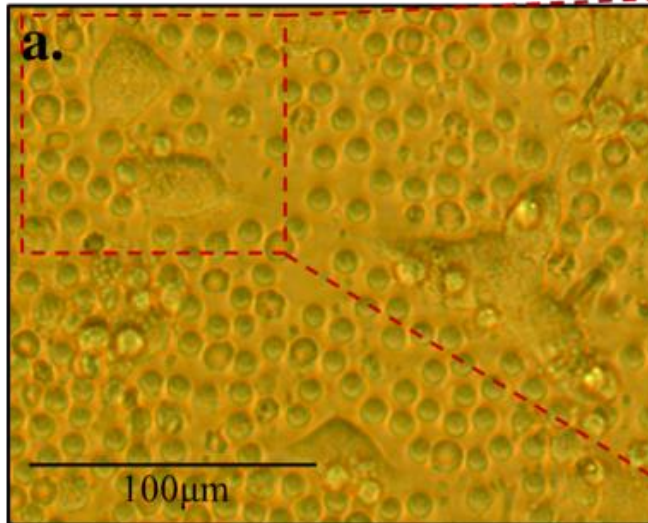
Spotted average 5 CTCs in wells
A well contain 10^6 WBCs (monolayer)

6×10^6 WBCs in 1 ml full blood

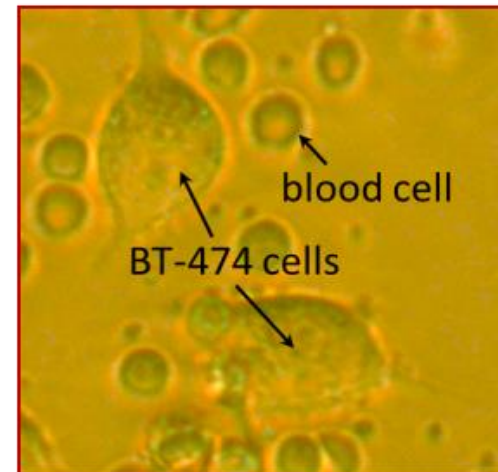
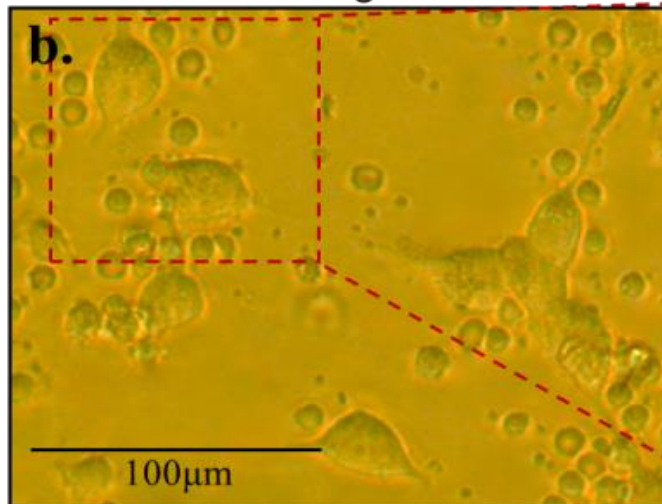
Predicted- 30 CTCs in 8 ml full blood

Tumor cells on-site Incubation

After 3 days incubation

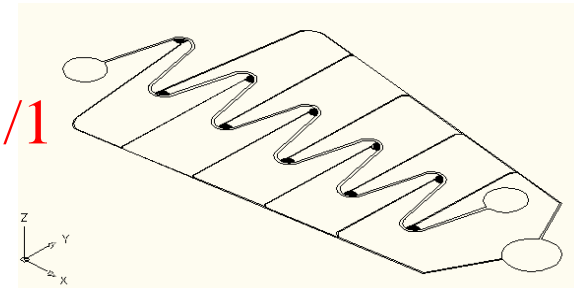


After PBS washing

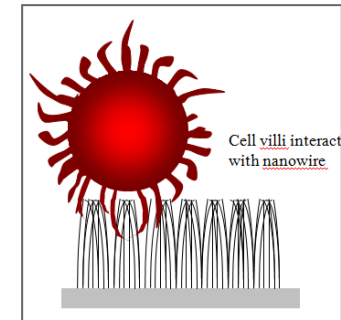


Conclusions

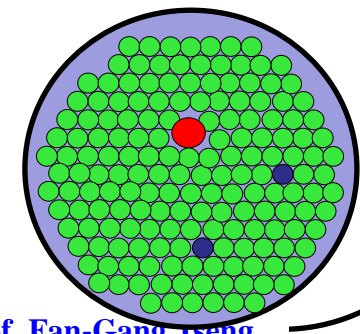
1. Blood cell separation: by using hydrodynamic and centrifugal forces,
 RBC/WBC: $1000/1 \Rightarrow 60/1 \Rightarrow 15/1 \Rightarrow <1/1$
 (goal)



2. CTC enrichment: by using Nanobundle arrays,
 CTC/WBC: $1/10^7 \Rightarrow \sim 1/10^5$ (80%:2% retention)



3. High density Cell Arrays: cell self assembly in descending flow, Density $>10^4$ cells/mm² ($>10^6$ cells/cm²), Distinguish ratio $1/10^6 \Rightarrow 1/10^7$





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4. **VUST Interdisciplinary Project, Taiwan**

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Thanks very much for Attention!

NTHU ESS

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Wei-Ru Tseng^b

Ren-Guei Wu^a

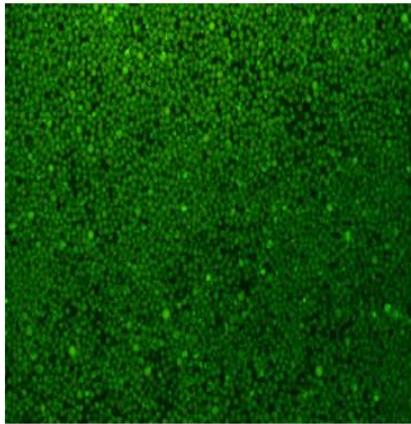
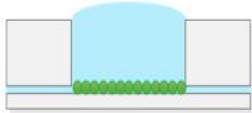
Yu-Cheng Chang^a



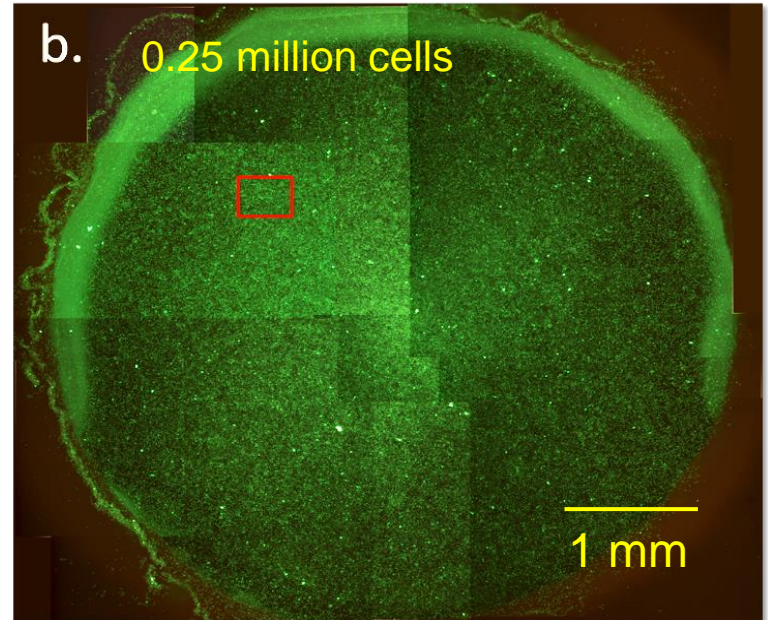
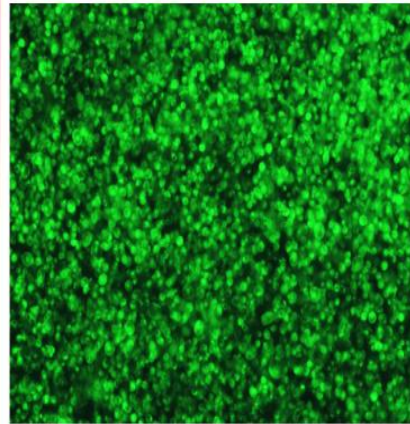
The importance of the Micro Slit



a well with slit channel



a well without slit channel



High cell arraying density in mono layer:

1,000,000 cells/cm²!!

