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UNIVERSITÀ  
DEGLI STUDI DI BARI  
ALDO MORO

Dipartimento di Chimica



# *In vitro* and *in silico* minimal cell models

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2<sup>nd</sup> International Summit on

**Integrative Biology**

August 04-05, 2014 Hilton-Chicago/Northbrook, Chicago, USA



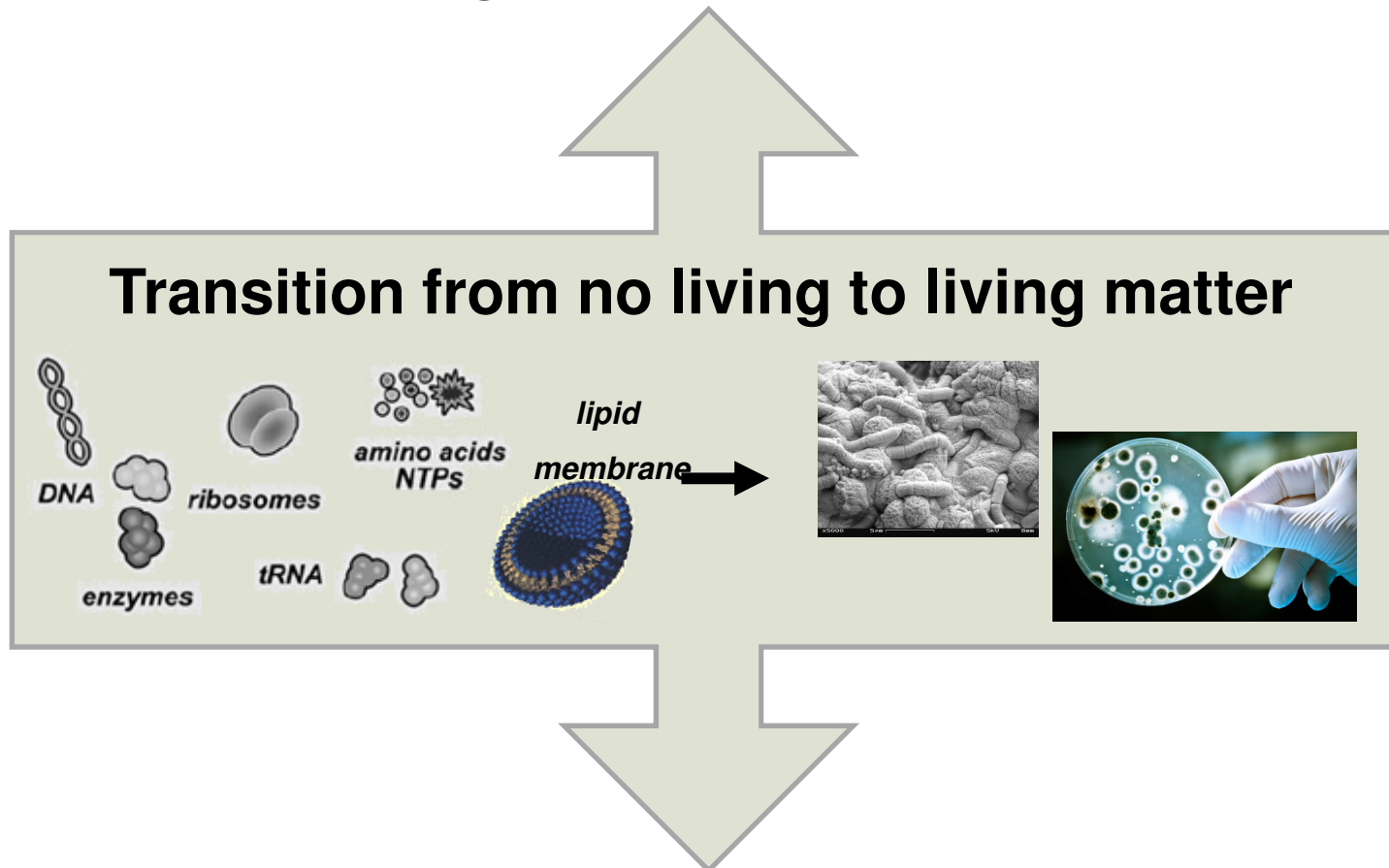
# Outline

- Introduction
  - Notion of minimal cell
  - Stochastic vs Deterministic models
- In vitro models
  - Polymer-Enzymes Complex for cascade reactions
  - Membrane Pore formation
  - Photosynthetic Reaction Renter membrane recostitution
- In silico models
  - Osmotic Synchronization
- Conclusions



Is it possible to construct a simplified cell from separated molecules? (\*)

## Origin of Life On Earth



## Emergence of Life in Test Tube

(\*) Luisi's group, ETH Zurich and Roma3 Univ.



# The notion of minimal cell

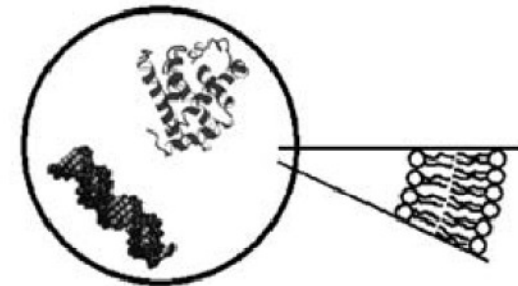
*“...the one having the minimal and sufficient number of components to be called alive. What does “alive” mean?”*

*Living at the cellular level means the concomitance of three properties:*

- *self-maintenance (metabolism),*
- *self-reproduction,*
- *and evolvability.”*

*“A living system is a system capable of self-production and self-maintenance through a regenerative network of processes which takes place within a boundary of its own making and regenerates itself through cognitive or adaptive interactions with the medium.”*

The notion of “minimal cell”



containing the minimum and sufficient number of components to be “alive”

<b>ALIVE</b>	self-maintenance
	reproduction
	evolvability



# Semi-synthetic approach to MC

modern living cells



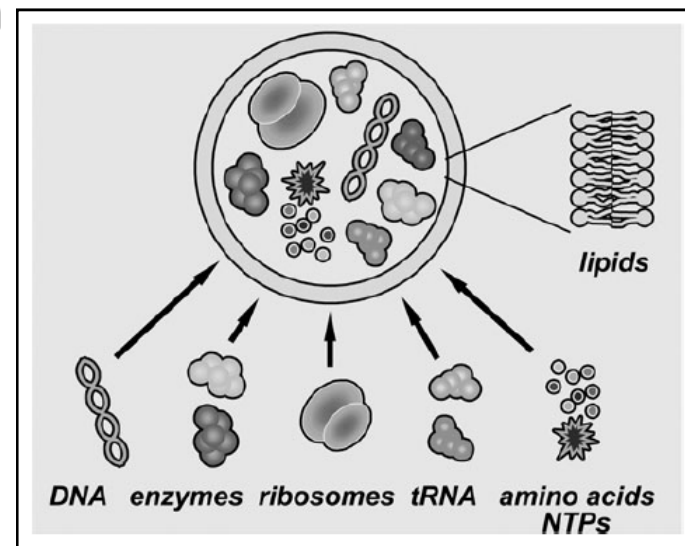
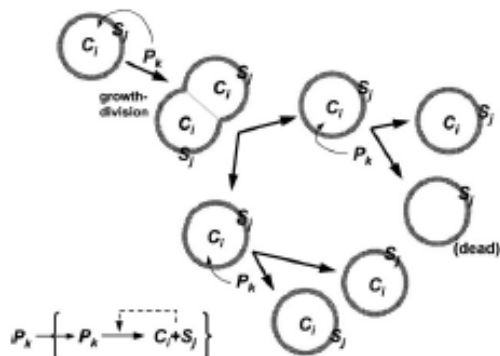
reducing  
complexity

Minimal Cell  
(minimal life)



bottom-up  
approach

simple molecules





# Aim of Modeling

Bridging the gap between *in silico* and *in vitro* experiments in the bottom-up approach to the minimal artificial cell



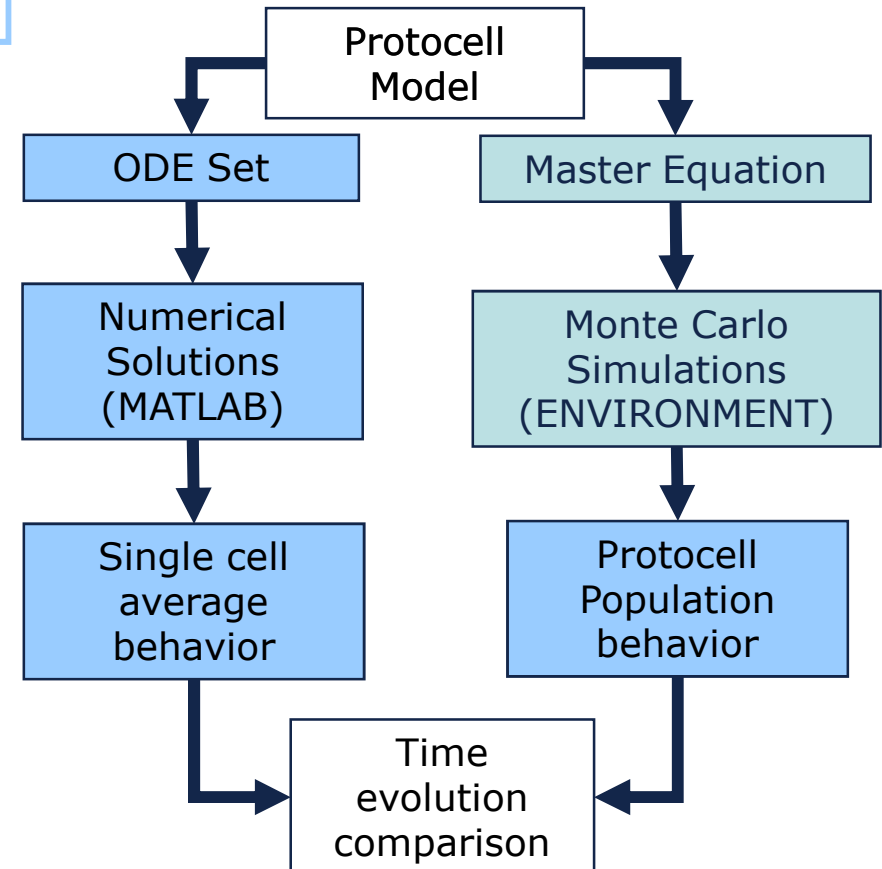
Theoretical approaches:

- **Deterministic**
- **Stochastic**

Theoretical models have been developed in order to analyse the feasibility of minimal cell models (*protocells*) able to exhibit:

- Self-maintenance
- Self-reproduction
- Evolution

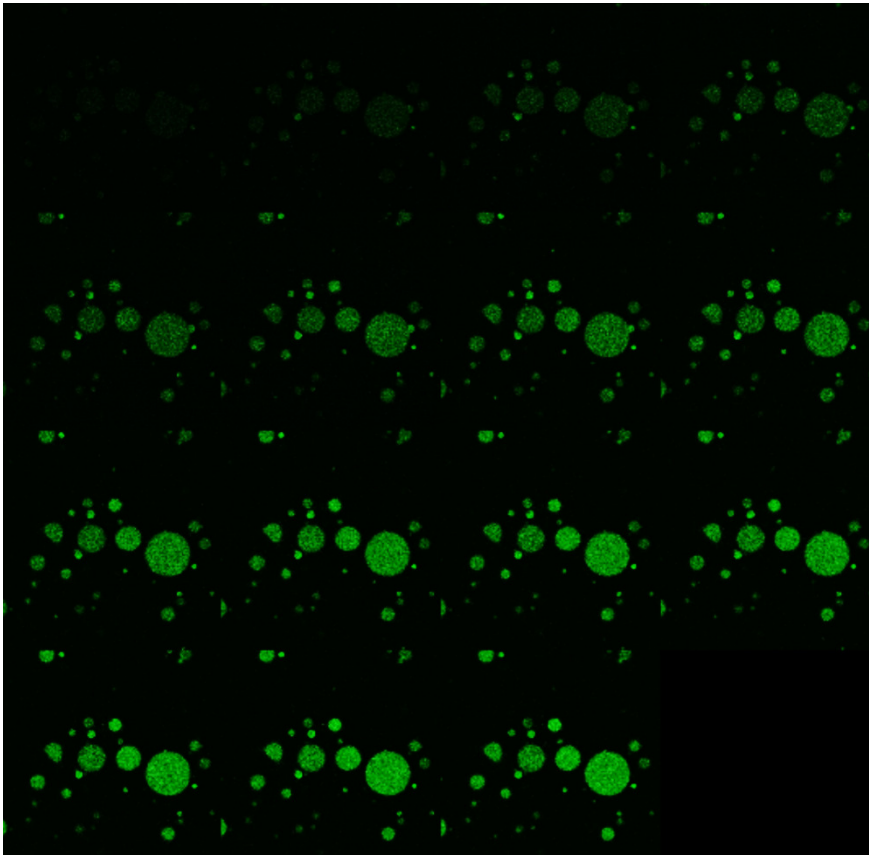
Deterministic	Stochastic
Macroscopic	Microscopic
Positive real numbers of Molecules	Integer numbers of Molecules
Reaction Rates	Propensity Probabilities
Average Behaviour	Average + Fluctuations







# GFP Expression in a w/o emulsion

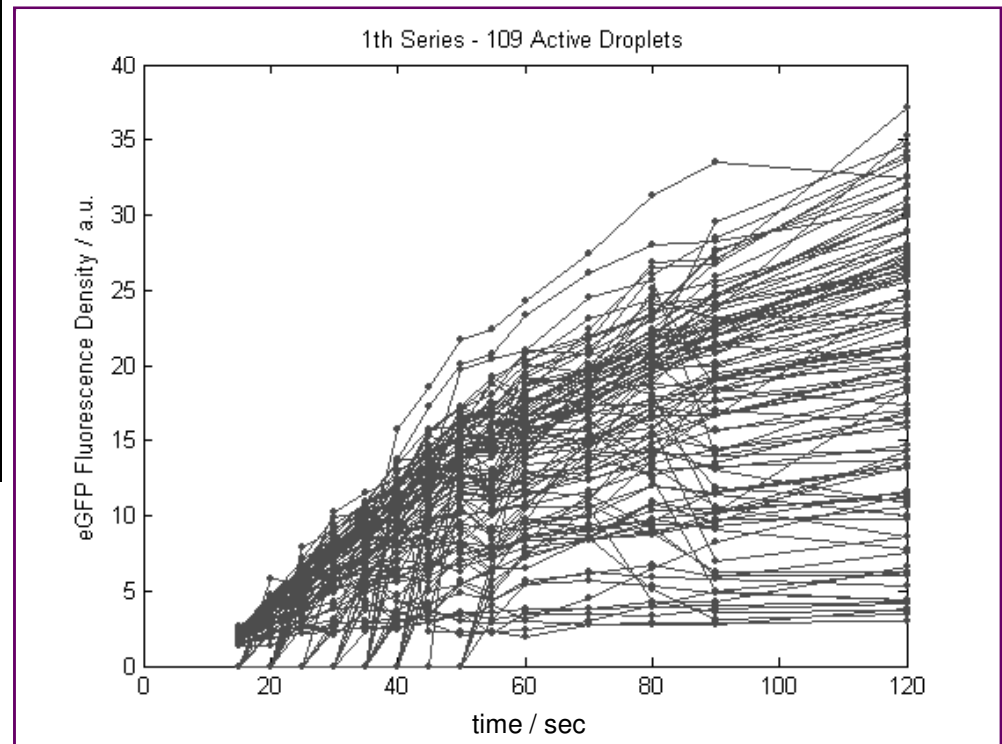


PURESYSTEM encapsulated in a water/oil emulsion

Data from Luisi's Lab experiments by P.Carrara, M.Caputo and P.Stano

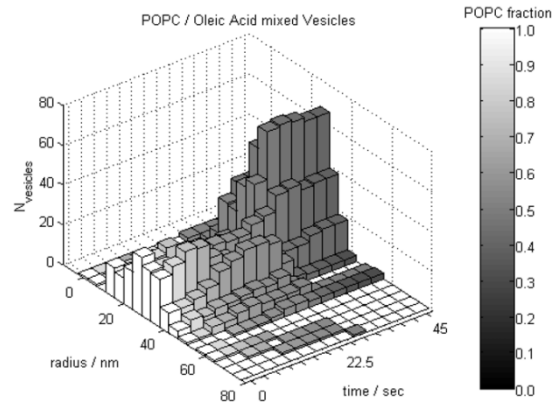
The time behavior of each compartment in the population could be highly affected by random fluctuations:

- Intrinsic effects
- Extrinsic effects



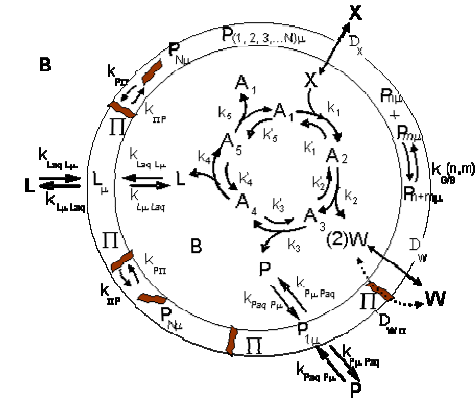


## Lipid Vesicle Dynamics



Mavelli F., Ruiz-Mirazo K.,  
Phys Biol 7, (2010), 36002

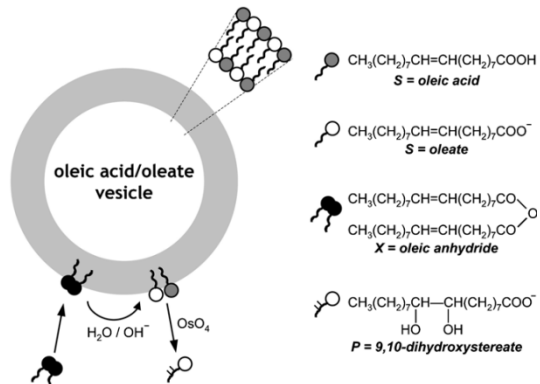
## Minimal Petide Cell



Ruiz-Mirazo K., Mavelli F.,  
*Biosystems* 91, (2008), 374



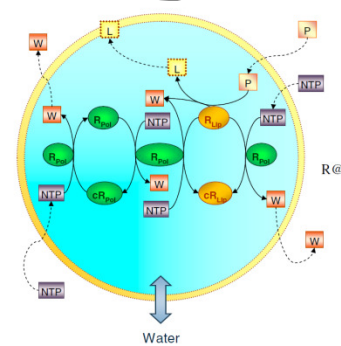
## Autopoietic Vesicles



Mavelli F., Stano P.,  
Phys Biol 7, (2010), 16010

Mavelli F., Ruiz-Mirazo K.,  
Phil. Trans. R. Soc. Lon. B  
362, (2007), 1789.

## Ribocell Model



Mavelli F. (2012) BMC Bioinformatics  
13, supp4, S10



# In vitro modeling



# Giant lipid vesicles (GVs)

## Features:

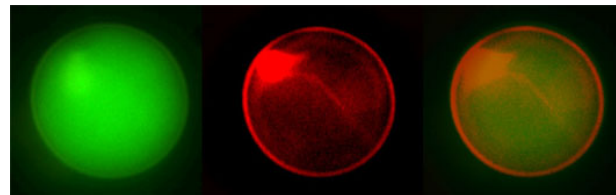
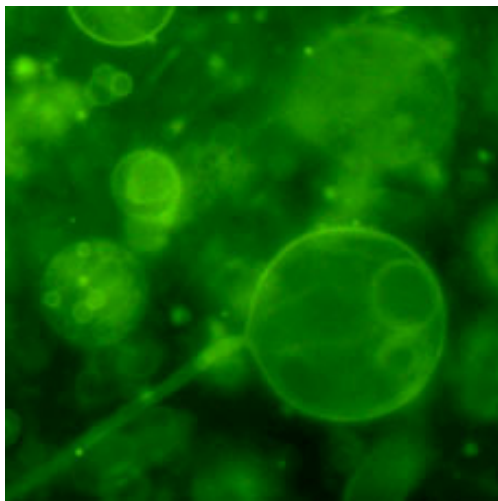
- Cell-like size
- Large encapsulation volume
- Single vesicle analysis
- Direct visualization by microscopy techniques
- Use of High-throughput analysis (flow cytometry)

## Conventional preparation methods:

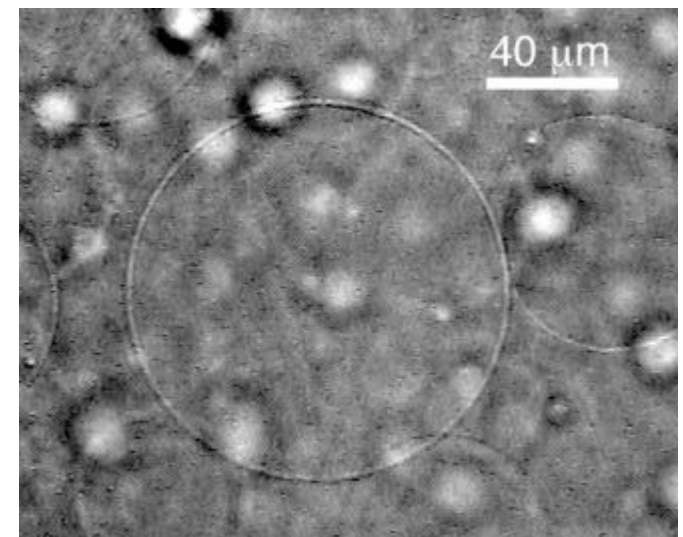
- Natural swelling
- Electroformation

## New Method

- Phase Transfer Method



*Giant vesicles (1-100 $\mu$ m)*

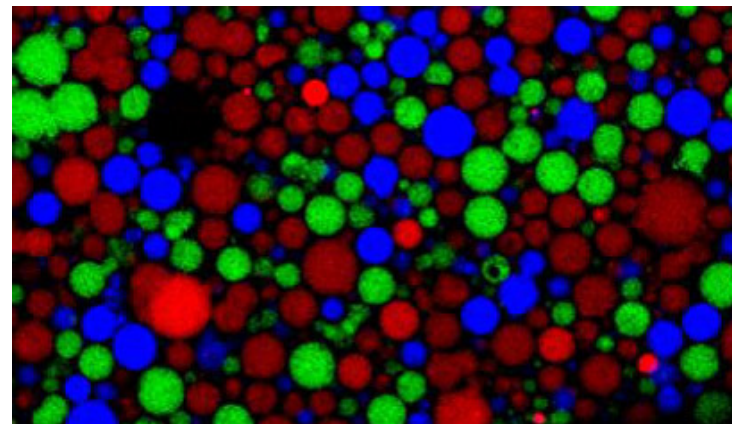
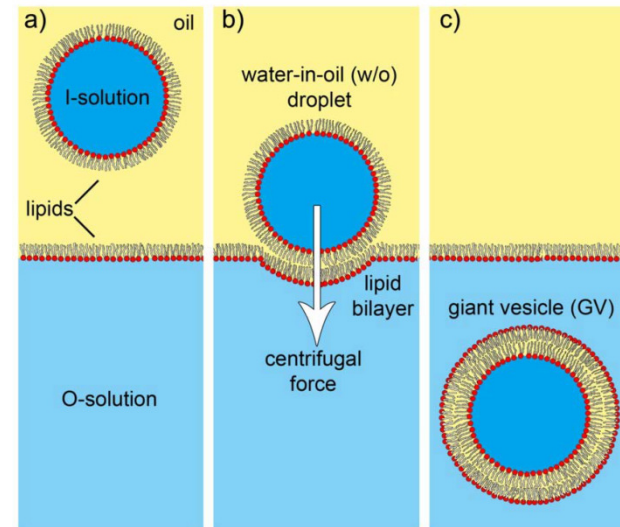
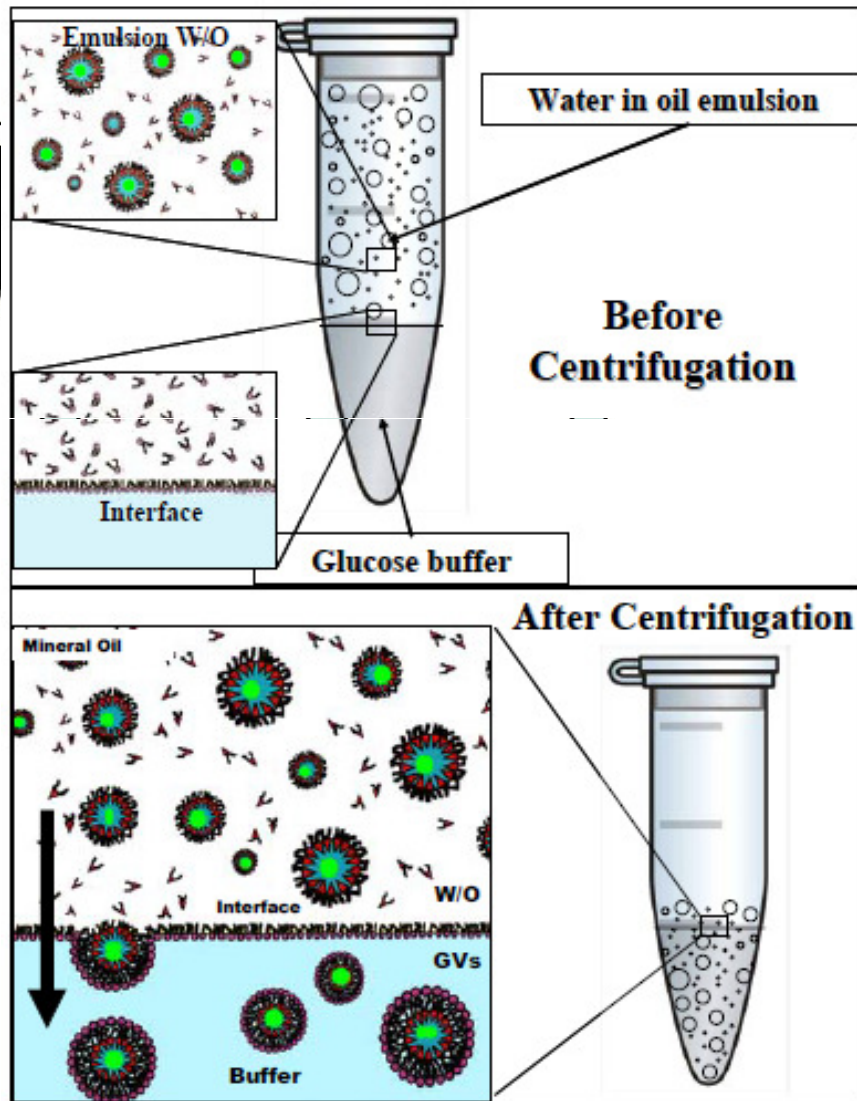




# The “Droplet transfer method”

## The “droplet transfer” method

Pautot et al., *Langmuir* 2003; *PNAS* 2003



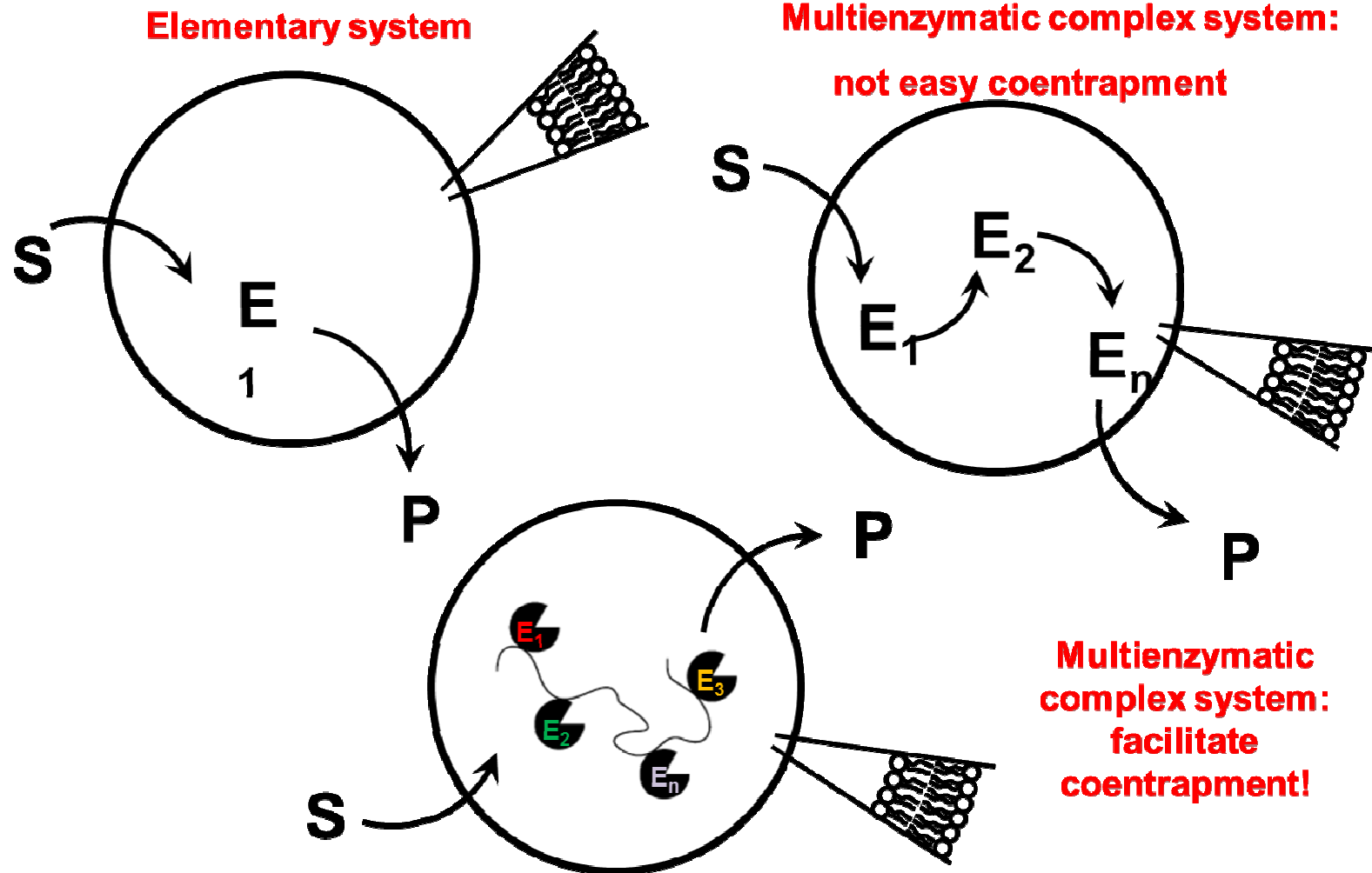
Carrara, Altamura, Stano, Luisi, *submitted*





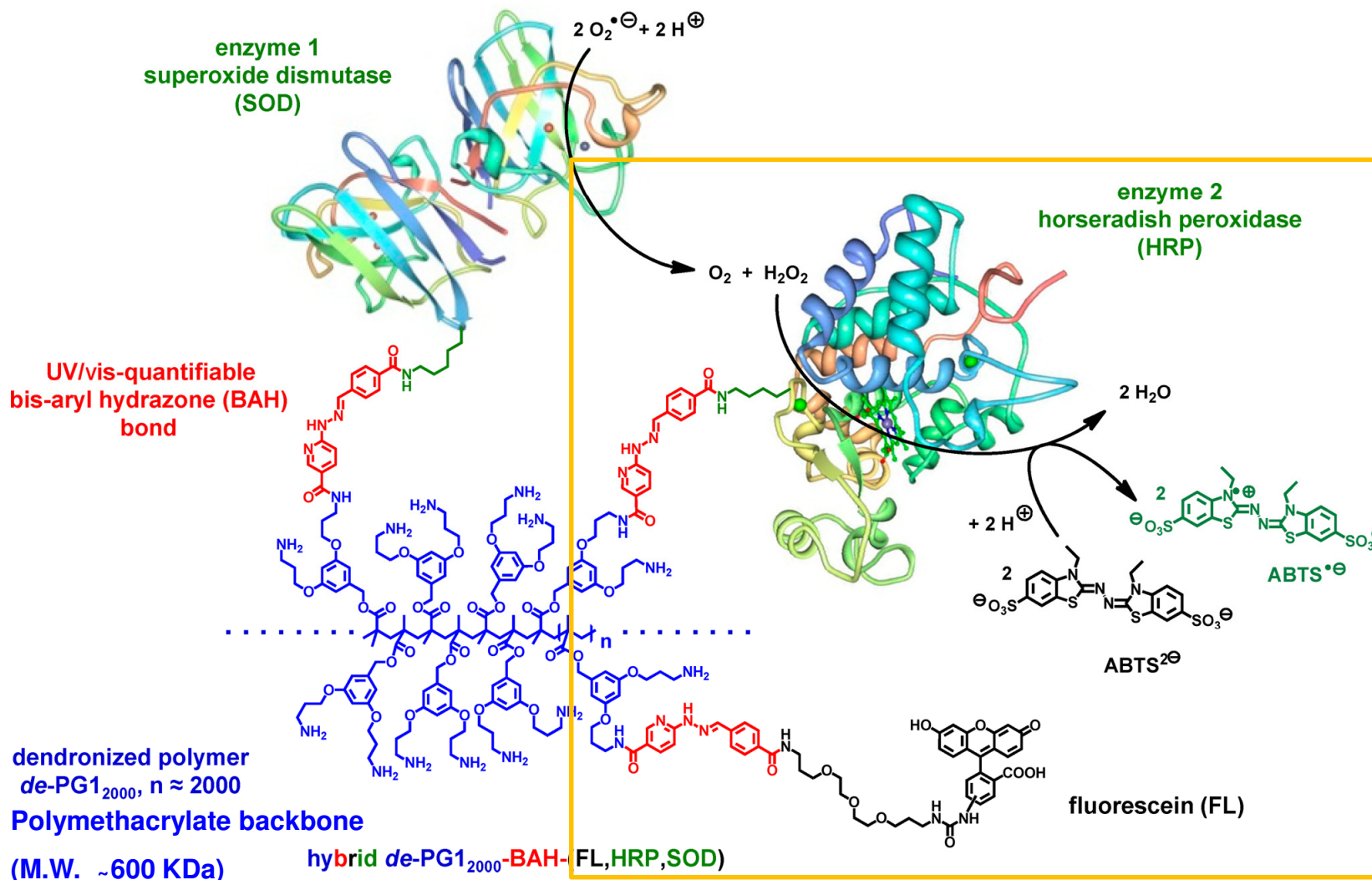
# GVs as biochemical reactors

## Improving enzyme encapsulation



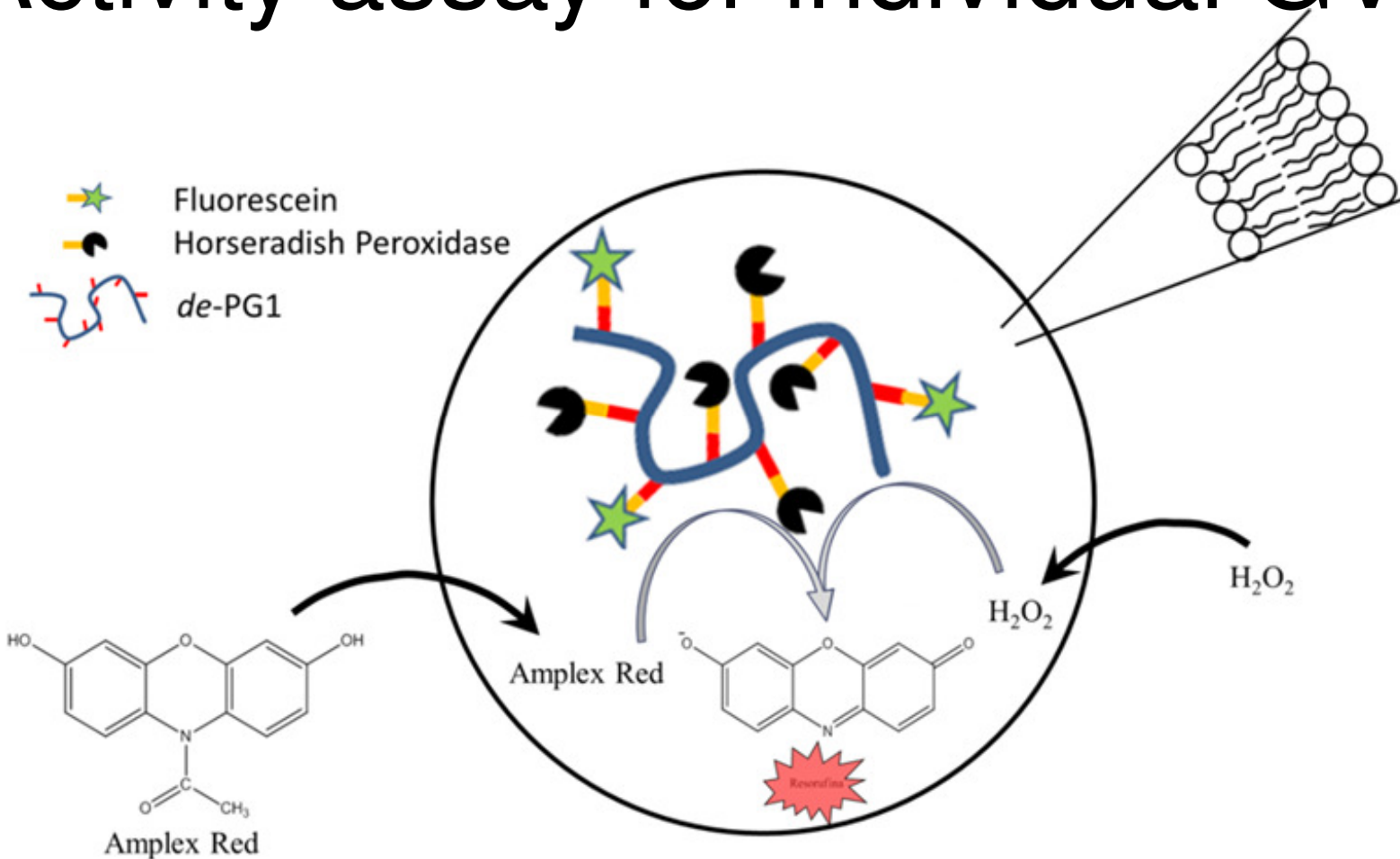


# de-PG1-(BAH)-FL-HRP-SOD synthesis:





# Activity assay for individual GVs



Enzymatic system





# *de*-PG1-FI-HRP entrapment

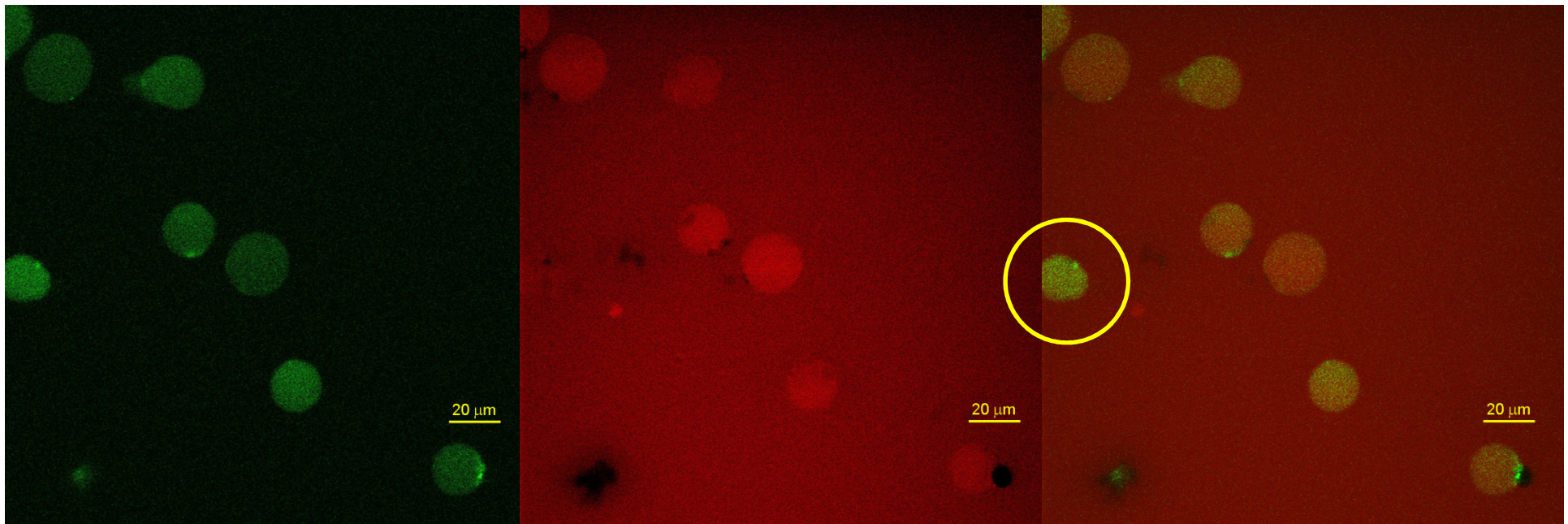
- [HRP] = 1  $\mu$ M,
- [Amplex Red] = 10  $\mu$ M,
- [H<sub>2</sub>O<sub>2</sub>] = 10  $\mu$ M

*Langmuir*

Article  
pubs.acs.org/Langmuir

Structure and Enzymatic Properties of Molecular Dendronized Polymer–Enzyme Conjugates and Their Entrapment inside Giant Vesicles

Andrea Grotzky,<sup>1</sup> Emiliano Altamura,<sup>1,2</sup> Jozef Adamec,<sup>3</sup> Paolo Carrara,<sup>4</sup> Pasquale Stano,<sup>5</sup> Fabio Mavelli,<sup>3</sup> Thomas Nausser,<sup>6</sup> Raffaele Mezzena,<sup>7</sup> A. Dieter Schlüter,<sup>7</sup> and Peter Walde<sup>6,7</sup>



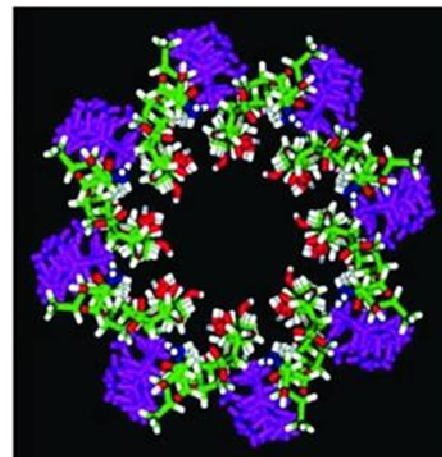
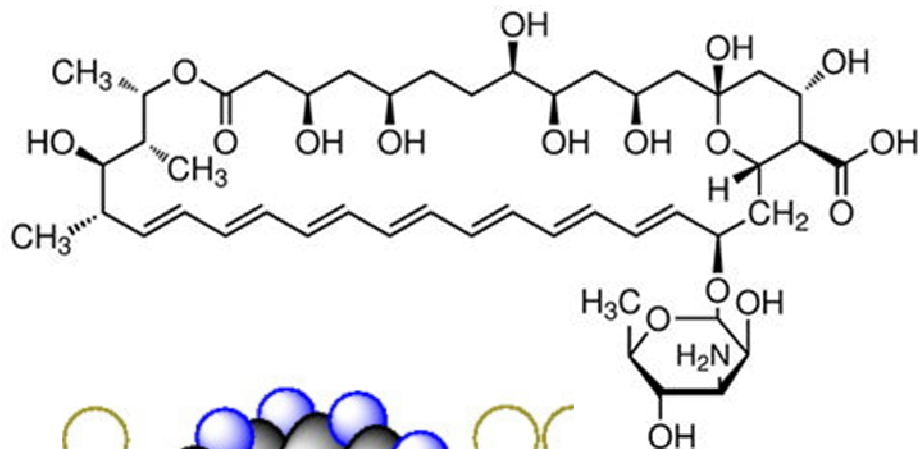
t = 200 s



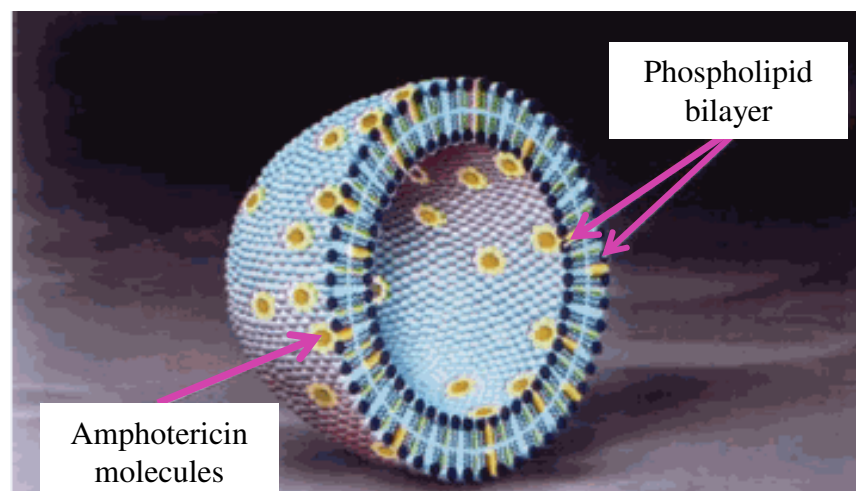
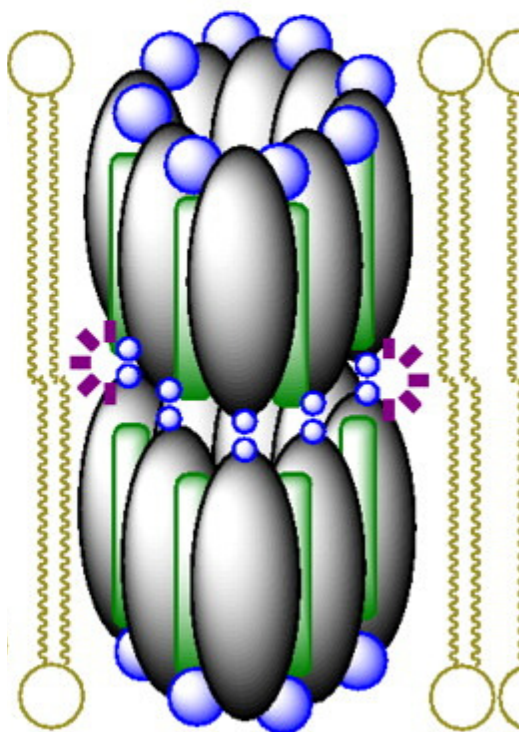
# Membrane engineering



# Pore formation: Amphotericin B



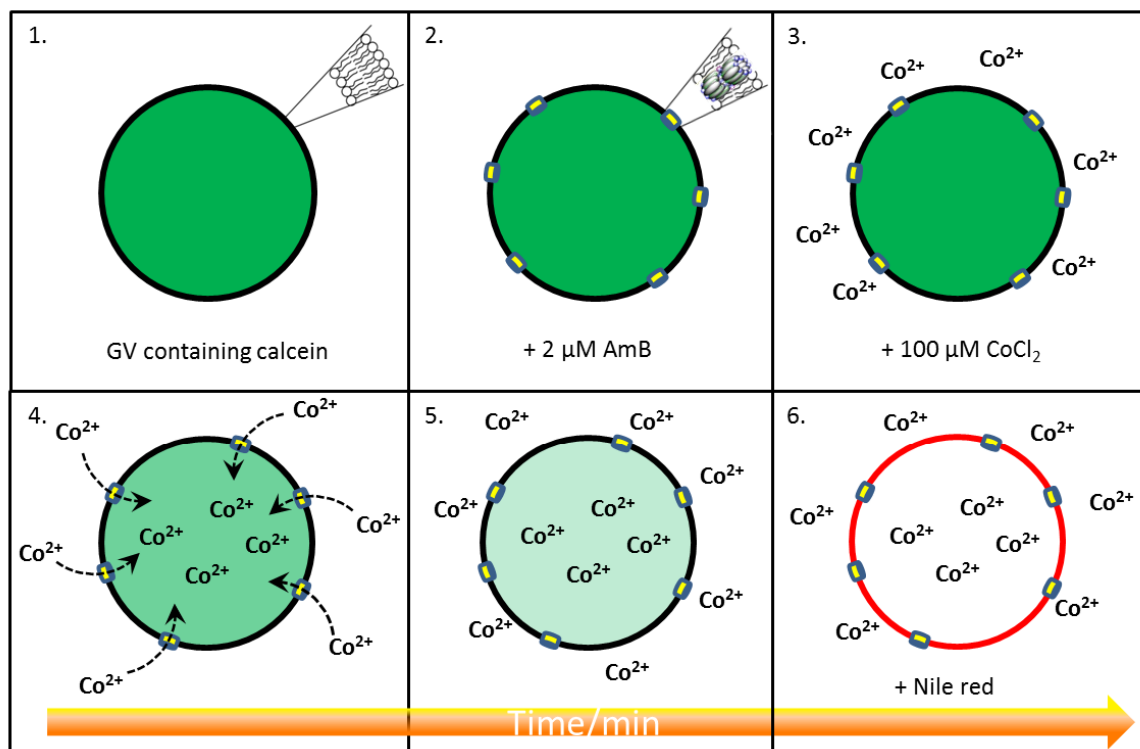
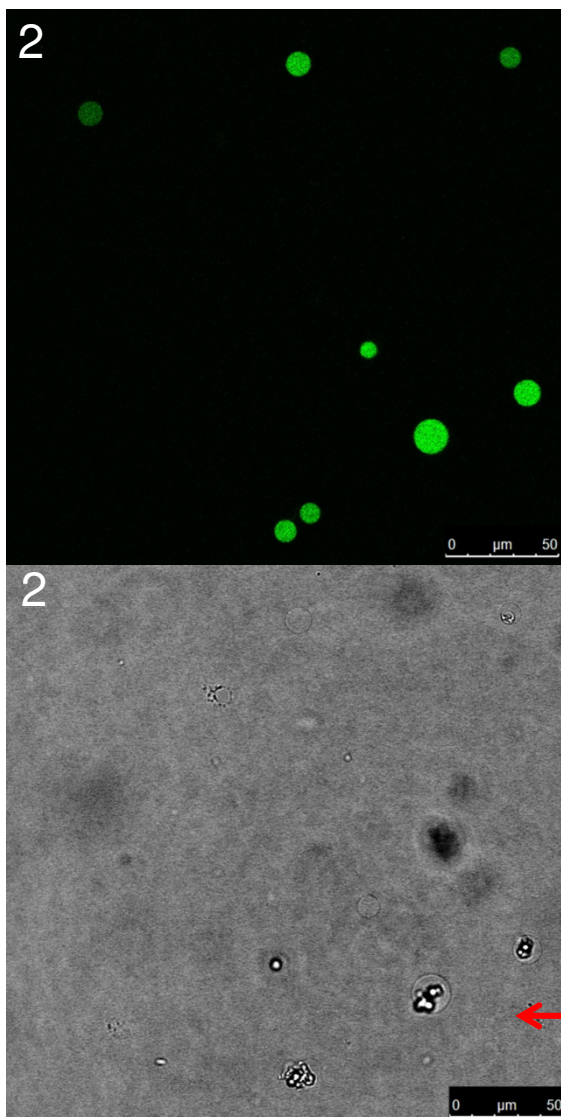
Among the variety of antifungal antibiotics Amphotericin B (AmB) is an amphiphilic molecule that forms pores in phospholipid membranes (Khutorsky 1996), allowing the diffusion of glucose, as experimentally demonstrated for nano-sized liposomes (Fujita *et al.* 2013).







# The assay for the pore formation

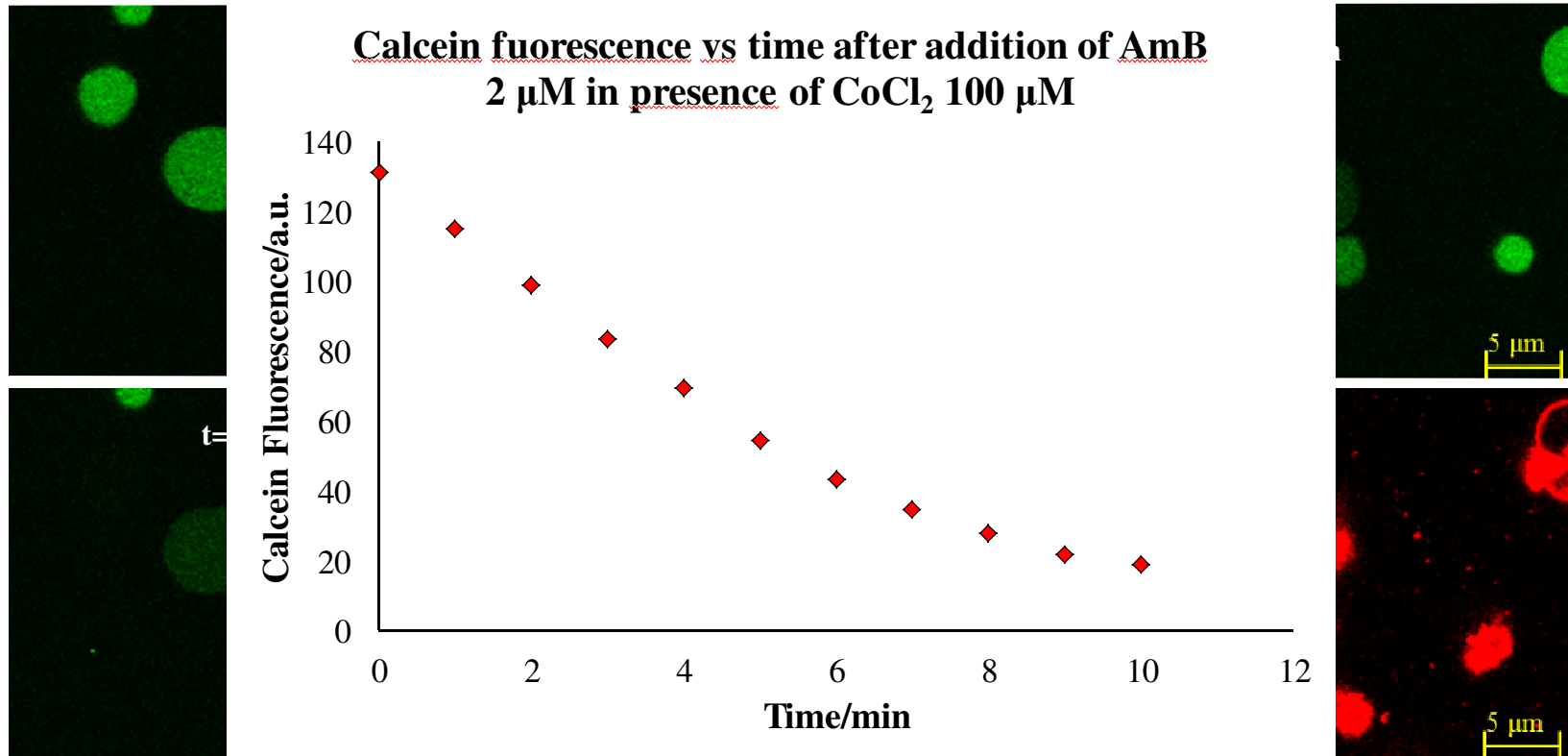


GV containing calcein and AmB pore formation, followed by internal calcein fluorescence extinction after addition of a  $\text{Co}^{2+}$  ions to the vesicle.

GVs encapsulating 10  $\mu\text{M}$  calcein after the addition of AmB 2  $\mu\text{M}$



# The assay for the pore formation

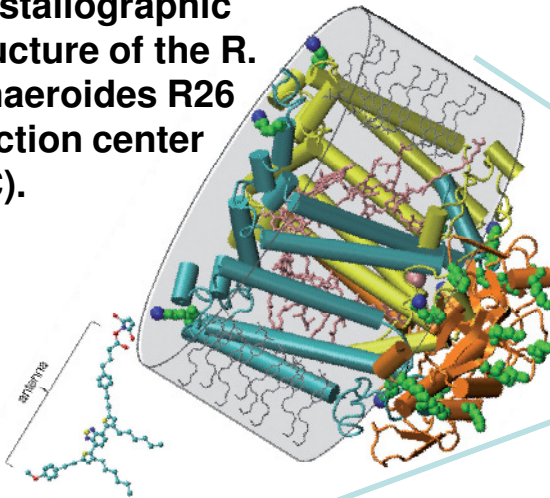


GVs containing Calcein 10  $\mu\text{M}$ . In the external buffer  $\text{CoCl}_2$  100  $\mu\text{M}$  is added as quencher. Time scan recorded after the addition of Amphotericin B 2  $\mu\text{M}$ , that is able to create pores through the membrane with an high dimensional selectivity. The graph indicates the decrease of calcein green fluorescence during time due to the quenching effect of the  $\text{Co}^{2+}$  ions.

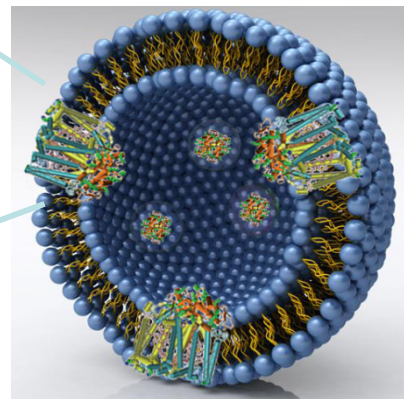


# Photosynthetic Reaction Center in GVs

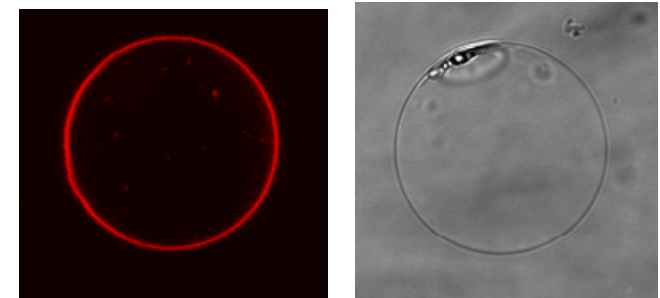
Crystallographic structure of the R. sphaeroides R26 reaction center (RC).



Reconstitution of the photosynthetic RC within the GV membrane

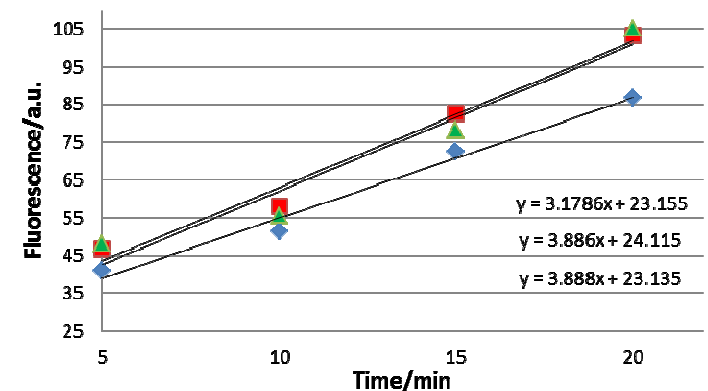
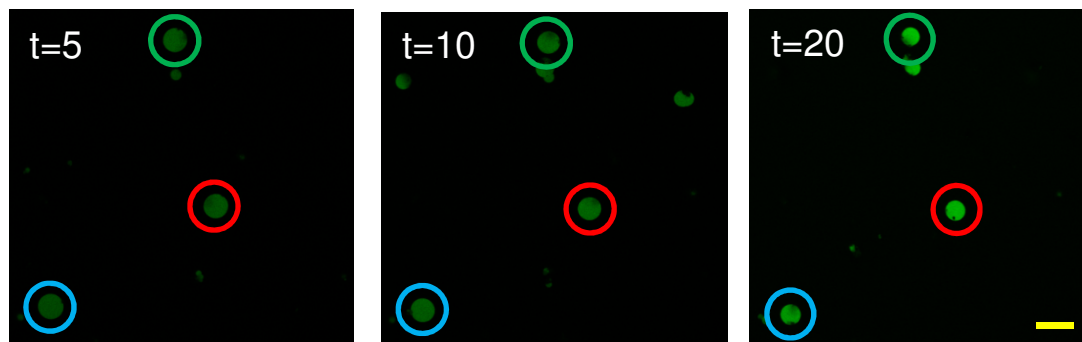


Confocal microscope images of GV made by POPC with RC reconstituted in membrane.



*Manuscript in preparation*

pH increase of the vesicle internal solution due to the activity of RCs monitored by the fluorescence of an entrapped dye (pyranine)



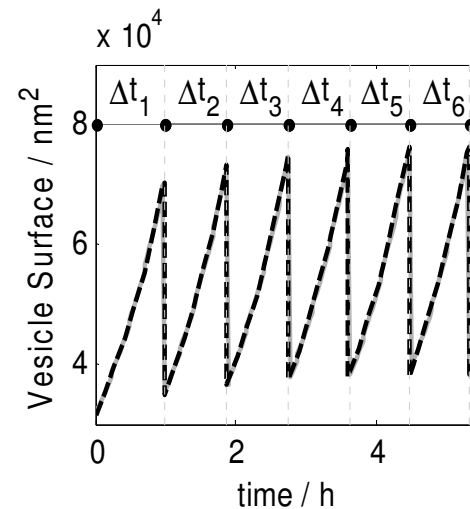
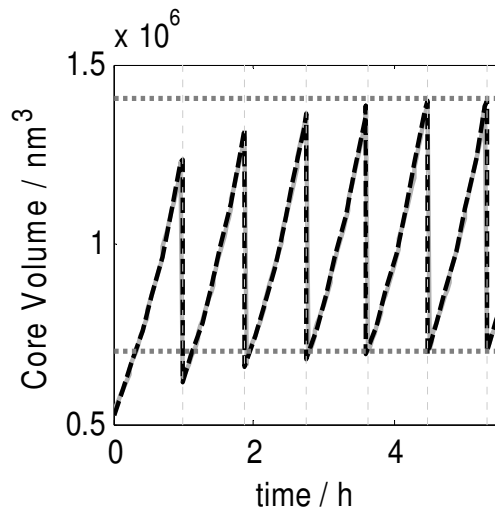
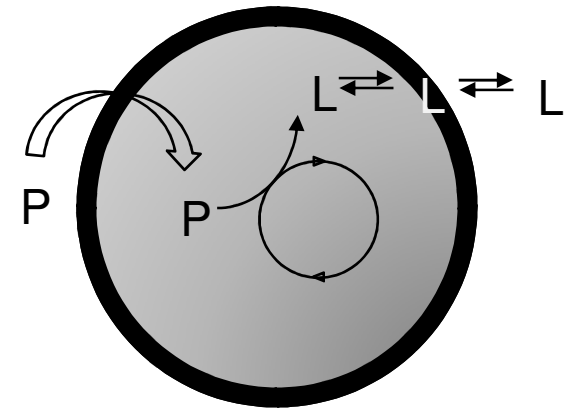


# In silico modeling



# Protocells stationary reproduction

We would like to determine the conditions that drive minimal self-producing vesicles to regular growth/division cycles, making possible that subsequent generations inherit size and internal chemical composition.







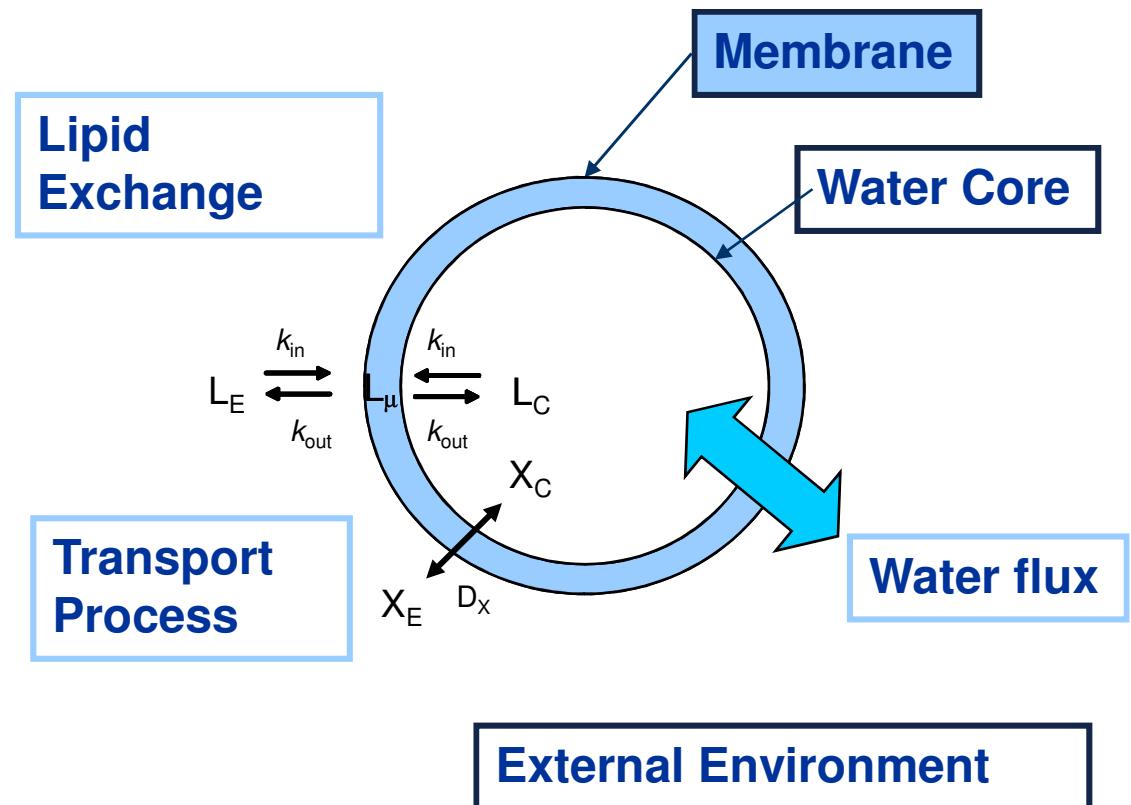
# In silico Vesicles

Vesicles are described as compartmentalized reacting systems (CSTR) made of two different homogeneous domains:

- the membrane
- the water core

Lipids and molecules can be exchanged between the membrane and water core, between the membrane and the external environment.

Transport processes can also occur, exchanging molecules directly from the external environment to the internal water pool.





# Stability of closed membrane

## Reduced Surface

$$\phi = S_{\mu} / \sqrt[3]{36\pi V_{core}^2}$$

ratio of the actual membrane surface  $S_{\mu}$  and the area of a sphere with the actual volume  $V_c$  of the core

### Vesicle

swollen

spherical

deflated

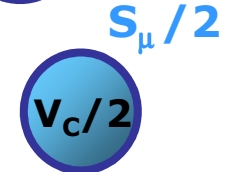
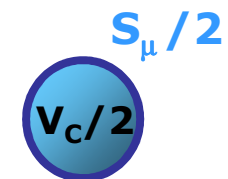
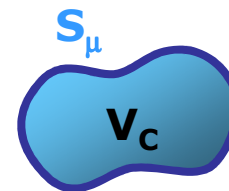
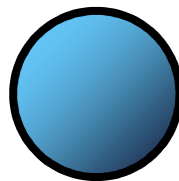
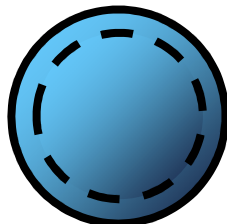
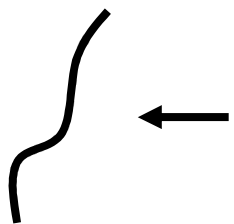
$$1 - \varepsilon$$

$$\leq$$

$$\phi = 1$$

$$\leq$$

$$\sqrt[3]{2}$$



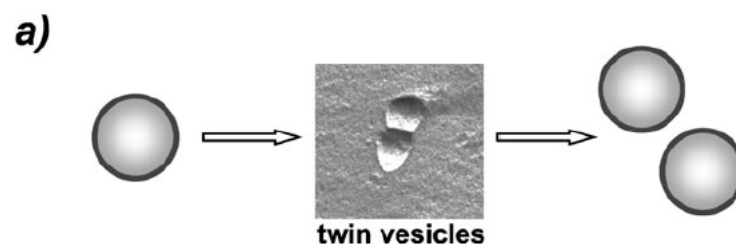
Osmotic  
Crisis

Division

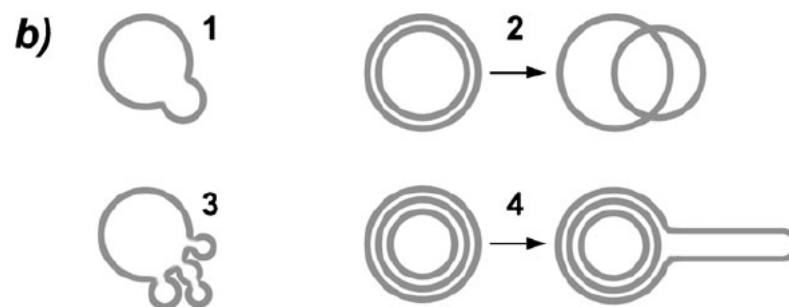


# Vesicle division

Small unilammellar vesicles



Giant uni-lammellar vesicles



Giant multi-lammellar vesicles

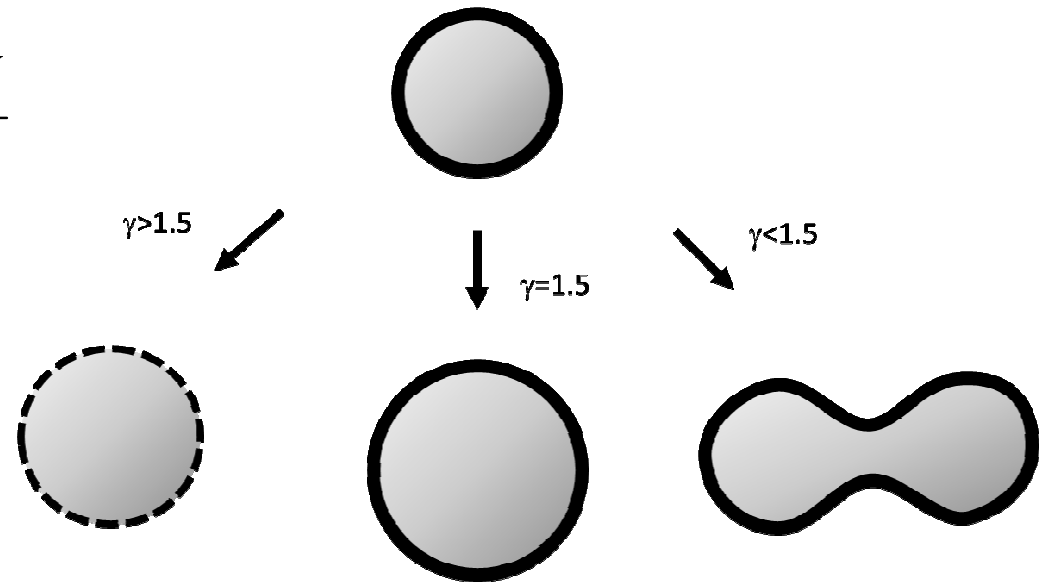
Fig. 5 Freeze-fracture electron microscopy of oleate vesicles (taken after 40 s from the addition of oleate micelles) reveals that “twin vesicles” can be the actual intermediate of the self-reproduction. Twin vesicles are not present at the end of the reaction. Adapted from Stano *et al.*<sup>29h</sup> Several intermediates have been observed in the case of the transformations occurring to oleate giant vesicles after the addition of oleate micelles: (1) budding mechanism;<sup>33a,b</sup> (2) translocation;<sup>33a,b</sup> (3) evagination;<sup>36b</sup> (4) tubular growth and division.<sup>36a</sup>



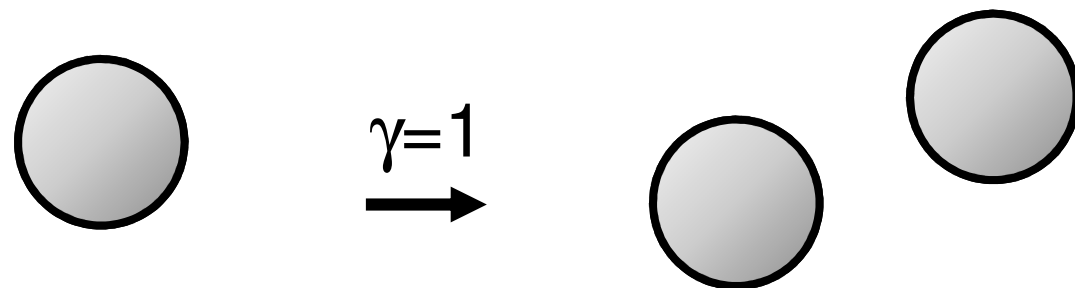
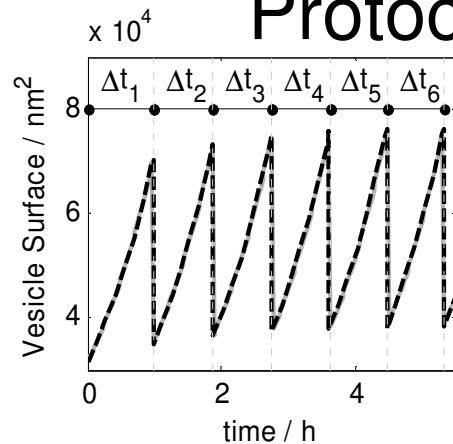
# Growth control coefficient

$$\gamma = \left( \frac{1}{V_g} \frac{dV}{dt} \right) / \left( \frac{1}{S_g} \frac{dS}{dt} \right) = \frac{S_g}{V_g} \frac{dV}{dS}$$

$\gamma$  is a dimensionless observable is defined as the ratio between the relative velocities of variation of core volume and membrane surface of a vesicle



## Protocells stationary reproduction





# Osmotic synchronization

$$\gamma = 1$$



$$\sum_{\rho}^{\text{Reactions}} \Delta m_{\rho} r_{\rho} + \frac{S}{V} \sum_i^{\text{Species}} \varphi_i ([X_i]_{Env} - [X_i]) - v_L = \frac{C^T \alpha_L}{2} \frac{N_A V}{S} v_L$$

$$v_R = \sum_{\rho}^{\text{Metabolic Reactions}} \Delta m_{\rho} r_{\rho}$$

different metabolic steps occurring inside the protocell, each at a particular rate  $r_{\rho}$ , with a net number of molecules produced or consumed  $\Delta m_{\rho}$

$$v_{Tp} = \frac{S}{V} \sum_i^{\text{Species}} \varphi_i ([X_i]_{Env} - [X_i])$$

net fluxes of molecules that can come in or escape across the membrane through passive transport

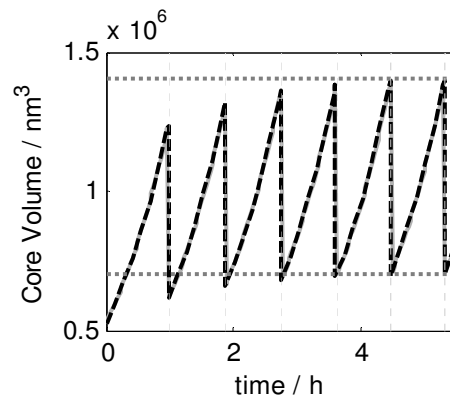
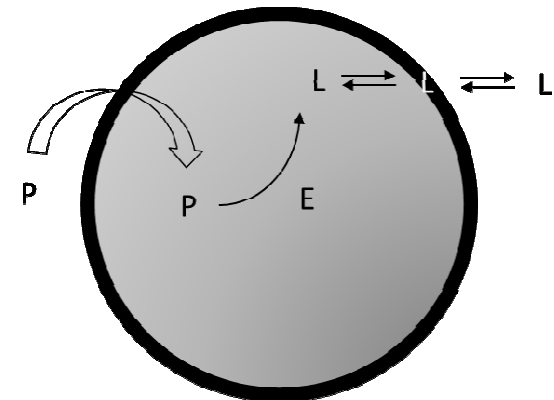
$v_L$

metabolic lipid production rate



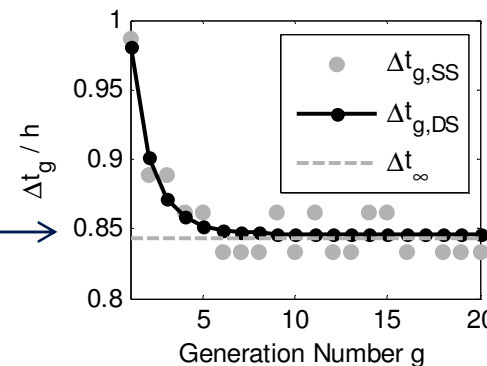
# Case Study

**Self-producing enzymatic vesicle:** a hypothetical model where the production of lipid L takes place through the chemical transformation of a precursor molecule P, assumed to occur only in the presence of an additional compound E,



$$R_{\infty} = \frac{6n}{\alpha_L N_A C^T}$$

$$\Delta t_{\infty} = \frac{2}{\alpha_L [P]_{Ext}} \left( \frac{\ln 2}{N_A \phi_P} + \frac{4\pi R_{\infty}^2}{k} \right)$$

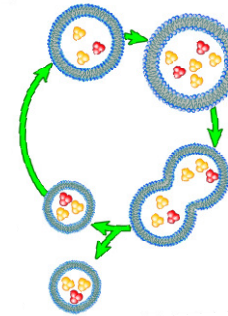
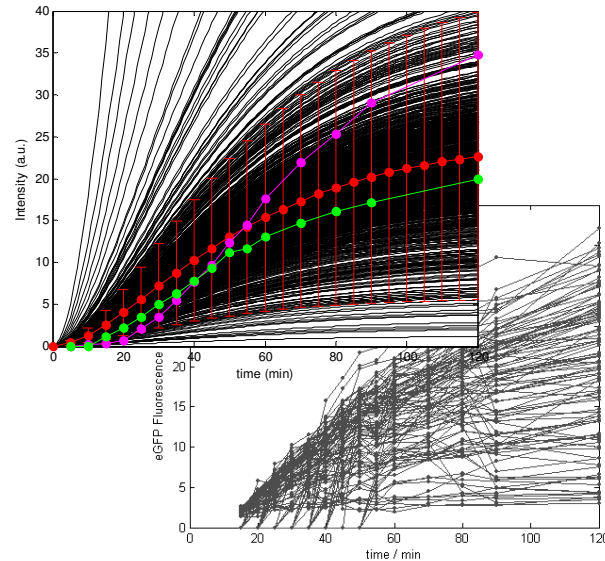
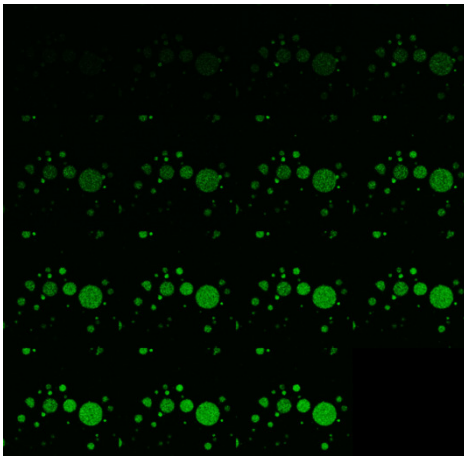


- Stochastic simulation outcomes (in grey)
- the deterministic curve by numerical integration (in black).

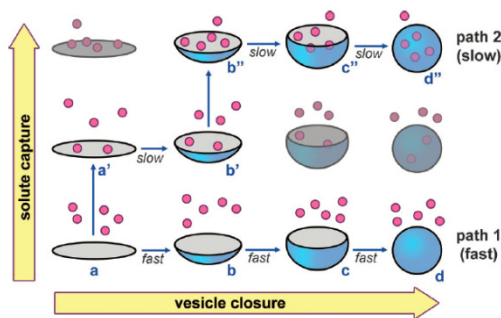
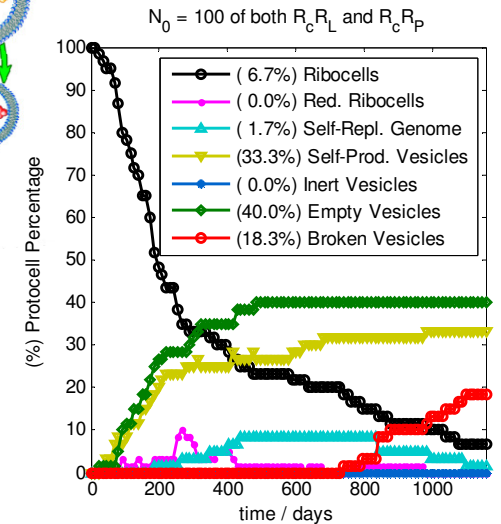


# Theoretical modelling

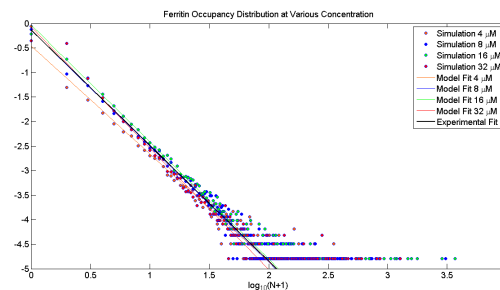
## GFP Expression in a w/o emulsion



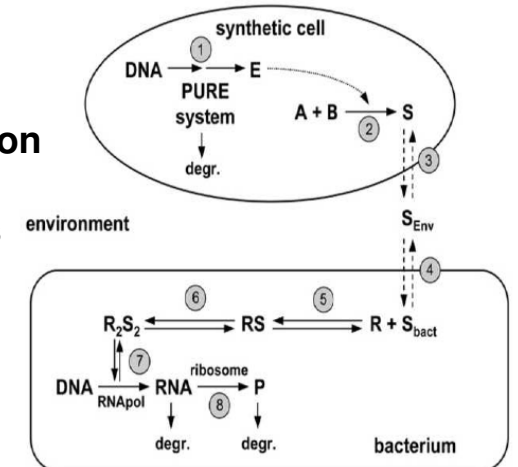
## the Ribocell model



## the Crowding effect



## Chemical communication between artificial cells and bacteria





# Acknowledgments



- Pasquale Stano
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- Peter Walde
- Pier Luigi Luisi



University of the Basque Country

colleagues

**ETH**

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Swiss Federal Institute of Technology Zurich

Marco Lerario

Pierluigi della Gatta

Angelo Lanzillotto

Gaetano Regina

Carmen Bonasia

Marika De Palo

students



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Coordination of Biological and Chemical IT Research Activities



COST ACTION  
CM0703

Action Chair Sijbren Otto  
Vice-Chair Gonzen Asikenasay  
STSM Coordinator Christoph Flamm  
Grant Holding University University of Groningen





# Conclusions

*In vitro* and *in silico* minimal artificial cell models (protocells) have been present.

By the integration between numerical model and protocells implemented in the test tube it will be possible to design and synthesize a real minimal artificial cell in a not too far future.

**Thank you for your attention**



*“All models are wrong,  
but some are useful”*

George Box



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