An Evaluation of Statistical Methods for DNA Methylation Microarray Data Analysis

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Outline



Introduction

- 2 Methylation array analysis methods
- Simulation Studies
- Real data example 4



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DNA methylation



Figure: DNAmolecule that is methylated at the two center cytosines. Source: $http: //en.wikipedia.org/wiki/DNA_methylation$

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DNA methylation and disease

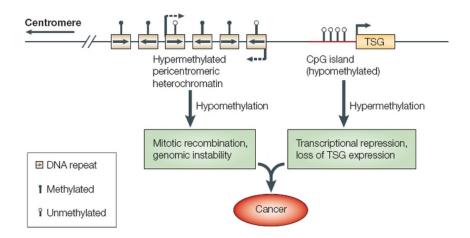


Figure: Diagram for DNA methylation and cancer. Source: 2005Nature Publishing GroupRobertson, K. DNA methylation and human disease.*Nature Reviews Genetics*6,598.

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Epigenome-wide association studies (EWAS)

- Illumina Methylation Assay
- Three platforms for DNA methylation assay
 - GoldenGate (1,563 methylation site per sample)
 - Infinium Human Methylation27 (> 27,000 methylation sites per sample)
 - Infinium HumanMethylation450 BeadChip (> 485,000 methylation sites per sample)

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Work flow of Illumina Assay

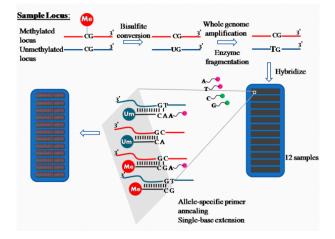


Figure: Source: http://en.wikipedia.org/wiki/Illumina_Methylation_Assay

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Methylation array downstream analysis

- Locus-by-Locus analyses are commonly used for EWAS
- Average β value denote the level or percentage of methylation for a locus
- *M* value, or log ratio of percentage of methylation, is also commonly used to measure methylation
- Relationship between the β -value and the *M*-value

$$M = \log_2 \frac{\beta}{1-\beta}$$

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Methods implemented in Bioconductor/R

- Wilcoxon rank sum test (methyAnalysis)
- t-test (methyAnalysis, CpGAssoc, RnBeads, and IMA package)
- Kolmogorov-Smirnov Tests (Used in some papers: Price et al. Epigenetics & Chromatin 2013, 6:4)
- Permutation test (CpGAssoc package)
- Empirical Bayes method (RnBeads, IMA and minfi package)
- Bump hunting method (minfi package)

Motivation for evaluation methylation data analysis methods

- Finding the most appropriate one to use for a specific data set is challenging
- Different methods have different assumptions to get validate results
- Multiple methods could provide inconsistent results for the same data set
- Exploring power and stability differences across different methods for the same data set
- Proving advice for investigators choosing appropriate method for their methylation data

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Definition of power and stability

	number not rejected	number rejected	
true null hypotheses	U	V	m_0
non-true null hypotheses	Т	S	m_1
total	m-R	R	m

Table: Possible outcomes from m hypotheses tests

$$Power = E(\frac{S}{m_1}|m > m_0),$$

$$Stability = Var(R) = Var(S + V) = Var(S) + Var(V) + 2Cov(S, V).$$

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Wilcoxon rank sum test

$$H_0$$
 : Median₁ = Median₂
 $z = rac{R - \mu_R}{\sigma_R}$

where

$$\mu_R = \frac{n_1(n_1 + n_2 + 1)}{2}$$

$$\sigma_R = \sqrt{\frac{n_1n_2(n_1 + n_2 + 1)}{12}}$$

$$R = sum \text{ of ranks for smaller sample size} (n_1)$$

$$n_1 = smalle \text{ of sample sizes}$$

$$n_2 = \text{ larger of sample sizes}$$

$$n_1 \ge 10 \text{ and } n_2 \ge 10$$

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$$H_0: \mu_1 = \mu_2 \\ t = \frac{\bar{y}_1 - \bar{y}_2}{s_{\bar{y}_1 - \bar{y}_2}}$$

where

$$s_{\bar{y}_1-\bar{y}_2} = \sqrt{\frac{s_1^2}{n_1} + \frac{s_2^2}{n_2}}$$
$$df = \frac{\left(\frac{s_{i1}^2}{n_1} + \frac{s_{i2}^2}{n_2}\right)^2}{\left(\frac{s_{i1}^2}{n_1}\right)^2 + \left(\frac{s_{i2}^2}{n_2}\right)^2}{\frac{n_1^2}{n_1 - 1} + \left(\frac{s_{i2}^2}{n_2}\right)^2}$$

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Kolmogorov-Smirnov Tests

$$H_0: F_{1,n_1}(y) = F_{2,n_2}(y)$$
$$D_{n_1,n_2} = \sup_{y} |F_{1,n_1}(y) - F_{2,n_2}(y)|$$

The null hypothesis is rejected at level α if

$$D_{n_1,n_2} > c(\alpha) \sqrt{\frac{n_1 + n_2}{n_1 n_2}}$$

where $c(\alpha) = 1.36$ for $\alpha = 0.05$

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Permutation test

$$H_0: F_{1,n_1}(y) = F_{2,n_2}(y)$$

- Compute the test statistic for the observed data set
- Permute the original data in a way that matches the null hypothesis
- Calculate the critical value of a level α test based on the upper α percentile of the reference distribution
- Obtain the raw *p*-value from the reference distribution.

Empirical Bayes method

$$H_0: eta_{gj}^* = 0$$

The moderated *t*-statistic, based on a hybrid classical/Bayes approach, is defined by: $\hat{}$

$$ilde{t}_{gj} = rac{eta_{gj}^*}{ ilde{s}_g \sqrt{v_{gj}}}$$

The posterior mean of σ_g^2 given s_g^2 is

$$\tilde{s}_{g}^{2} = E(\sigma_{g}^{2}|s_{g}^{2}) = rac{d_{0}s_{0}^{2} + d_{g}s_{g}^{2}}{d_{0} + d_{g}}$$

The prior estimator s_0^2 and d_0 degrees of freedom is estimated from the data by equating empirical to expected values for the first two moments of $logs_g^2$

Bump hunting method

$$H_0:\beta^*(t_j)=0$$

Fit a linear model between methylation and disease type, covariates, and potential confounding variables

$$Y_{ij} = \mu(t_j) + \beta^*(t_j)X_i + \sum_{k=1}^p \gamma_k(t_j)Z_{i,k} + \sum_{l=1}^q \mathsf{a}_{l,j}W_{i,l} + \epsilon_{i,j}$$

where i is ith subject and j is j-th genomic locus

- Estimate $\beta(t_j)$ for each t_j
- **2** Use these to estimate the smooth function $\beta(t)$
- **③** Use this to estimate the regions R_n , n = 1, ..., N for which $\beta(t) \neq 0$ for all $t \in R_n$
- Use permutation tests to assign statistical uncertainty to each estimated region

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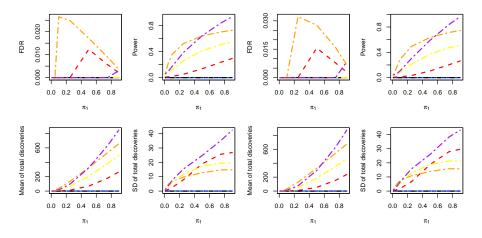
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Simulation Set up

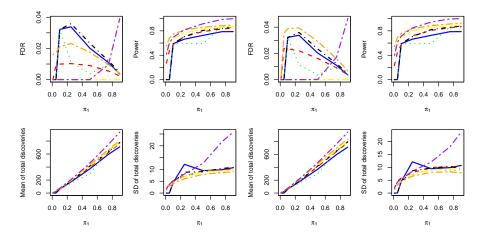
- Methylation data are generated from mixed beta distributions to mimic real methylation data
- Proportion of methylated loci are 0.01, 0.05, 0.10, 0.25, 0.50, 0.75, 0.90 to cover all possible scenarios
- Mean β value differences were set to be between 0.1 and 0.4 with steps equaling $1/(m m_0)$ such as 1/10, 1/50, 1/100, 1/250, 1/500, 1/750, 1/900
- 1000 loci and 1000 independent simulations
- Sample sizes are 3, 6, 12, and 24 in each group with two-group comparisons
- Both β values and M values are compared

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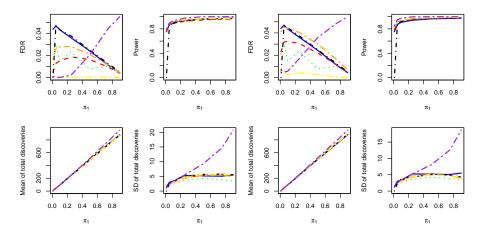
Methylation Results for sample size 3 in each group (Left: β values and Right: *M* values)



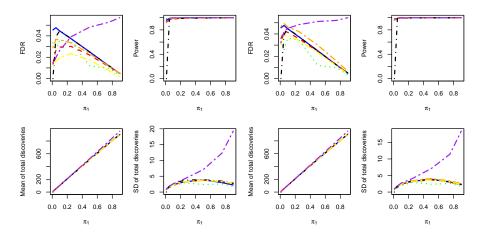
Methylation Results for sample size 6 in each group (Left: β values and Right: *M* values)



Methylation Results for sample size 12 in each group (Left: β values and Right: *M* values)



Methylation Results for sample size 24 in each group (Left: β values and Right: *M* values)



Real data example

- Genome wide DNA methylation profiling of United Kingdom Ovarian Cancer Population Study (UKOPS) with GEO accession number GSE19711
- Illumina Infinium 27k Human DNA methylation Beadchip v1.2 with 27578 CpGs in whole blood sample from 3, 6, or 12 cases and 3, 6, or 12 controls
- Total number of rejections at 10 significance levels were recorded using raw *p*-values
- Both β values and M values are compared
- Raws *p*-values are used for comparisons and no rejections for Bump Hunting method using Storey's *q*-value adjustment

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Apparent test power comparisons for n = 3 in each group

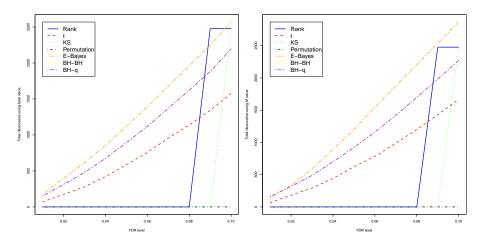


Figure: Blue: rank test; Red: *t*-test; Green: KS test; Black: permutation test; Orange: Empirical Bayes; Yellow: Bump Hunting BH.

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Apparent test power comparisons for n = 6 in each group

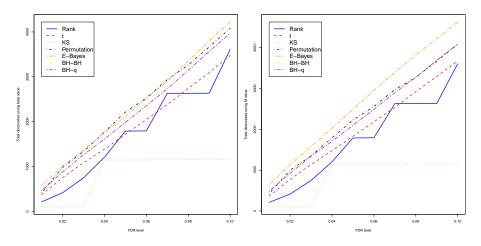


Figure: Blue: rank test; Red: *t*-test; Green: KS test; Black: permutation test; Orange: Empirical Bayes; Yellow: Bump Hunting BH.

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Apparent test power comparisons for n = 12 in each group

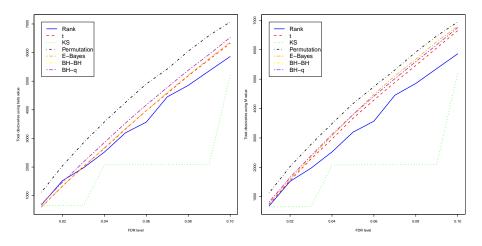


Figure: Blue: rank test; Red: *t*-test; Green: KS test; Black: permutation test; Orange: Empirical Bayes; Yellow: Bump Hunting BH.

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Discussion

- No significant differences were detected in terms of FDR control, power, and stability between β values and M values
- For small sample size, both empirical Bayes method and bump hunting method showed good FDR control and much larger power than all other methods compared
- For medium to large sample size, all methods compared have good FDR control except the bump hunting method with large proportion of differentially methylated loci
- For medium to large sample size, all methods compared have almost equivalent power except permutation test with very low proportion of differentially methylated loci
- For all sample sizes, bump hunting method has lowest stability in terms of variance of total discoveries

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Conclusion

- Either β values or M values are good to use for methylation data analysis
- Empirical Bayes method is recommended for methylation studies with small sample size
- For medium to large sample size, all methods except the bump hunting method are good for differentially methylation data analysis

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Acknowledgement

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