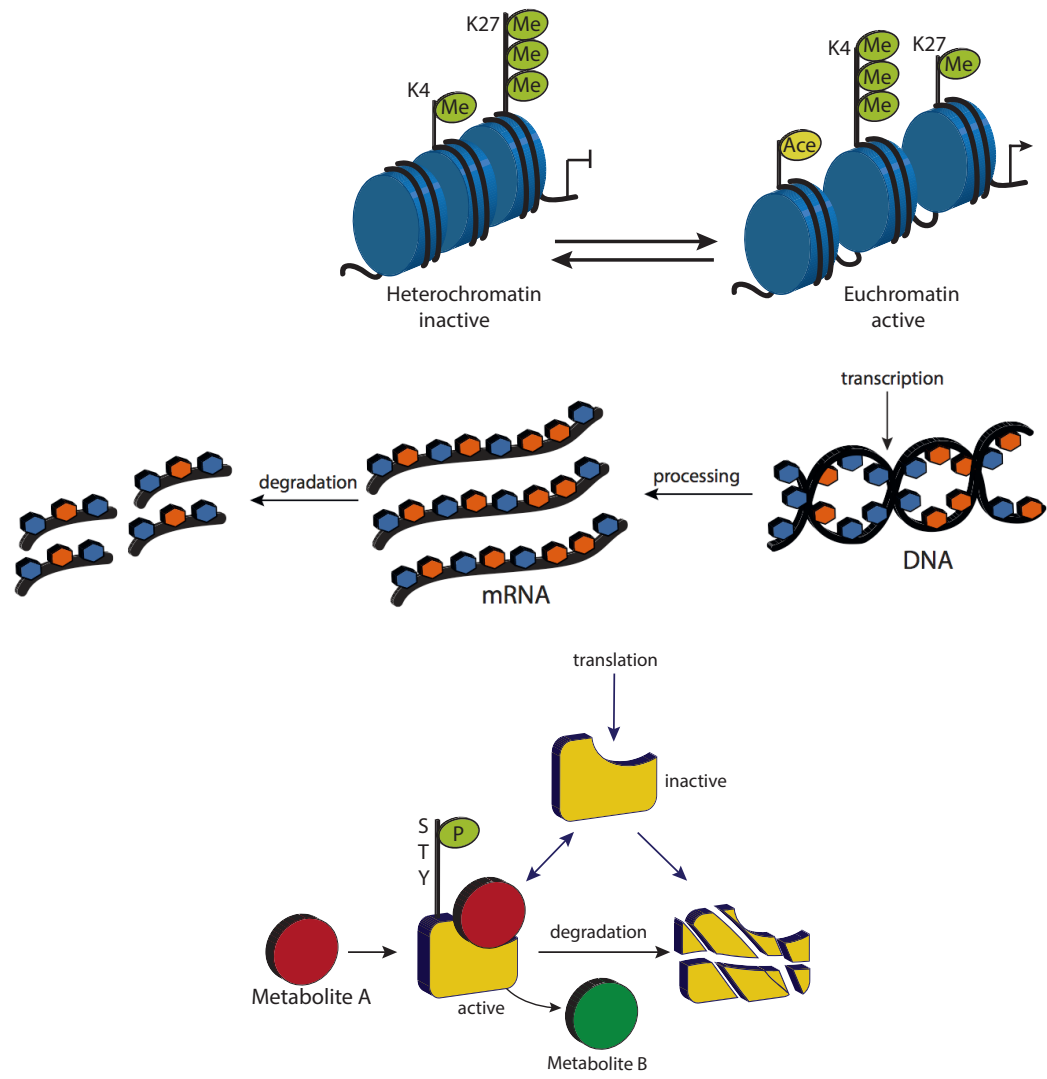


QUANTIFYING METABOLIC FLUXES IN CANCER AND STEM CELLS

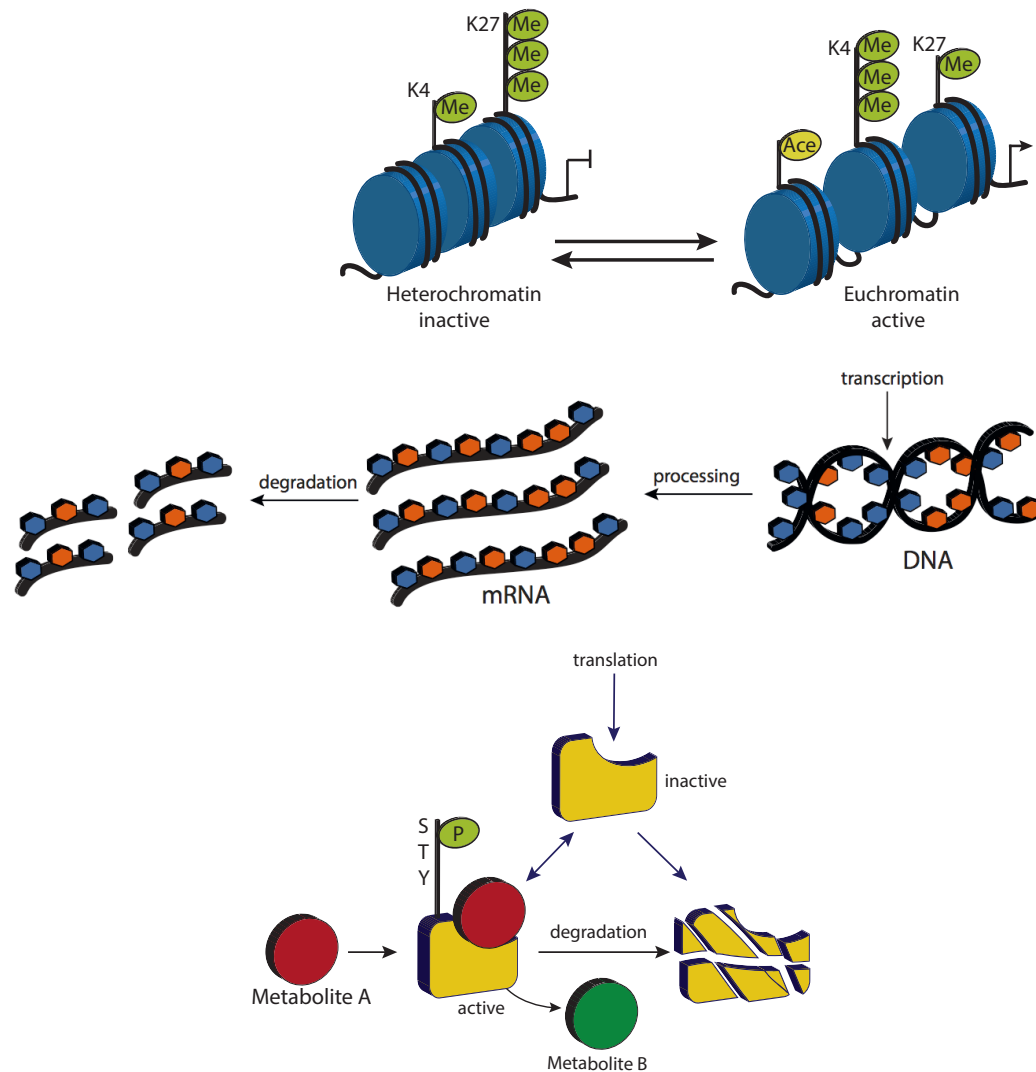
An integrative approach monitoring metabolic reprogramming

Christin Zasada
Kempa Lab - Integrative Proteomics & Metabolomics

Genome-regulatory code



Genome-regulatory code



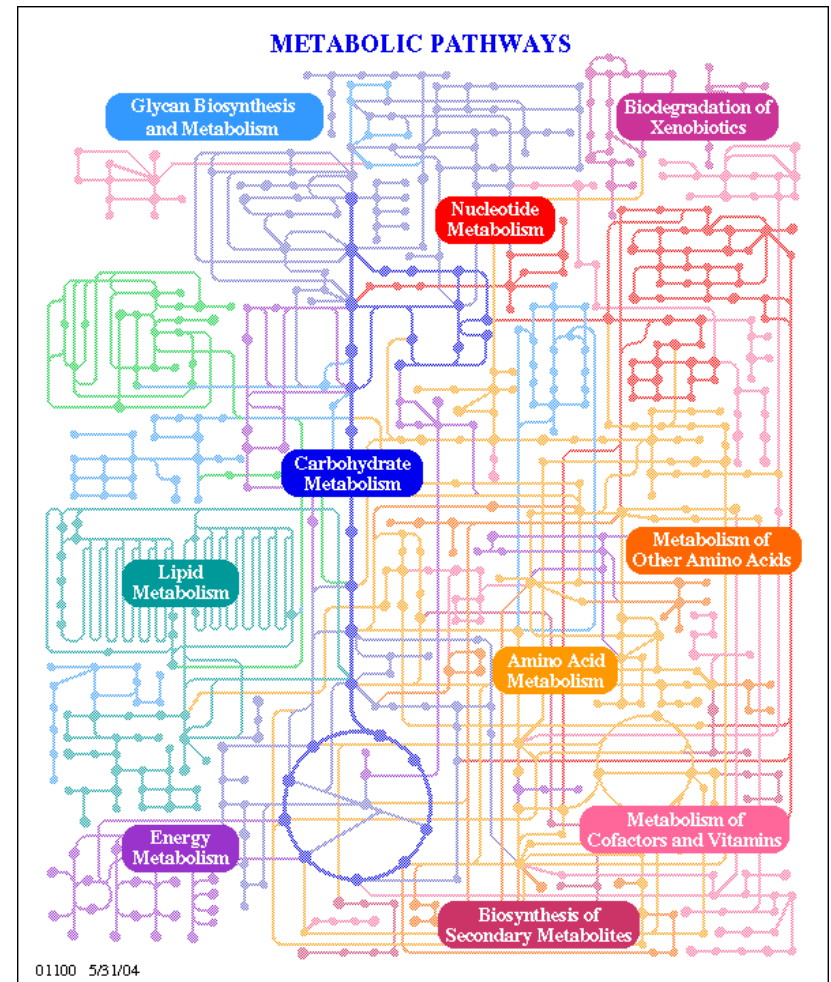
Genomics
three billion DNA base pairs

Transcriptomics
2 % of the genome

Proteomics
20'000 - 25'000 protein-coding genes

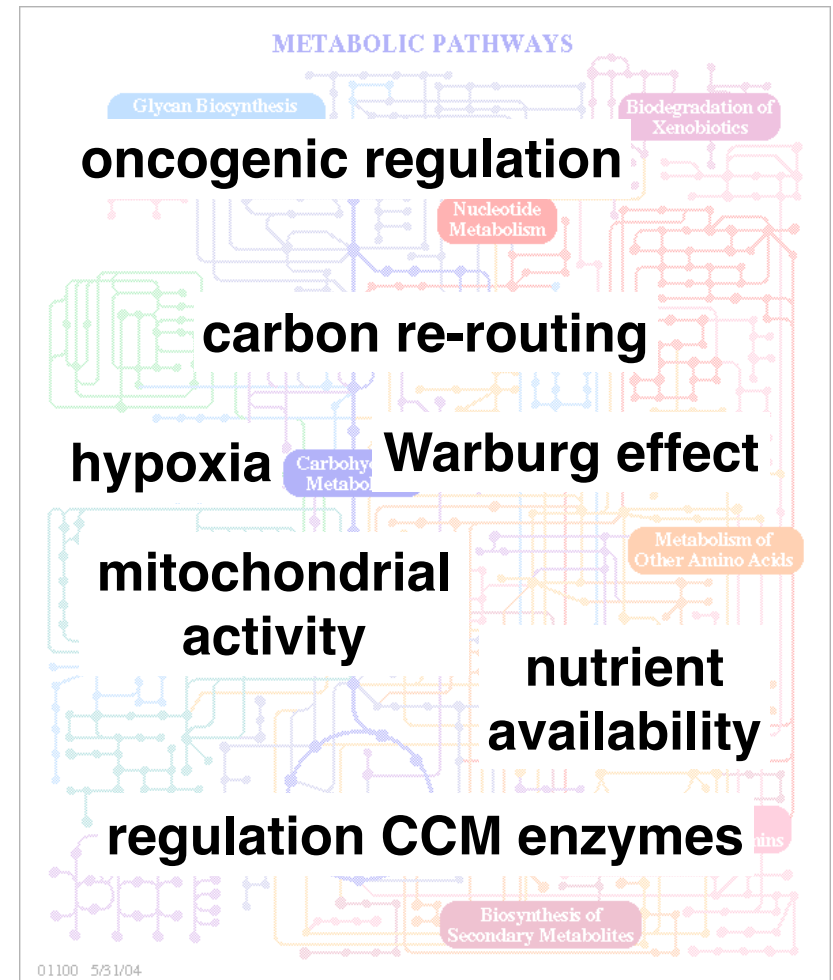
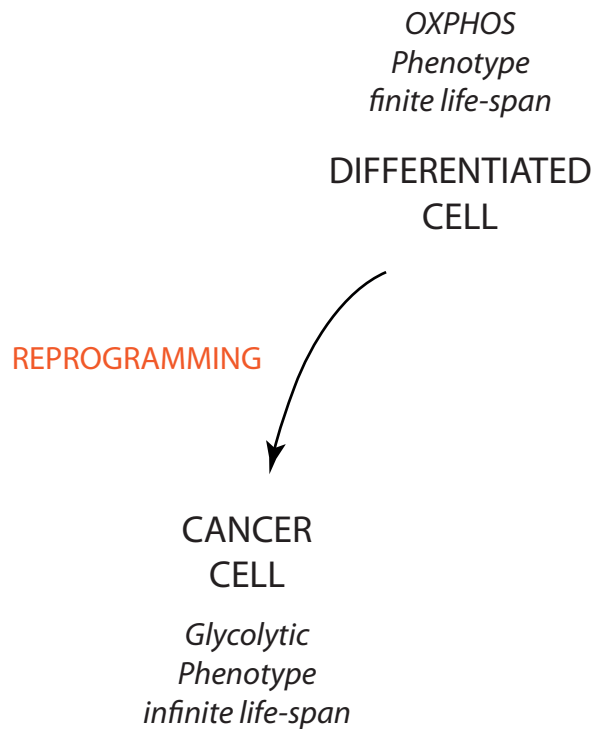
Metabolomics
~ 5 000

Reprogramming of metabolism



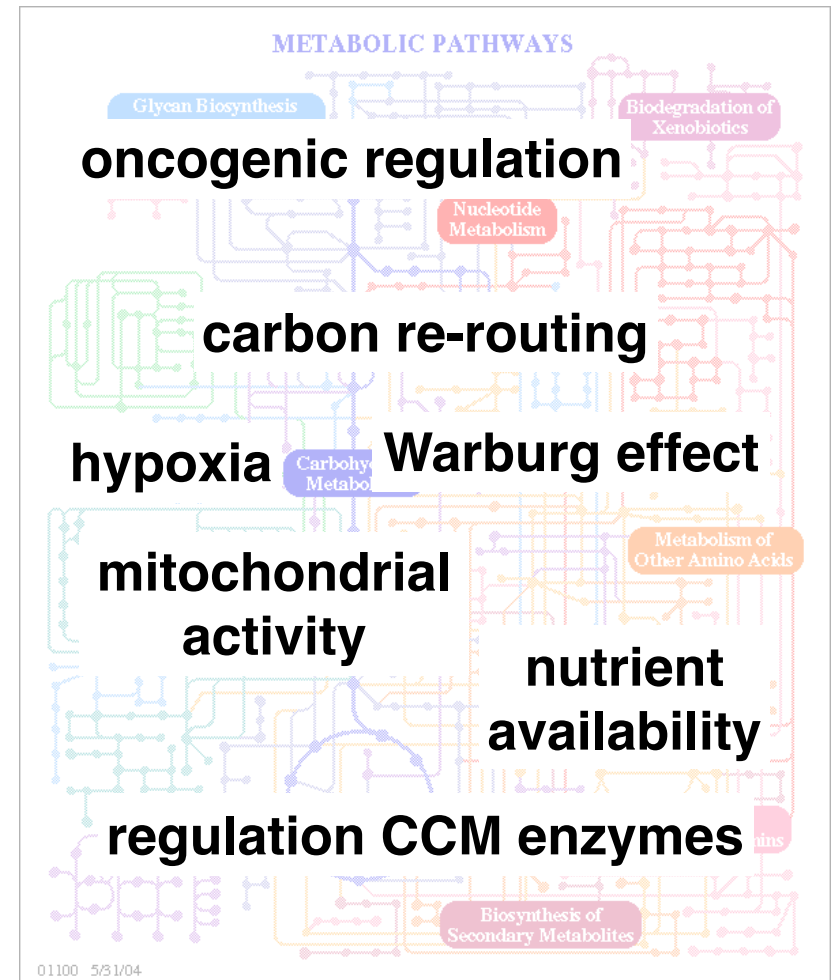
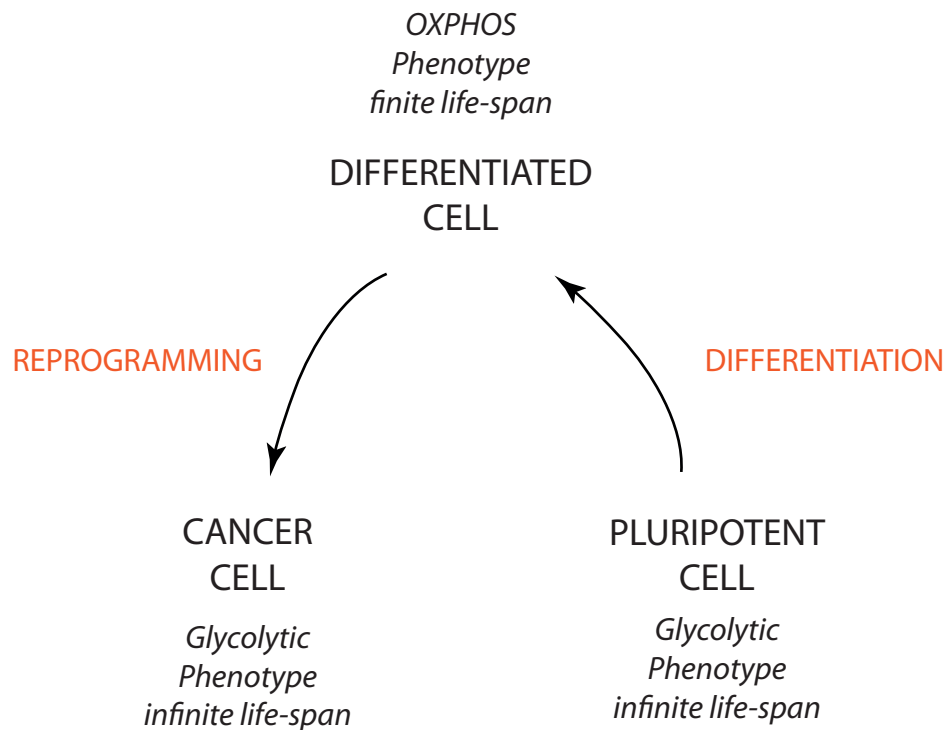
www.kegg.org

Reprogramming of metabolism



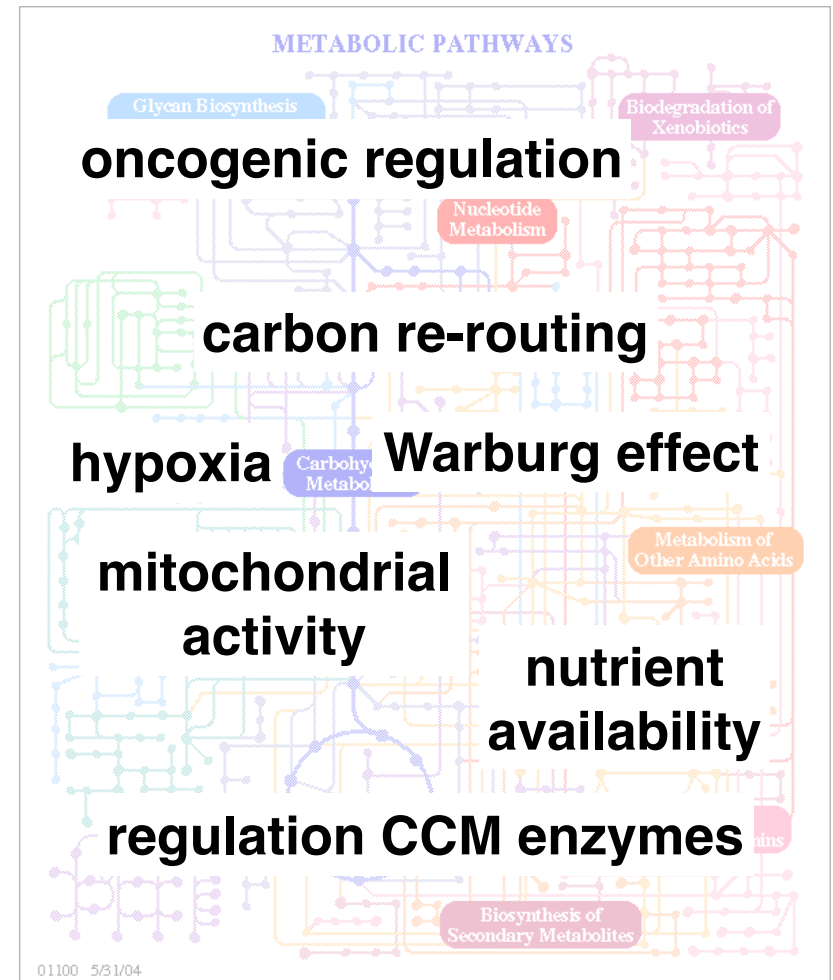
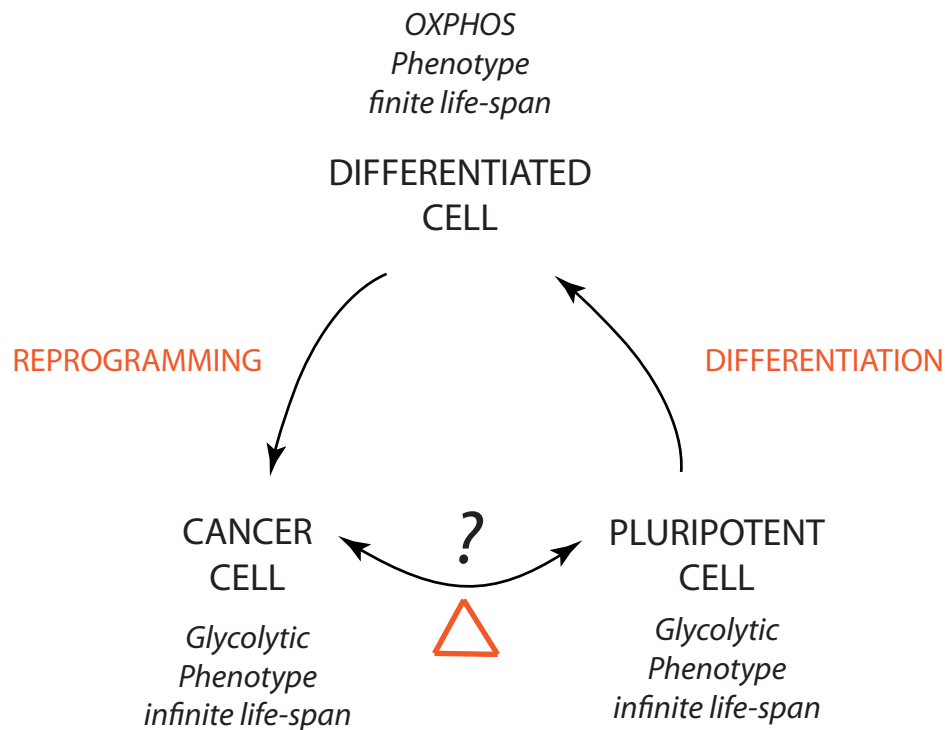
www.kegg.org

Reprogramming of metabolism



www.kegg.org

Reprogramming of metabolism



www.kegg.org

Quantitative information turns functional

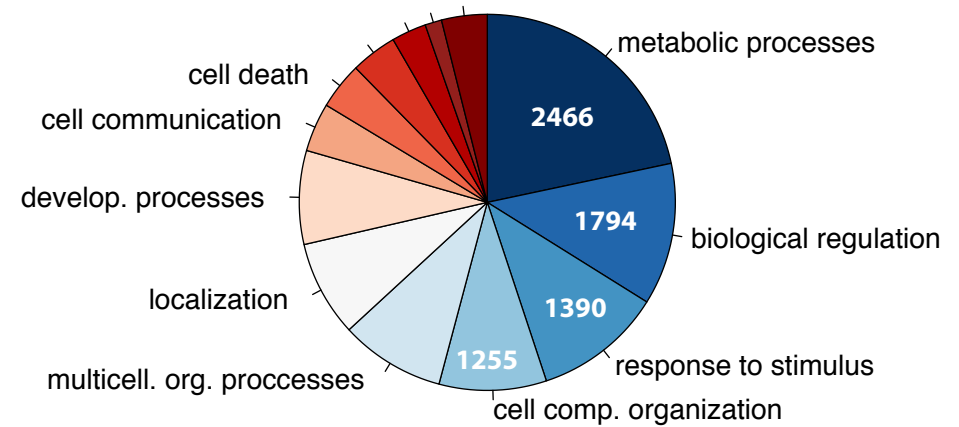
Quantitative information turns functional

Proteomics

LC-MS/MS

label-free, SILAC quantification

~ 3500 proteins



Quantitative information turns functional

Proteomics

LC-MS/MS

label-free, SILAC quantification

~ 3500 proteins

pulsed stable isotope resolved metabolomics (pSIRM)

GC-MS approach

identification

~ 100 metabolites

absolute quantification

~ 40 metabolites

Quantitative information turns functional

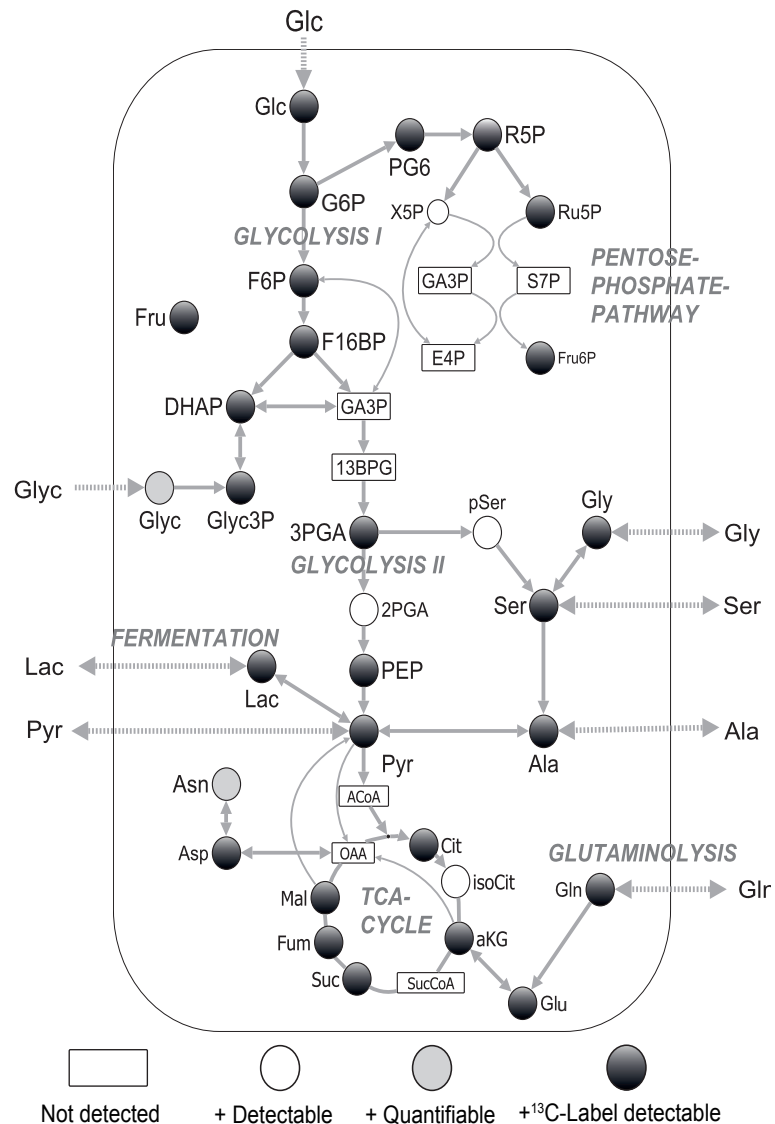
Proteomics

LC-MS/MS
label-free, SILAC quantification
~ 3500 proteins

pulsed stable isotope resolved metabolomics (pSIRM)

GC-MS approach
identification
~ 100 metabolites
absolute quantification
~ 40 metabolites

application of stable isotopes
—> time-resolved
—> position-specific
—> indirect measurement of flux



Pietzke & Zasada et al. Cancer & Metabolism 2014, 2:9

Non-stationary metabolic flux analysis

in cooperation with Katharina Nöh
and Sebastian Niedenführ
Forschungszentrum Jülich GmbH

**metabolic steady &
isotopic non-stationary**

Time courses for:

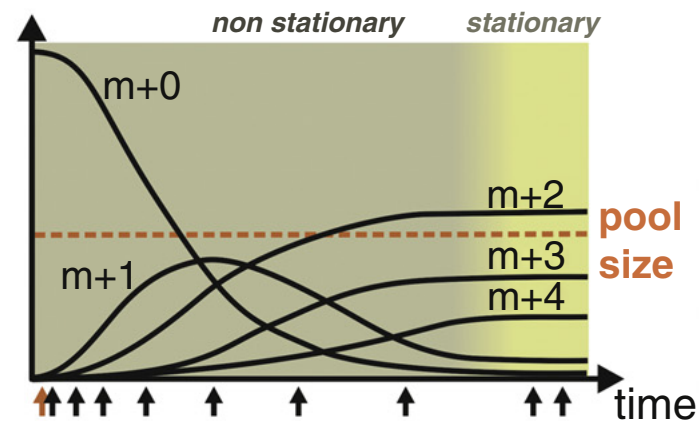
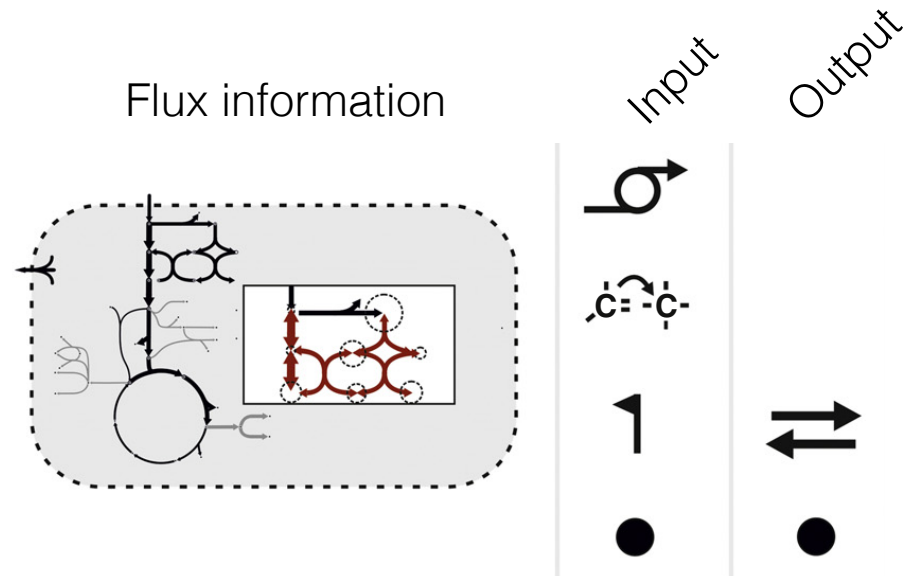
Glucose
Glutamine

Model parameter

168 labeling data point
5 measured rates
14 measured pool sizes

Pathways coverage

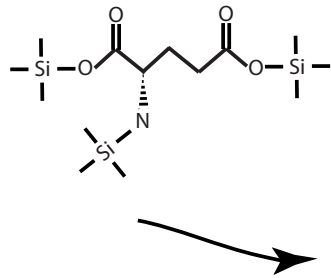
Glycolysis
TCA-Cycle
Pentose-Phosphate Pathway
Glutaminolysis
Amino acid synthesis
Biosynthesis



Niedenführ S, Nöh K, Wiechert W
Current Opinion in Biotechnology 2015, 34:82–90

GC-MS fragment mapping to molecule level

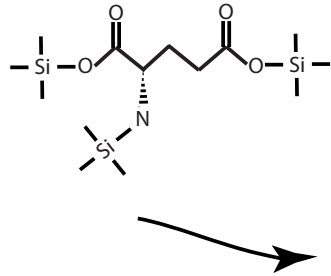
Glutamine DL (3 TMS)



modified from
S. Niedenführ

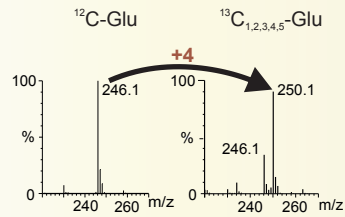
GC-MS fragment mapping to molecule level

Glutamine DL (3 TMS)

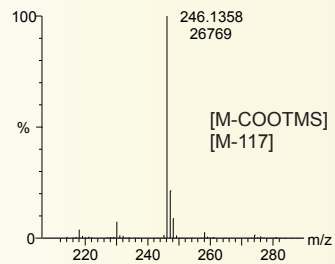


Fragment identification

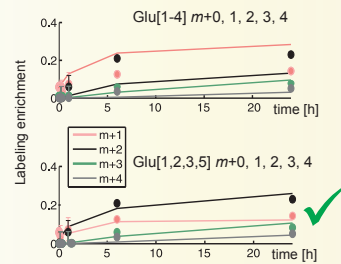
I. Isotope standards



II. High-resolutions mass spectrometry



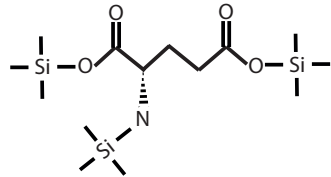
III. *in silico* consistency check



modified from
S. Niedenführ

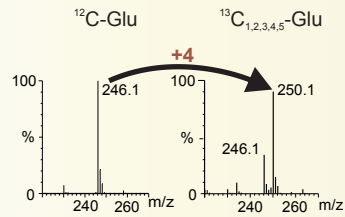
GC-MS fragment mapping to molecule level

Glutamine DL (3 TMS)

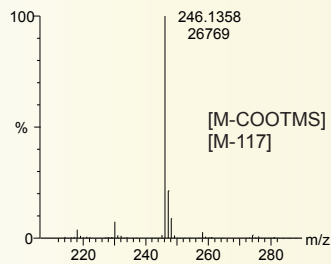


Fragment identification

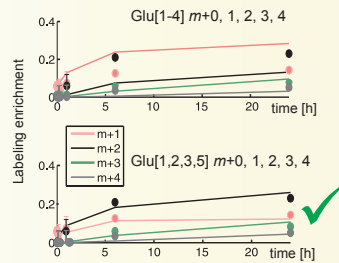
I. Isotope standards



II. High-resolutions mass spectrometry



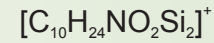
III. in silico consistency check



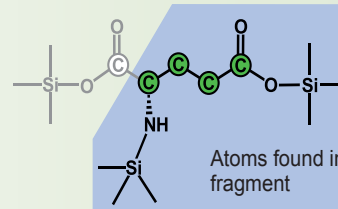
modified from
S. Niedenfür

Identified fragment

I. Chemical formula

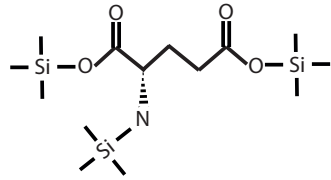


II. Fragment and precursor mapping



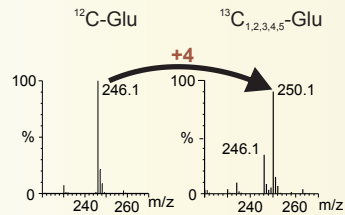
GC-MS fragment mapping to molecule level

Glutamine DL (3 TMS)

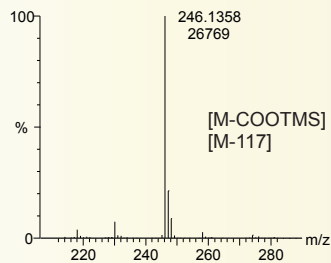


Fragment identification

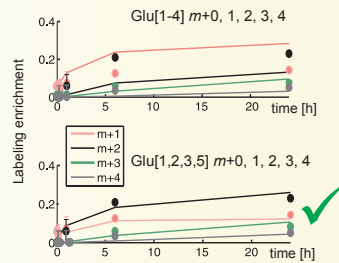
I. Isotope standards



II. High-resolutions mass spectrometry



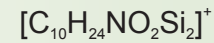
III. *in silico* consistency check



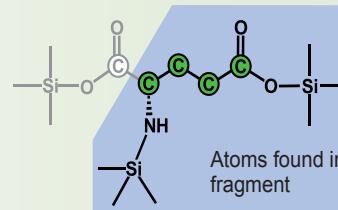
modified from
S. Niedenführ

Identified fragment

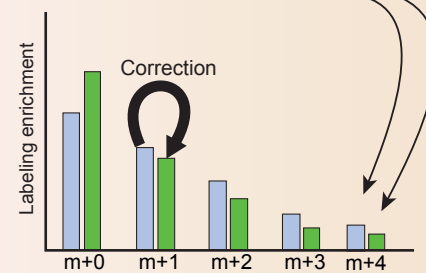
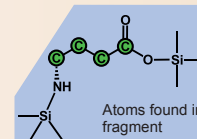
I. Chemical formula



II. Fragment and precursor mapping

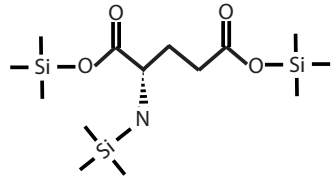


Correction for natural abundance



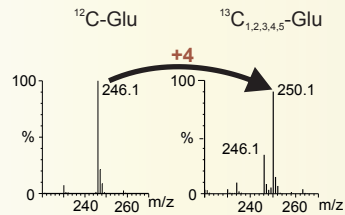
GC-MS fragment mapping to molecule level

Glutamine DL (3 TMS)

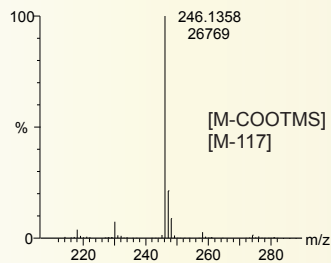


Fragment identification

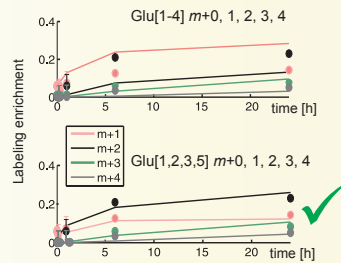
I. Isotope standards



II. High-resolutions mass spectrometry



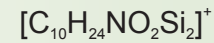
III. *in silico* consistency check



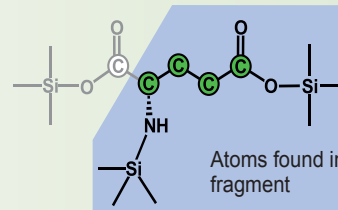
modified from
S. Niedenführ

Identified fragment

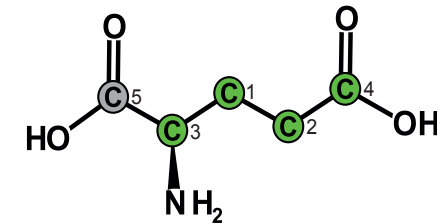
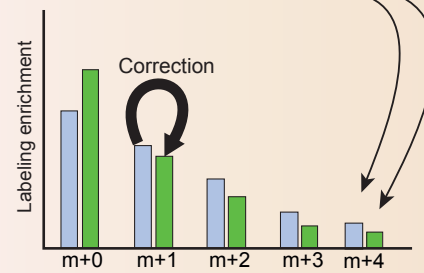
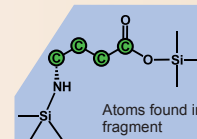
I. Chemical formula



II. Fragment and precursor mapping

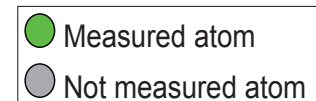


Correction for natural abundance



Measurement specification

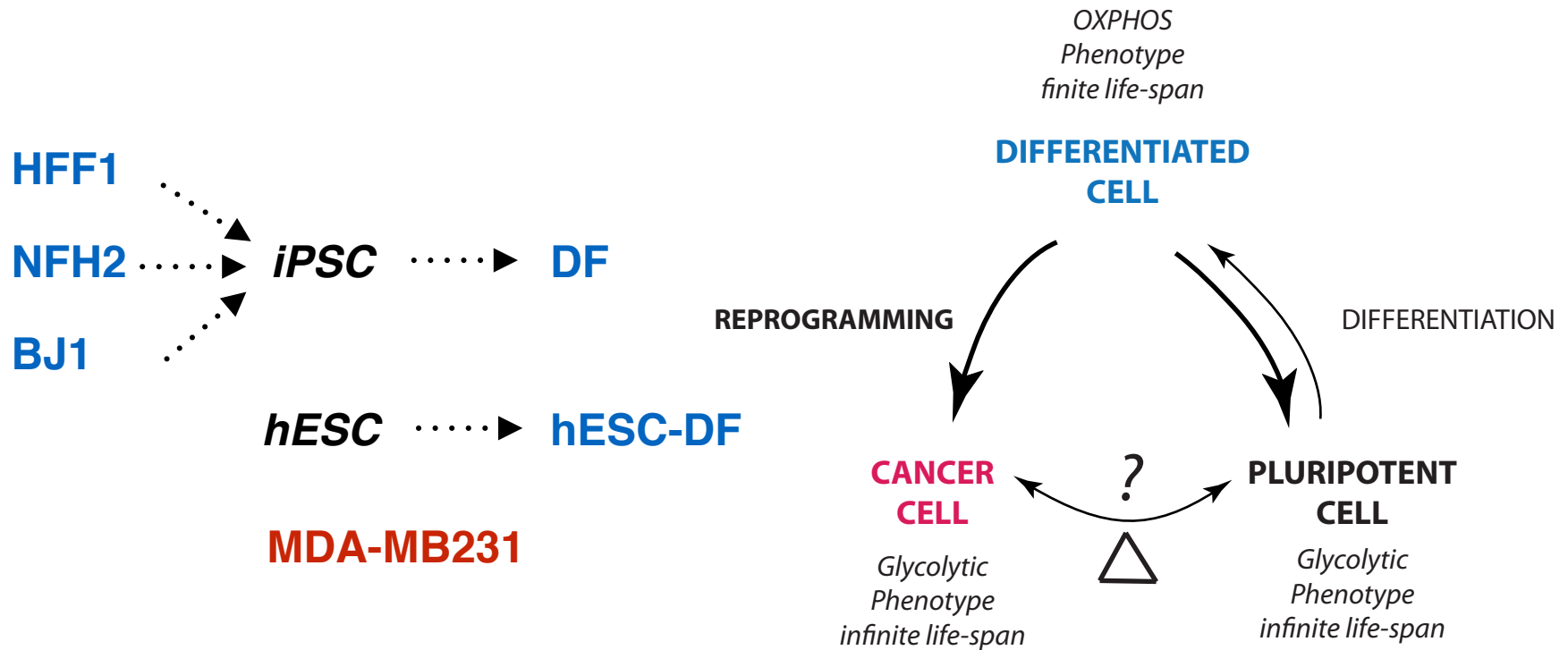
Glu[1-4] m+0, 1, 2, 3, 4



Monitoring metabolic reprogramming - experimental setup

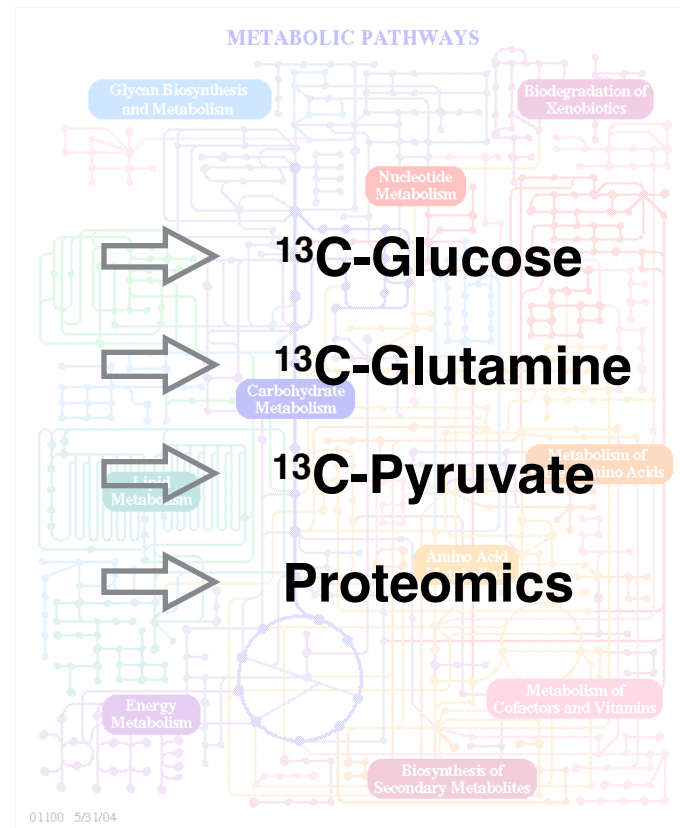
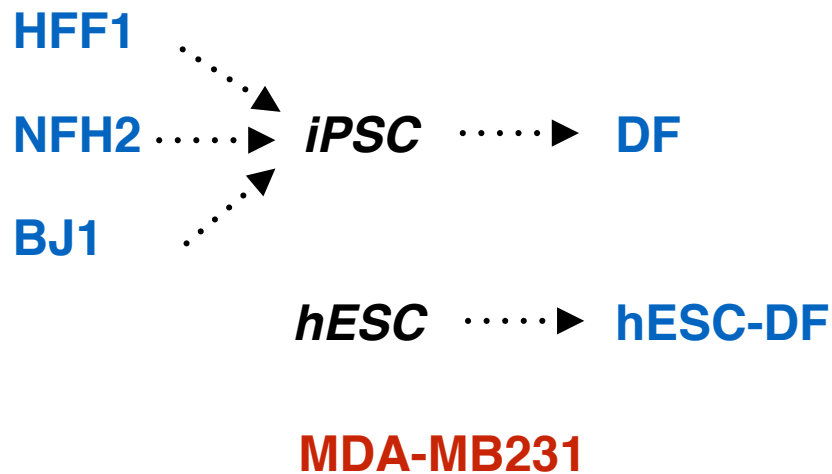
in cooperation with R. Bucowiecki and A. Prigione (MDC, Berlin, Germany)

Monitoring metabolic reprogramming - experimental setup



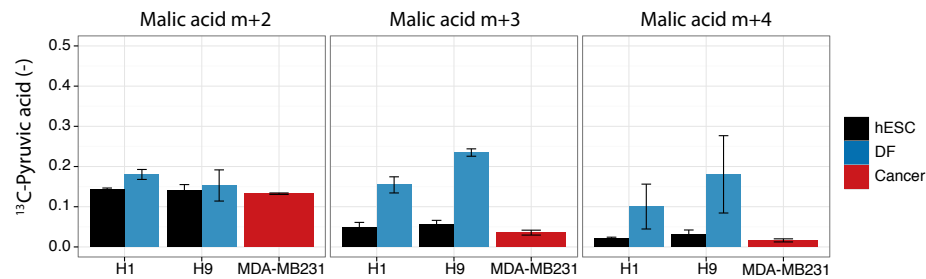
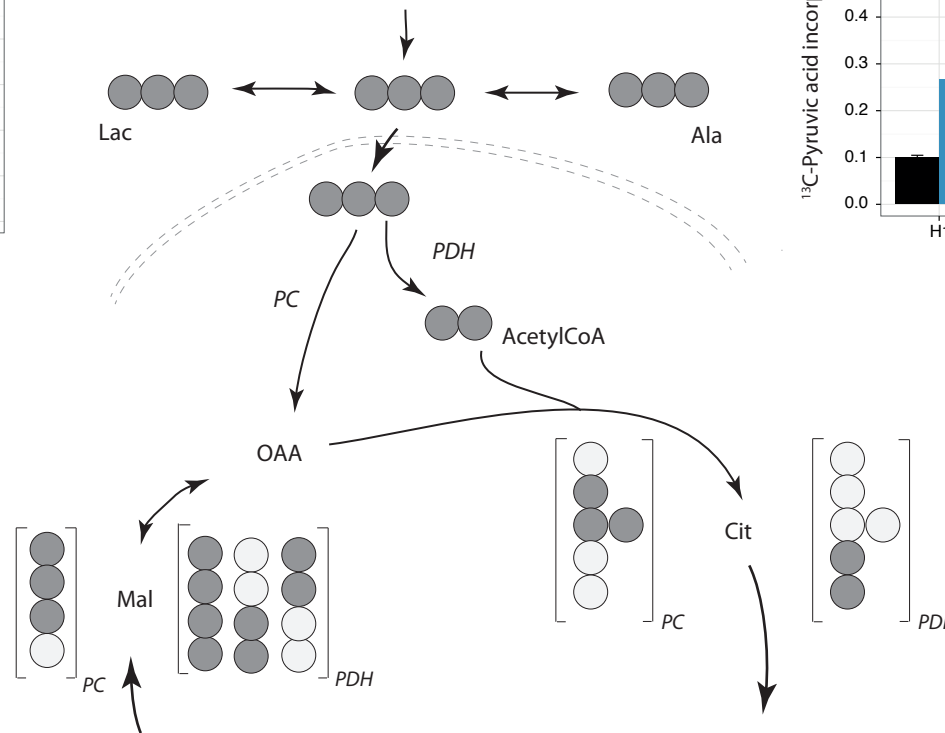
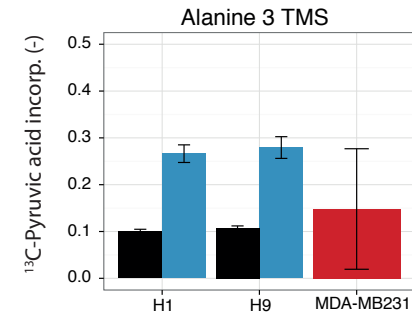
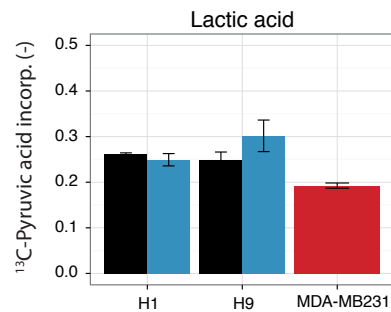
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Monitoring metabolic reprogramming - experimental setup



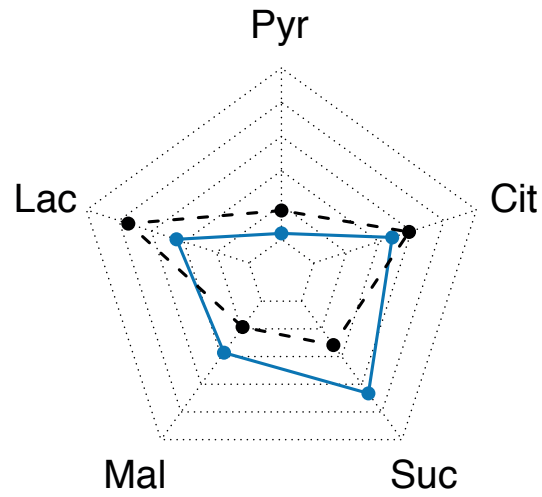
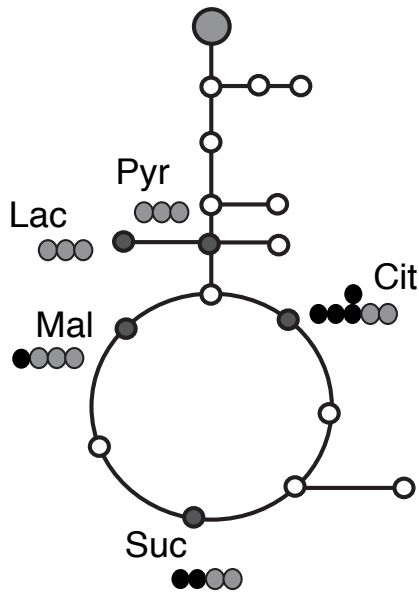
in cooperation with R. Bucowiecki and A. Prigione (MDC, Berlin, Germany)

Routing of ^{13}C -Pyruvic acid

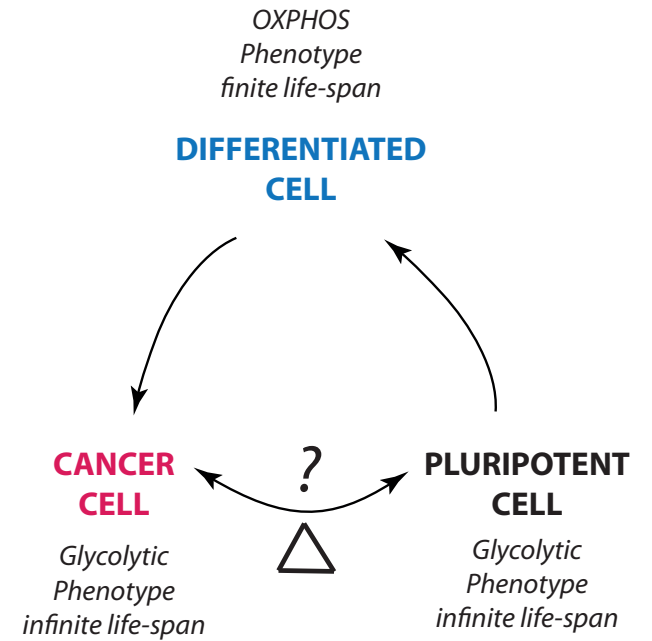
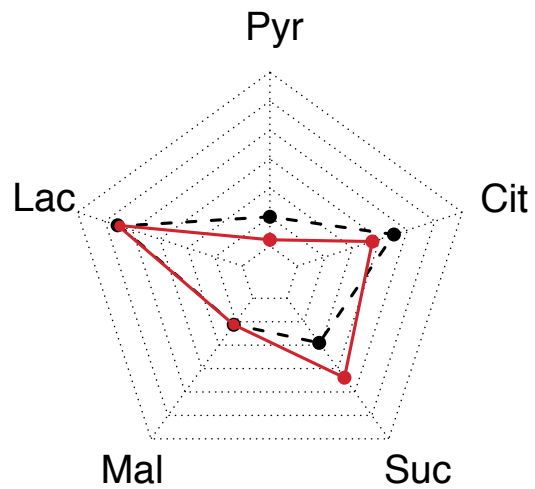


Summary

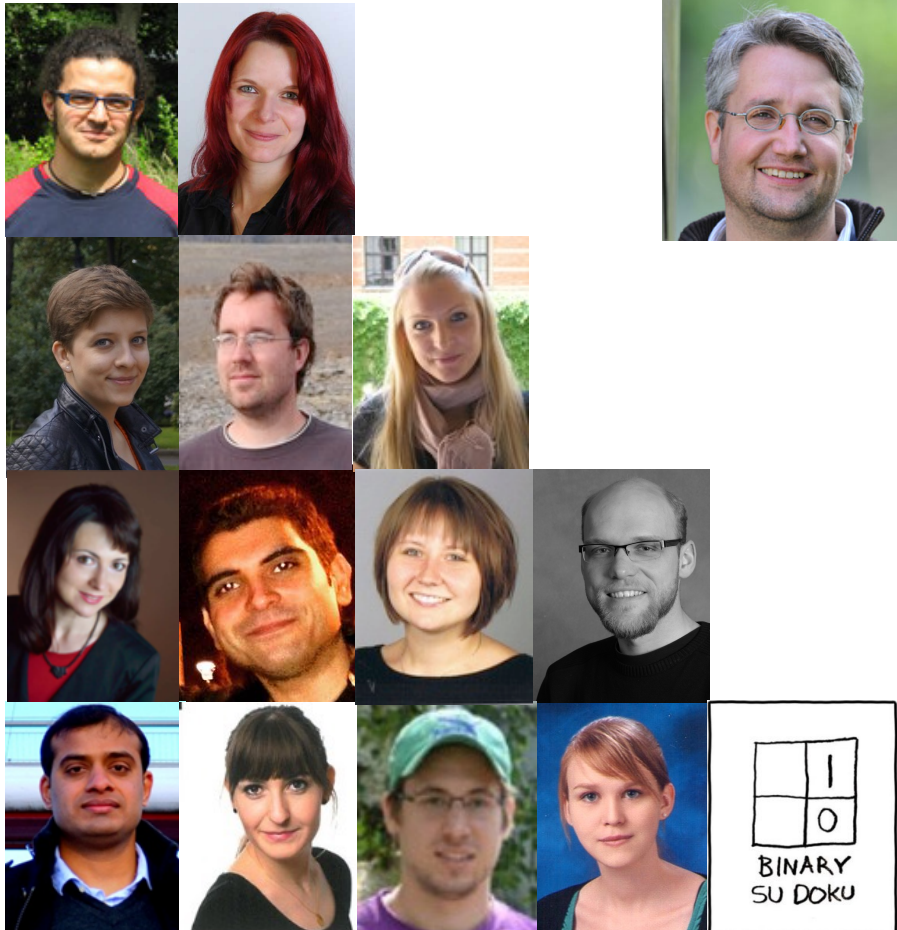
pluripotent vs. differentiated



pluripotent vs. tumorigenic



Acknowledgments



Alessandro Prigione
Raul Bukowiecki

MDC Berlin-Buch, Germany

Katharina Nöh
Sebastian Niedenführ

Forschungszentrum Jülich
Germany

... thank you for your
attention