

# Improved testing for the pathogenic fungi *Aspergillus fumigatus*

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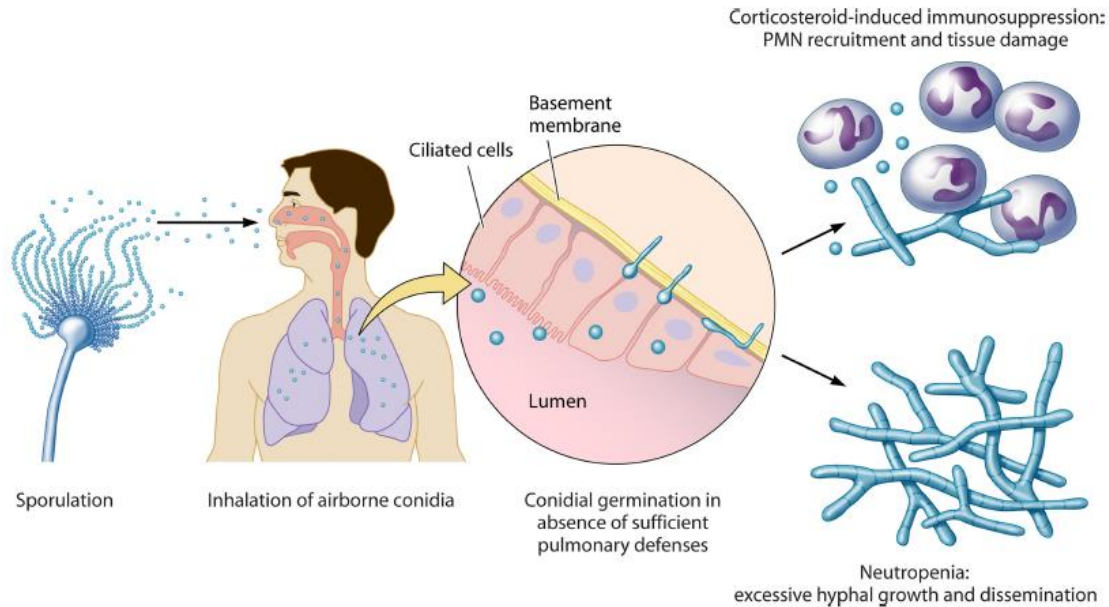
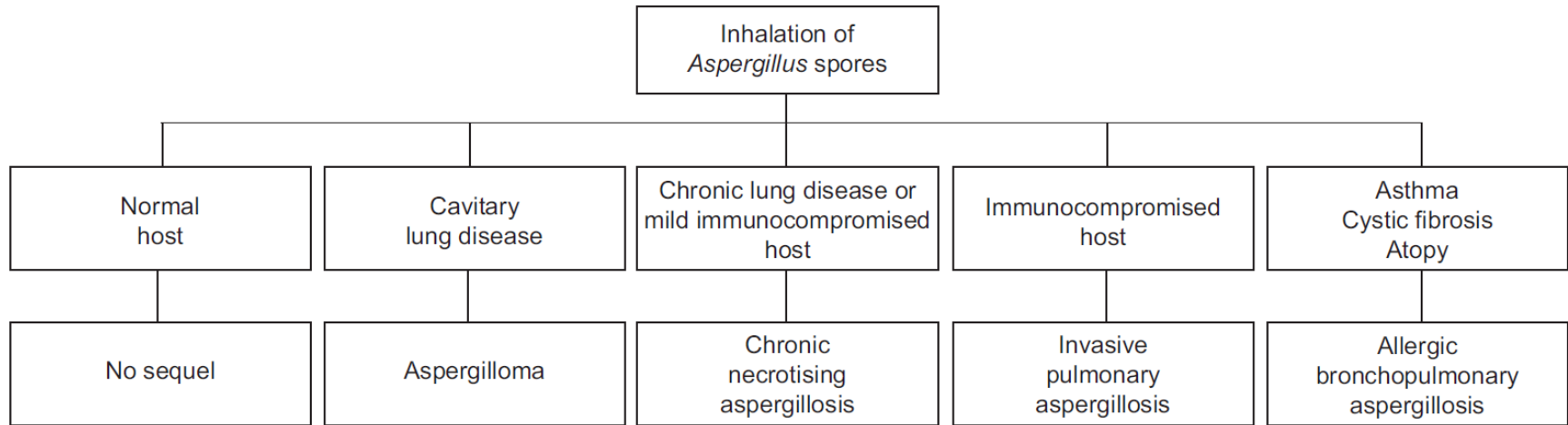
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# Aims:

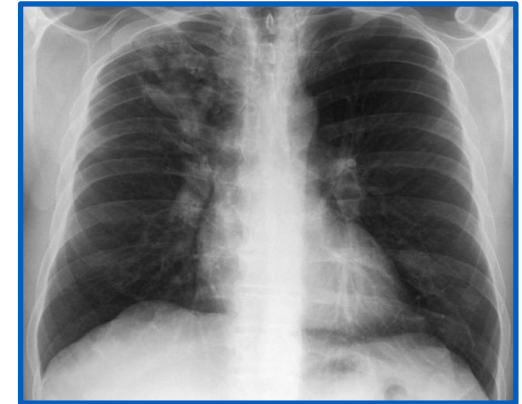
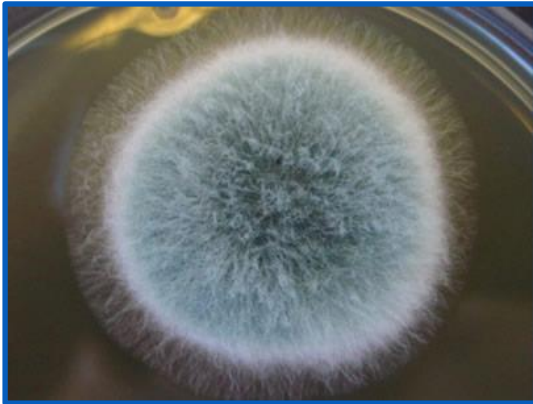
- What is *Aspergillus fumigatus*?
- Why is it a problem for the waste industry?
- Current environmental monitoring methods
- New and improved testing: RNA extraction
- New and improved testing: RT-qPCR assay
- What next?



# Aspergillus fumigatus: Leading cause of invasive aspergillosis



# Implications for the waste industry



## Case 1

- **Coughing** with expectoration of rubbery brown casts and blood from the lower respiratory tract

carrying sarcoidosis

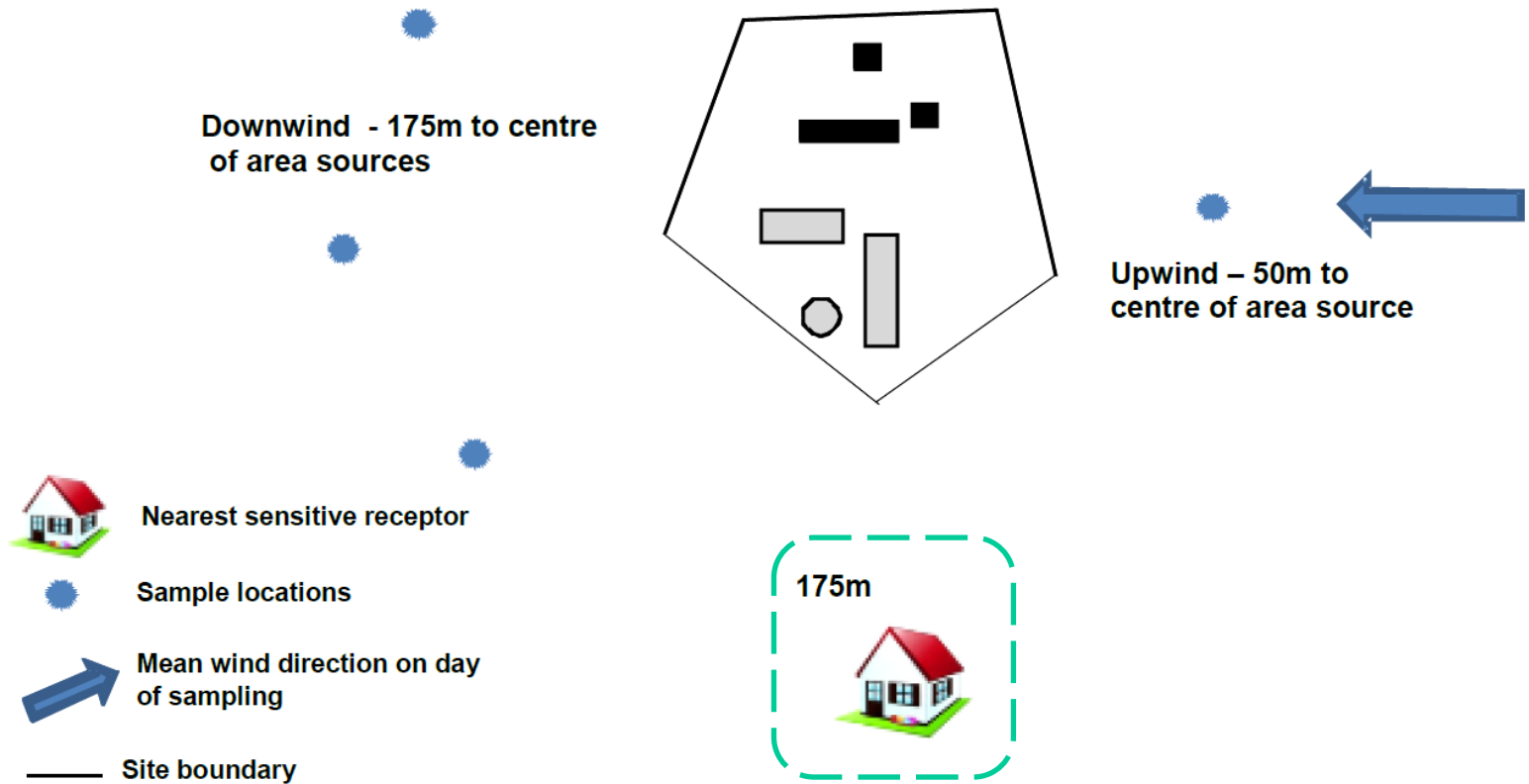
## Case 2

Existing allergic rhinitis and asthma

**ADVISED NOT TO WORK WITH COMPOST**



# Implications for the waste industry



**Sensitive receptor: nearest place to the permitted activities where people are likely to be for prolonged periods (250 m).**



# Current bioaerosol monitoring methods



Impaction



Filtration

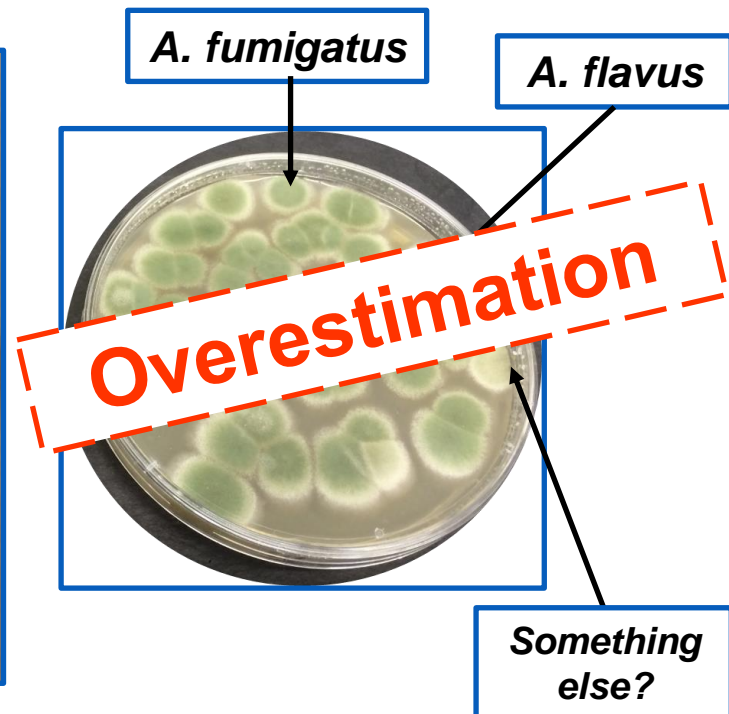
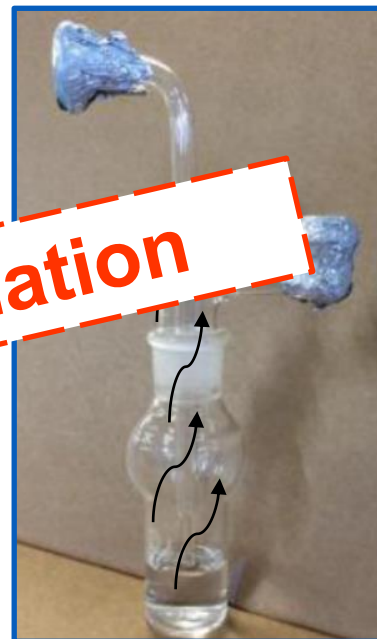


Impingement



# Current bioaerosol monitoring methods

- Only 10% of microorganisms are culturable
- Stressed microorganisms may not grow on SELECTIVE media
- VBNC microorganisms won't grow at all
- Current sampling techniques come with their own stressors



# Why select for *A. fumigatus*?



*A. fumigatus* causes **90%**  
of all Aspergillosis cases

Environment agency stipulate AF in permits

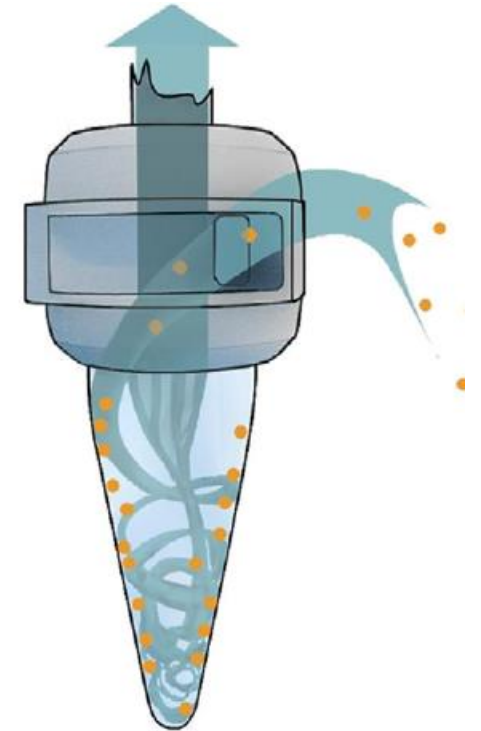
Correct antimicrobial treatment depends on correct diagnosis

Single assay for both purposes





# New and improved testing



# Quantitative PCR

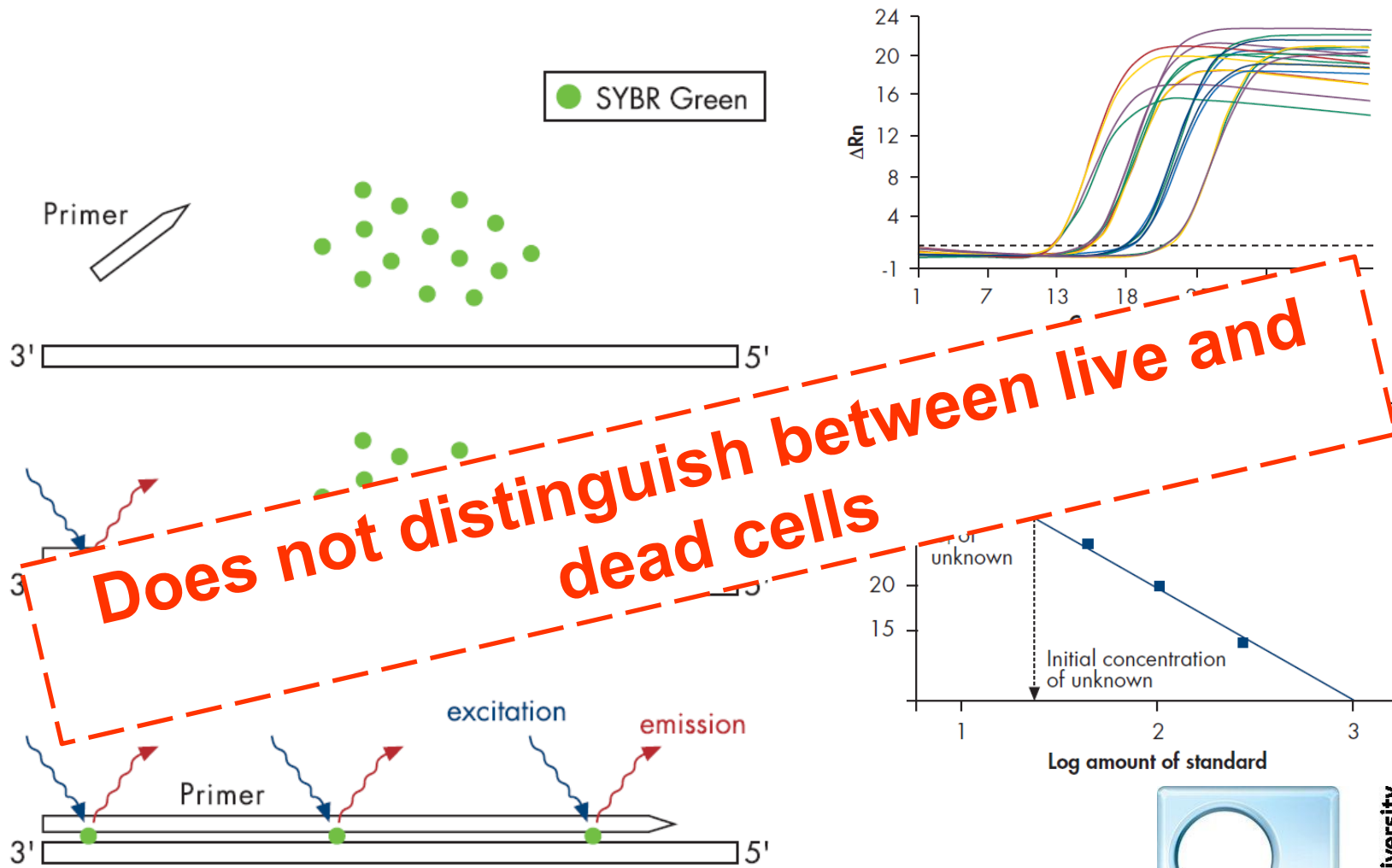


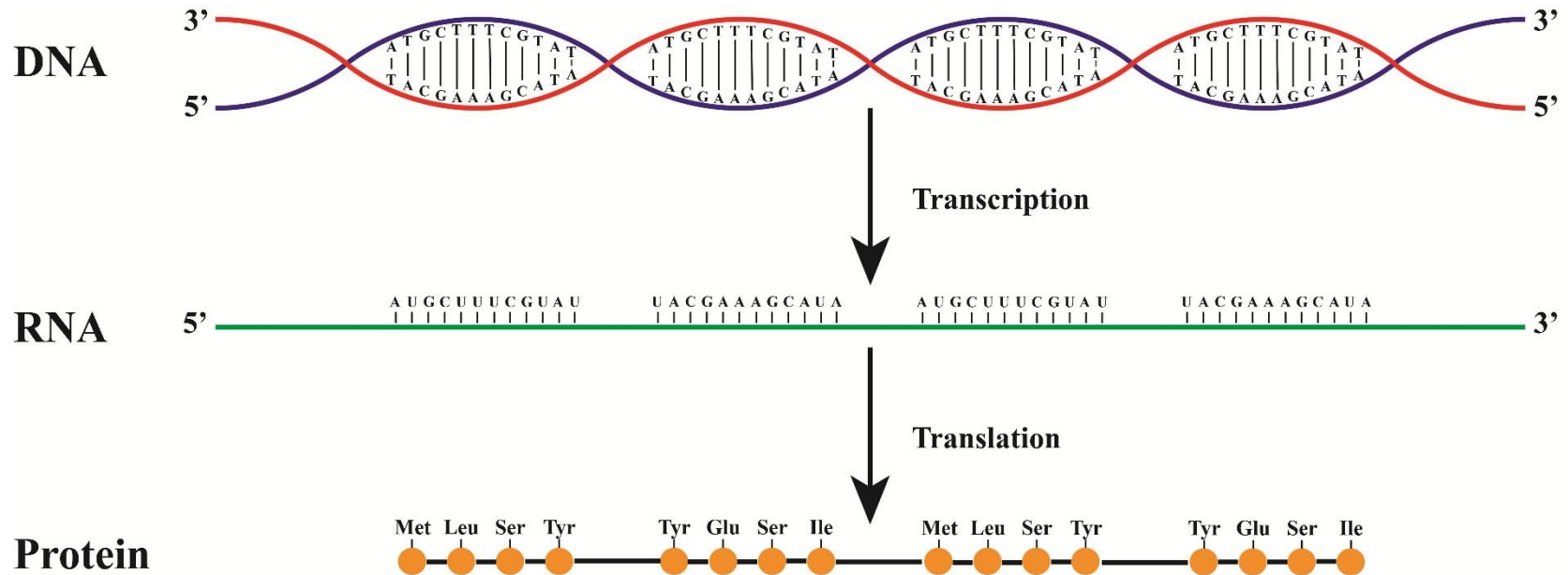
Figure 1. SYBR Green principle. Principle of SYBR Green-based detection of PCR products in real-time PCR.

Ref: Qiagen, 2010.



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# Reverse transcriptase-qPCR



# New and improved testing

Two objectives:

- Extract RNA from bioaerosols
- Design a reverse transcriptase – quantitative PCR assay specific for *A. fumigatus*



# RNA extraction from pure culture

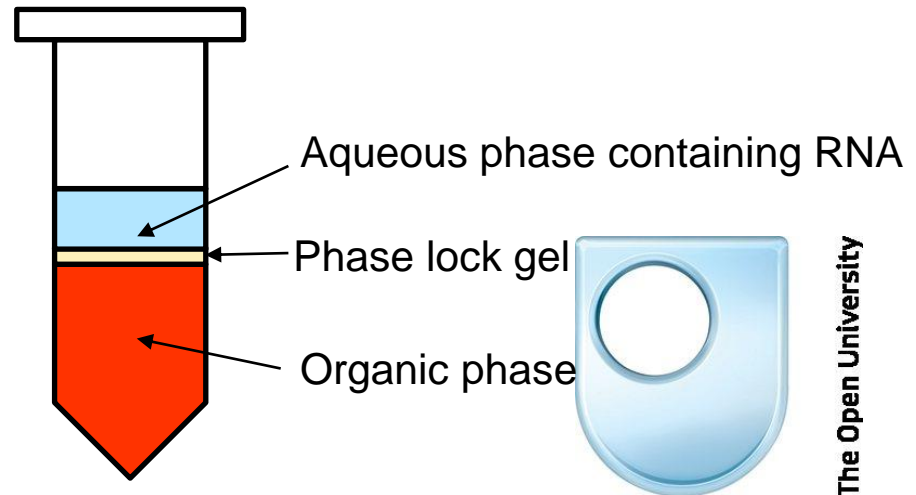
- Freeze +

**Unable to break the conidial cell wall**

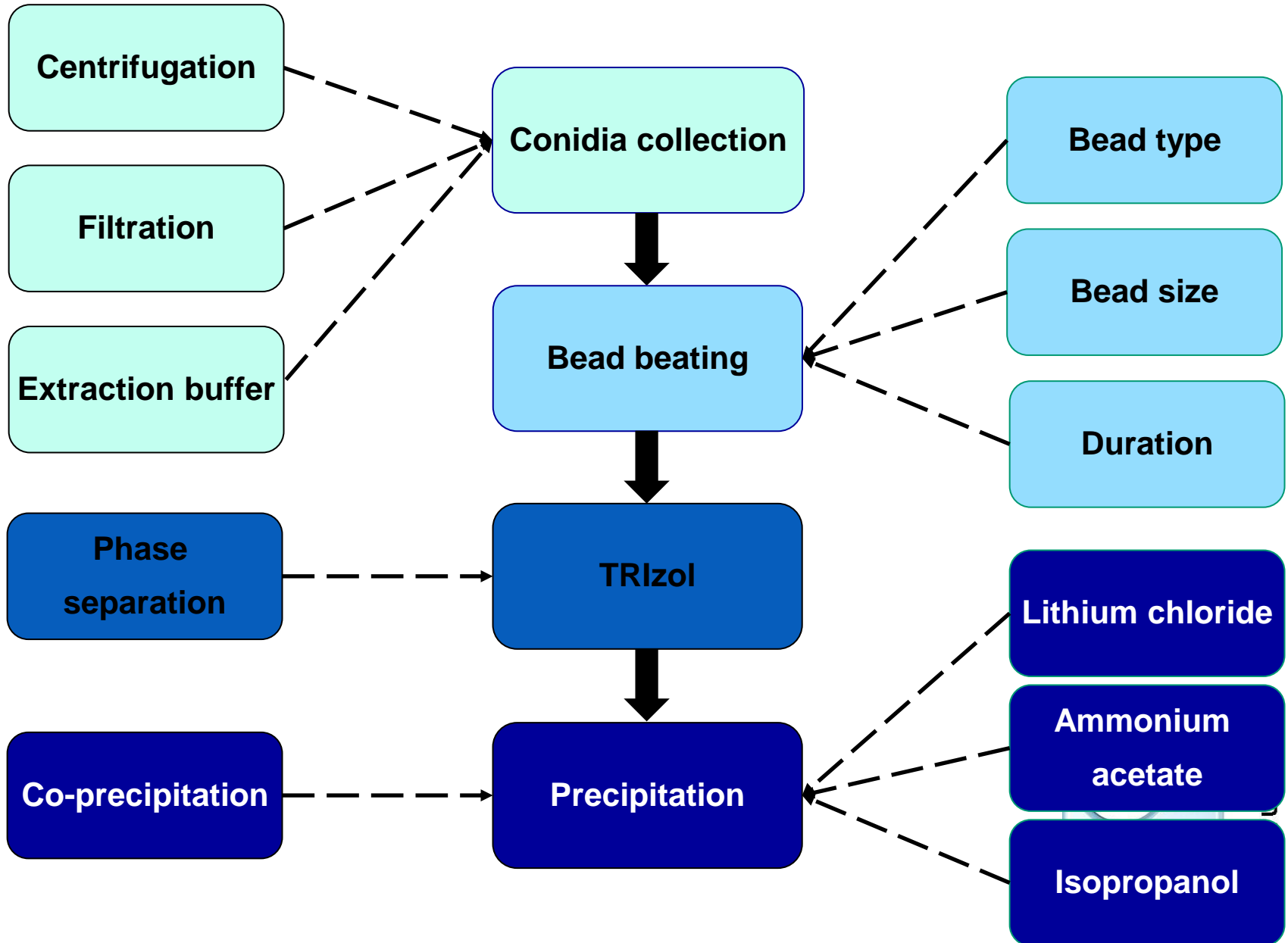
Red Kit



Phase lock tubes

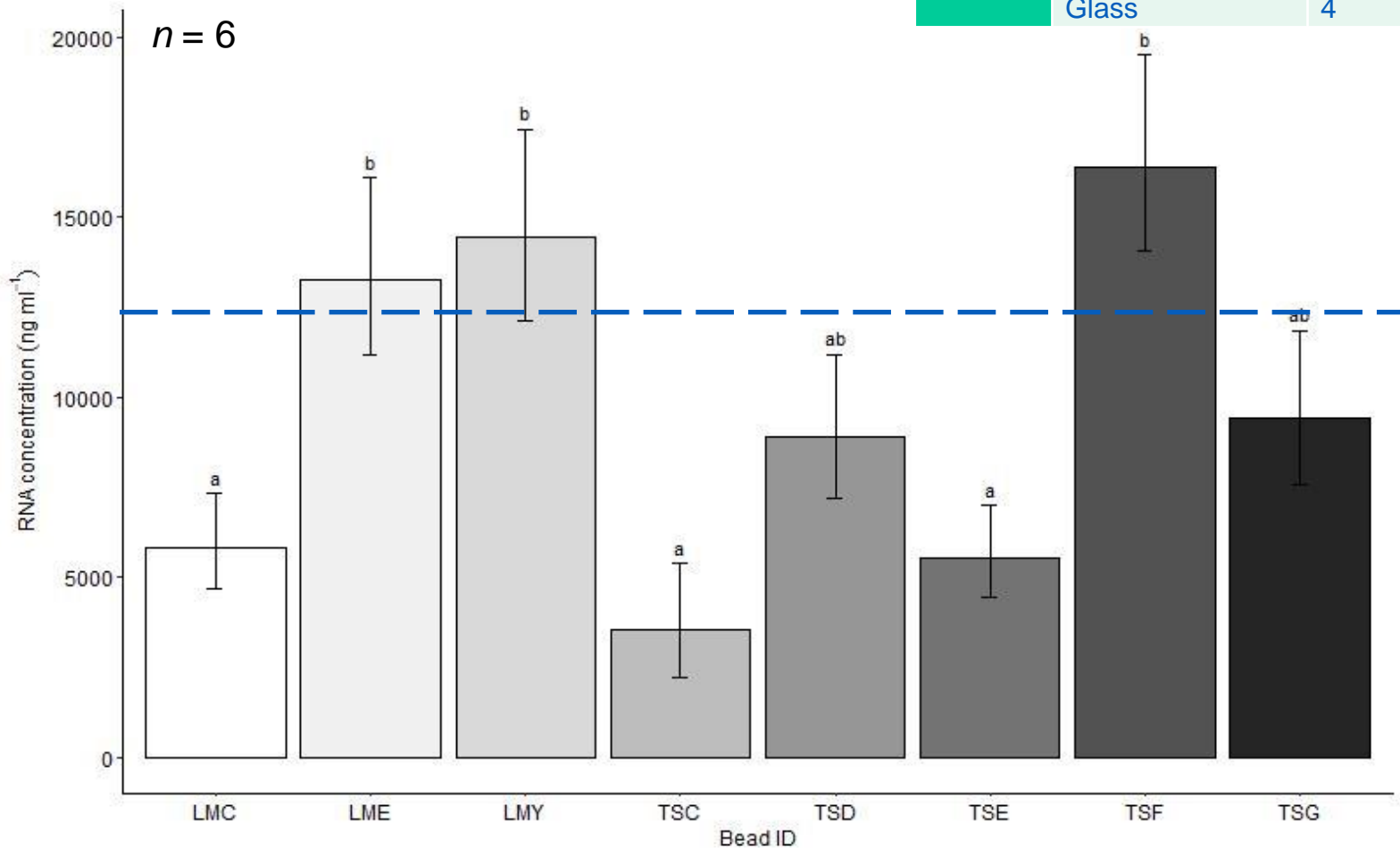


# RNA extraction from pure culture



# Optimising the assay: Bead size

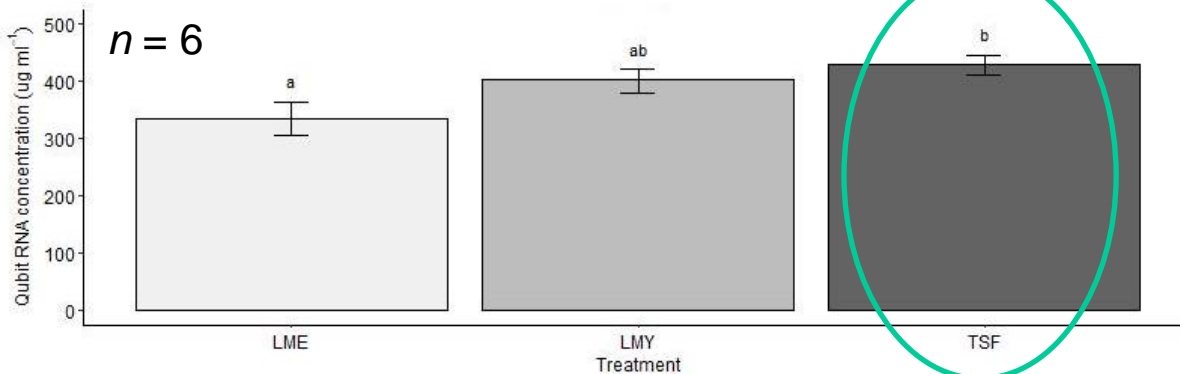
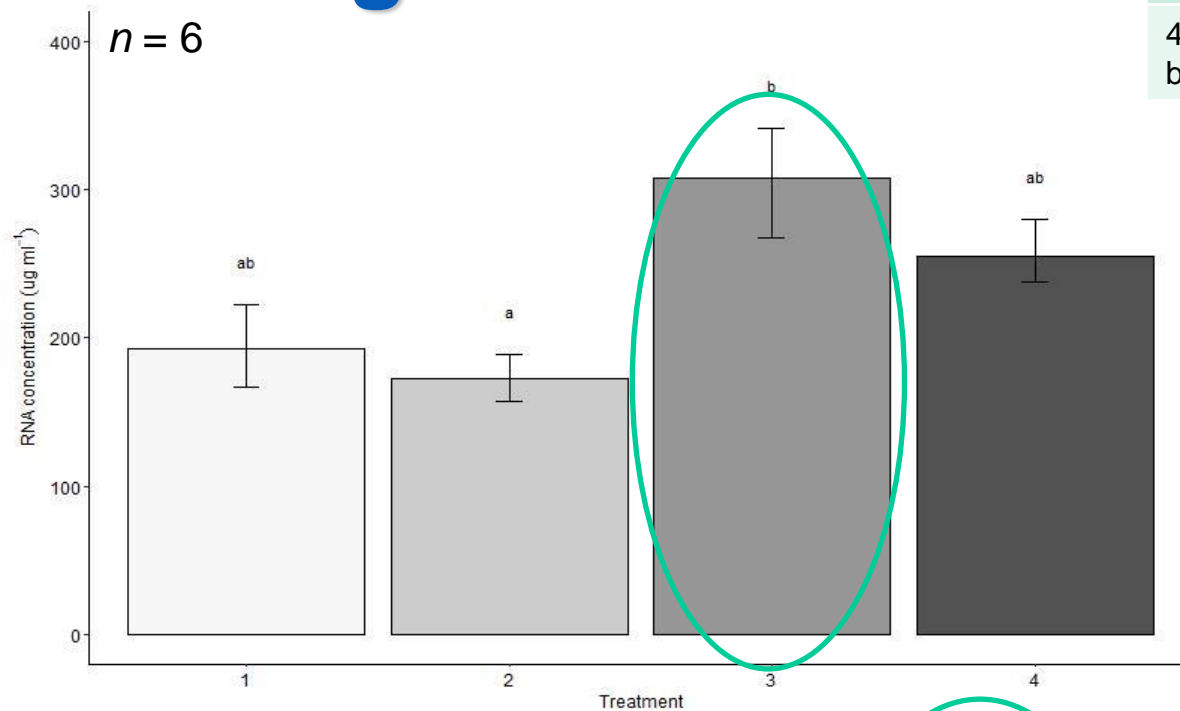
Bead ID	Material	Size (mm)
LMC	Silica	1.0
TSC	Glass	1.0
TSD	Zirconia/silica	1.0
LMY	Zirconium oxide	0.5
TSE	Glass	0.5
TSF	Zirconia/silica	0.1
TSG	Glass	0.1
LME	Ceramic	1.4
	Silica	0.1
	Glass	4



# Optimising the assay: Beating times

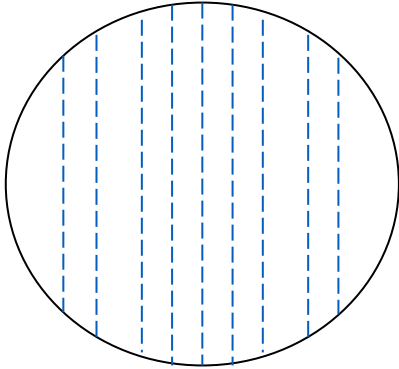
## Beating time

1. 40 sec @ setting 6.5
2. 25 sec, 5 sec break, 25 sec @ 6.5
3. 1 min, 2 min on ice, 1 min @ 6.0
4. As 3, but frozen for 30 min prior to beating

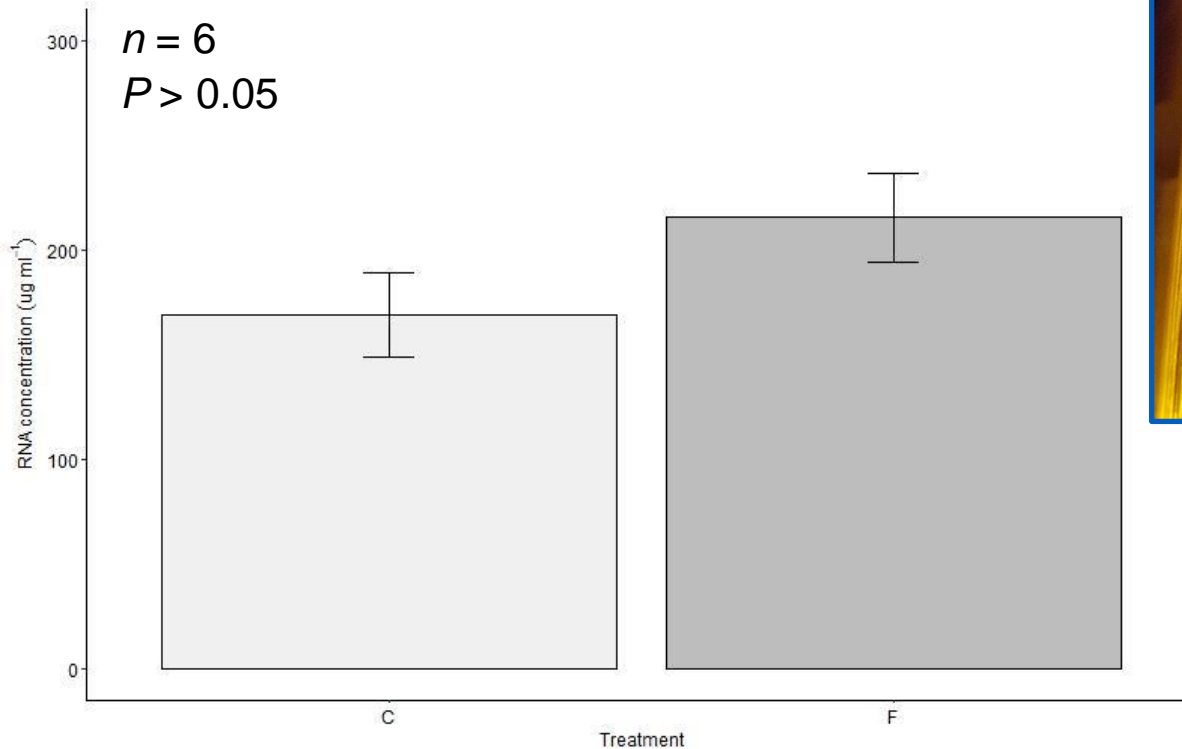




# Optimising the assay: filters

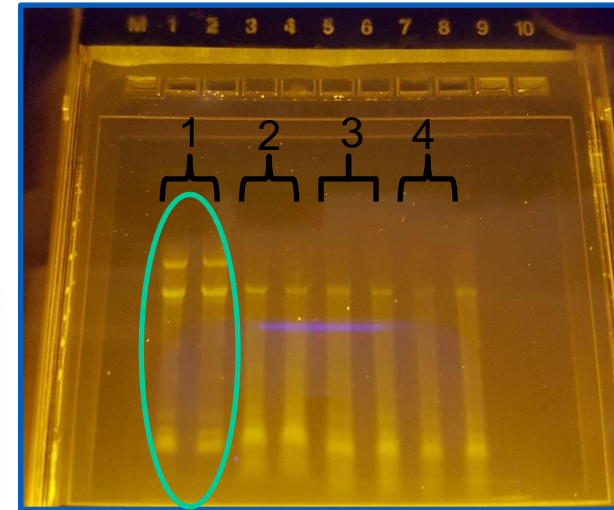


25 mm, 0.8  $\mu\text{m}$   
polycarbonate filter

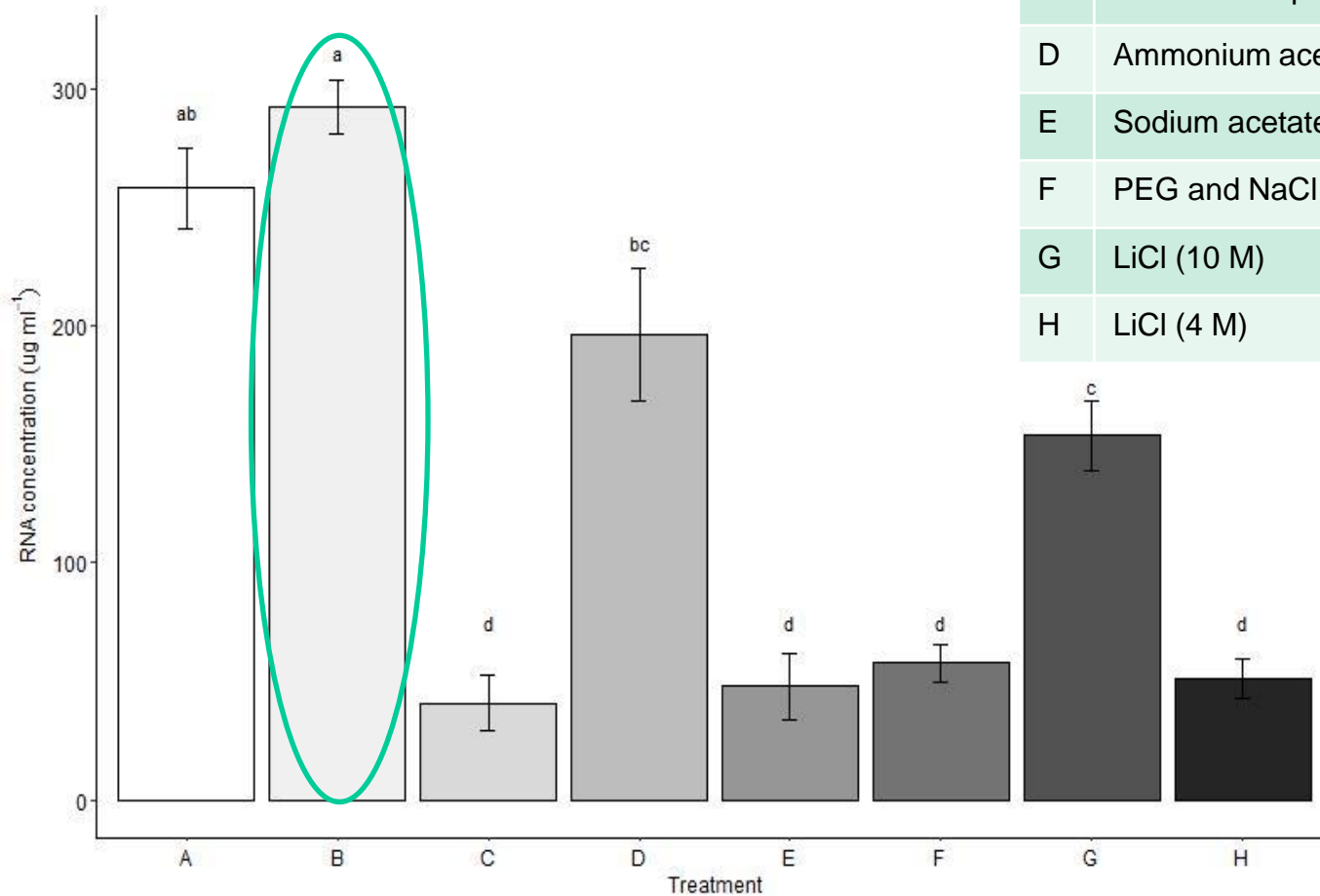


## Beating time

1. 1 min, 2 min on ice, 1 min @ 6.0
2. As above for a total of 5 min
3. 5 min continuous @ 6.0
4. 5 min continuous @ 4.0



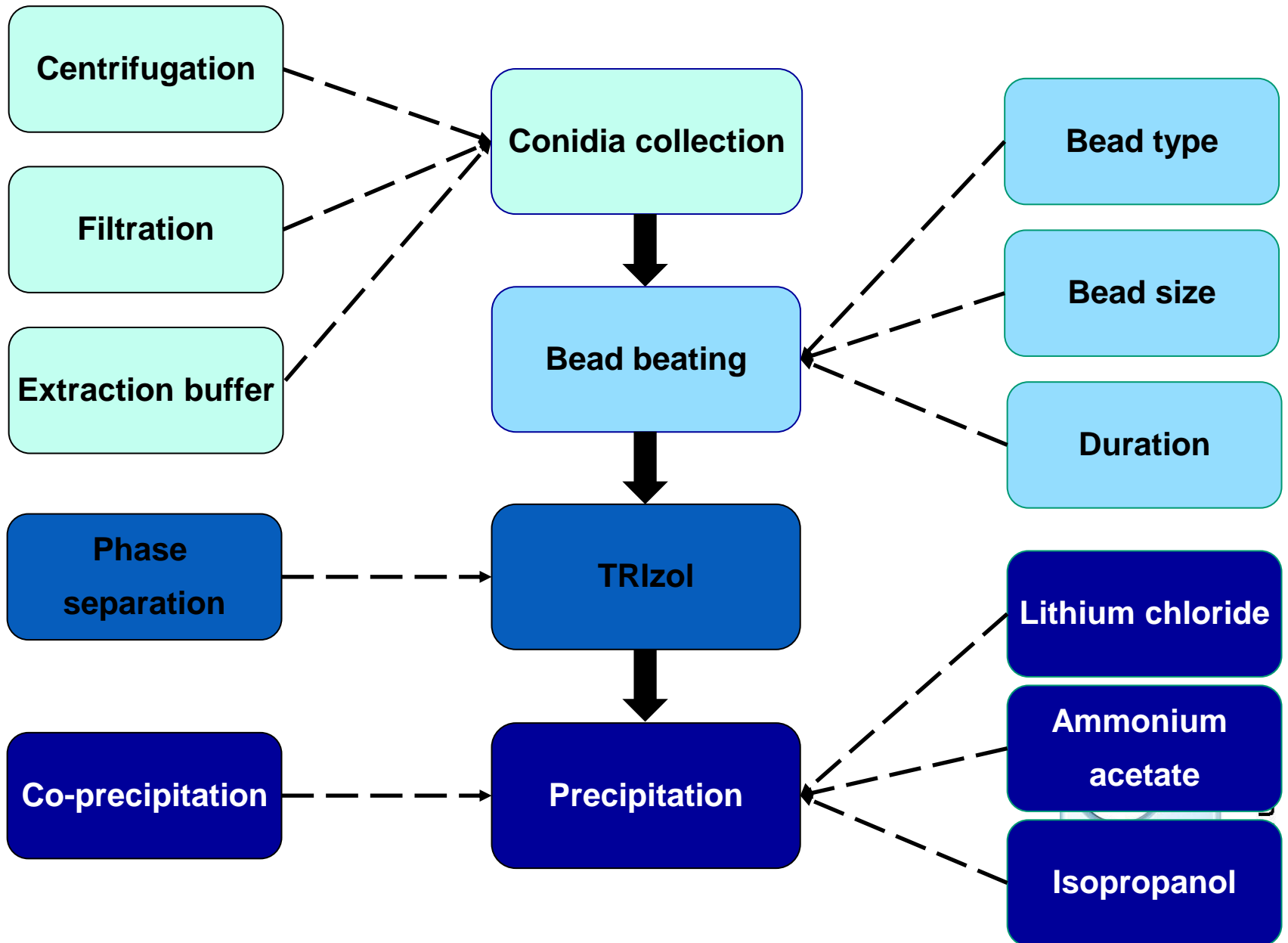
# Optimising the assay: precipitation



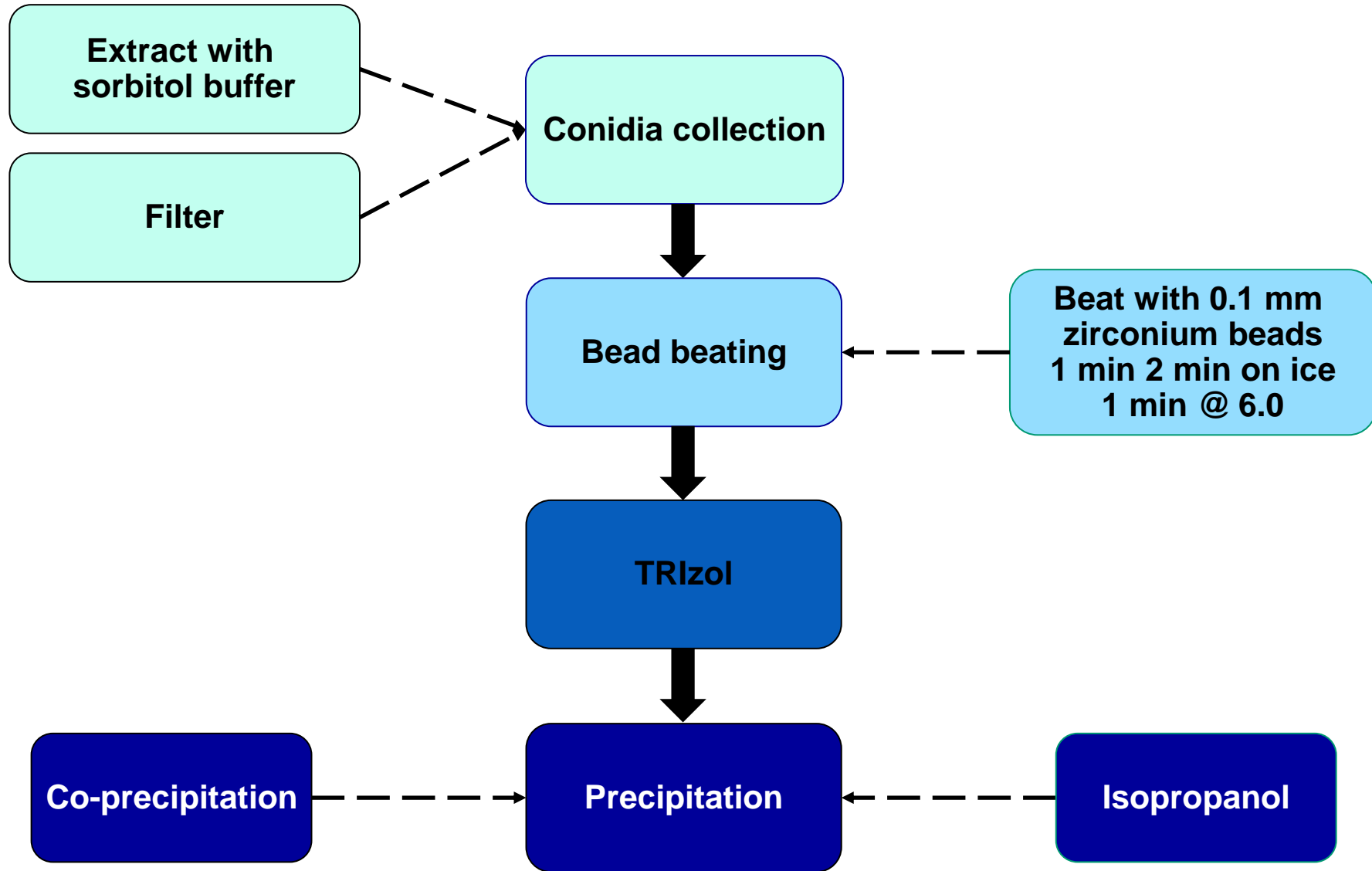
	Precipitation method
A	Isopropanol (12,000g, 10 min)
B	Isopropanol (12,000g, 60 min)
C	NaCl and isopropanol
D	Ammonium acetate and isopropanol
E	Sodium acetate and ethanol
F	PEG and NaCl
G	LiCl (10 M)
H	LiCl (4 M)



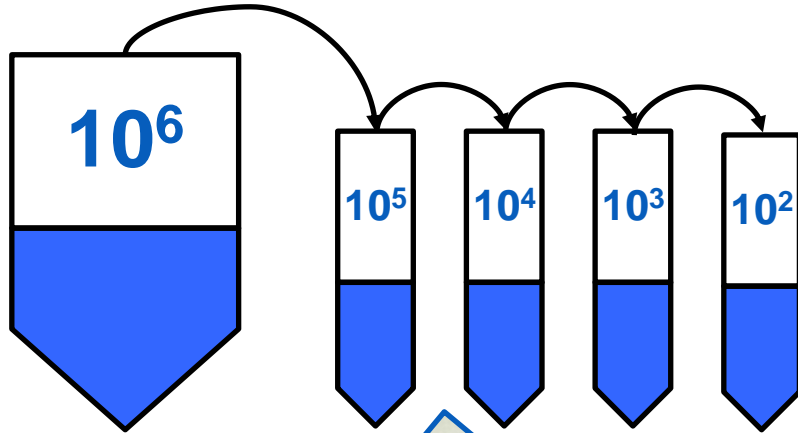
# RNA extraction



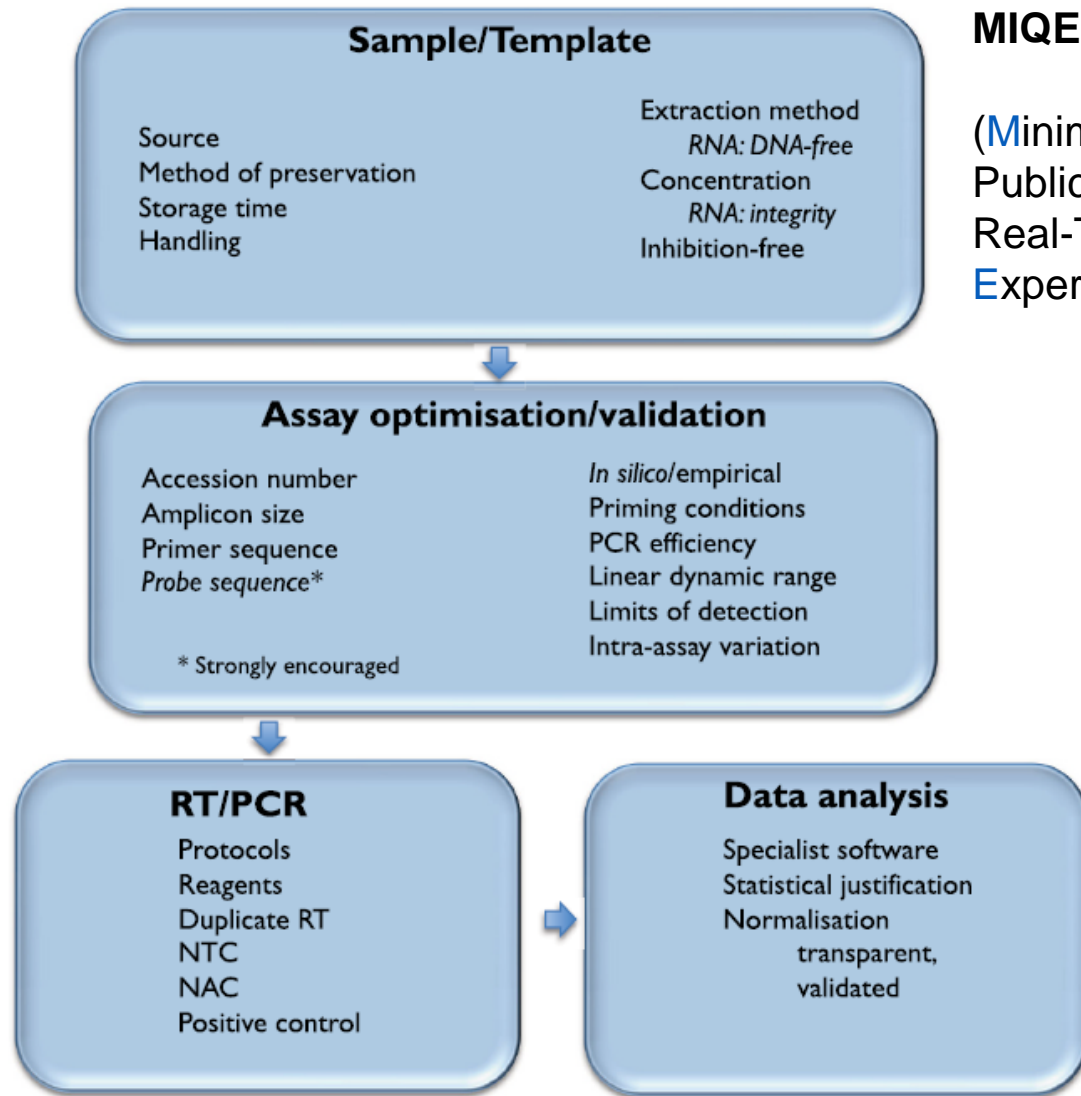
# RNA extraction



# RNA Extraction: What next?



# RT-qPCR assay



## MIQE Guidelines

(Minimum Information for Publication of Quantitative Real-Time PCR Experiments)



Figure 1 Key criteria delineating essential technical information required for the assessment of a RT-qPCR experiment .

# Gene of interest

## FKS1 gene

- Codes for the fksP protein
- Involved in the production of glucan in the cell wall
- Single copy gene

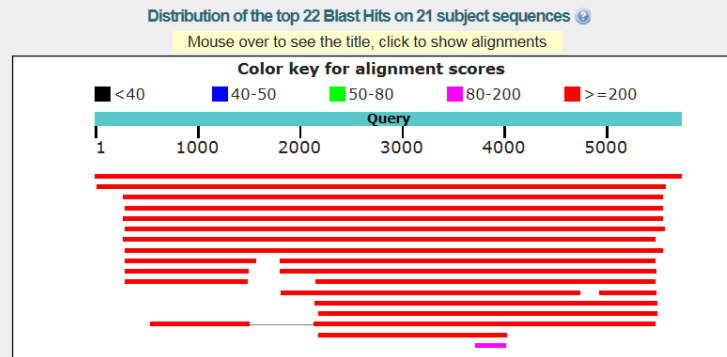
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TCGCCATCAACGCAGGTGTTGCTGGTGTTCCTCGGCATGGCCTGCTGTATGGGACCCATCTTCAGCATGTGCTGCAAGAAATTCGGTGC  
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CTCATTCTGTTCTTGTCTTCTCAT**TGCCCACTTGTTGTTTCGCTCTAT**GGGATTGGTTAAAACTCCCAATCTTCATTCA**ACTTGCTCCAGC**  
**CTTTGAC**AAGGACAACAACGATACTATGGTGACATACCCGGCAACAACATTCCCGCTGGATTTGAGCCAGTTGAGAGCGCCTCGAGCG  
TGGCTACTGCAACGAGCTAG

# Primer specificity

## NCBI Blastn database

Other reports: [Search Summary](#) [Taxonomy reports](#) [Distance tree of results](#) [MSA viewer](#)

### Graphic Summary



### Descriptions

#### Sequences producing significant alignments:

Select: [All](#) [None](#) Selected:0

[Alignments](#) [Download](#) [GenBank](#) [Graphics](#) [Distance tree of results](#)

Description	Max score	Total score	Query cover	E value	Ident	Accession
<input checked="" type="checkbox"/> <a href="#">Neosartorya fischeri NRRL 181 1,3-beta-glucan synthase catalytic subunit FksP (NFIA_058360) partial mRNA</a>	9385	9385	100%	0.0	96%	<a href="#">XM_001258382.1</a>
<input type="checkbox"/> <a href="#">Aspergillus aculeatus ATCC 16872 glycosyltransferase family 48 protein partial mRNA</a>	3897	3897	96%	0.0	79%	<a href="#">XM_020199838.1</a>
<input type="checkbox"/> <a href="#">Cyphellophora europaea CBS 101466 1,3-beta-glucan synthase component FKS1 partial mRNA</a>	2604	2604	91%	0.0	76%	<a href="#">XM_008721799.1</a>
<input type="checkbox"/> <a href="#">Cladophialophora yegresii CBS 114405 1,3-beta-glucan synthase component FKS1 partial mRNA</a>	2556	2556	91%	0.0	76%	<a href="#">XM_007757823.1</a>
<input type="checkbox"/> <a href="#">Cladophialophora carrionii CBS 160.54 1,3-beta-glucan synthase component FKS1 partial mRNA</a>	2505	2505	91%	0.0	76%	<a href="#">XM_008725355.1</a>



# Primer specificity

Species	Strain	Accession no.
<i>Aspergillus fumigatus</i>	Af293	XM_746025.1
<i>Aspergillus clavatus</i>	NRRL 1	XM_001268294.1
<i>Aspergillus flavus</i>	NRRL 3357	XM_002383365.1
<i>Aspergillus nidulans</i>	FGSC A4	XM_656241.1
<i>Aspergillus niger</i>	CBS 513.88	XM_001390940.2
<i>Aspergillus nominus</i>	NRRL 13137	XM_015556015.1
<i>Aspergillus oryzae</i>	RIB40	XM_001816621.2
<i>Aspergillus terreus</i>	NIH 2624	XM_001212456.1
<i>Baudoinia panamericana</i>	UAMH 10762	XM_007675218.1
<i>Candida dubliniensis*</i>	CD36	XM_002417149.1
<i>Capronia coronate</i>	CBS 617.96	XM_007727032.1
<i>Cladophialophora bantiana</i>	CBS 173.52	XM_016768205.1
<i>Cladophialophora carrionii</i>	CBS 160.54	XM_008725355.1
<i>Cladophialophora immunda</i>	Unknown	XM_016395370.1
<i>Cladophialophora yegresii</i>	CBS 114405	XM_007757823.1
<i>Cyphellophora europea</i>	CBS 101466	XM_008721799.1
<i>Exophiala oligosperma</i>	Unknown	XM_016405660.1
<i>Exophiala spinifera</i>	Unknown	XM_016384822.1
<i>Fonsecaea erecta</i>	Unknown	XM_018837737.1
<i>Fonsecaea pedrosoi</i>	CBS 160.54	XM_013423062.1
<i>Neosartorya fischeri</i>	NRRL 181	XM_001258382.1
<i>Neurospora tetrasperma</i>	FGSC 2508	XM_009854667.1
<i>Penicillium chrysogenum Wisconsin</i>	54-1255	XM_002564403.1
<i>Penicillium digitatum</i>	Pd1	XM_014679162.1
<i>Saccharomyces cerevisiae</i>	S288C	NM_001182231.1
<i>Talaromyces atrovirens</i>	Unknown	XM_020260475.1
<i>Talaromyces marneffeii</i>	ATCC 18224	XM_002147459.1
<i>Talaromyces stipitatus</i>	ATCC 10500	XM_002481595.1
<i>Trichoderma gamsii</i>	Unknown	XM_018810875.1
<i>Ustilago maydis</i>	521	XM_011389324.1
<i>Verticillium albo-atrum*</i>	VaMs.102	XM_003001921.1

**In silico  
specificity:  
Sequence is  
specific for *A.  
fumigatus***



# Primer specificity

Species	Strain
<i>Aspergillus fumigatus</i>	Af293
<i>Aspergillus alliaceus (Petromyces)</i>	DSMZ 813
<i>Aspergillus clavatus</i>	DSMZ 816 (NRRL 1)
<i>Aspergillus flavus</i>	DSMZ 1959
<i>Aspergillus nidulans (Emericella)</i>	DSMZ 820
<i>Aspergillus niger</i>	DSMZ 2143
<i>Aspergillus terreus</i>	DSMZ 826
<i>Neosartorya fischeri</i>	DSMZ 3700 (NRRL 181)
<i>Penicillium glabrum</i>	DSMZ 2017

**In vivo specificity:**  
**Awaiting results**



# RT-qPCR assay: What next?

- Test the primers against microorganisms similar to *A. fumigatus*
- Test assay against *A. fumigatus* isolates that have been collected from field samples



# And finally...

Air sampling + RNA extraction+ RT-qPCR assay =  
Improved method of *A. fumigatus* detection

Expand the methods to include other pathogens of  
interest as part of our commercial Pathogen Suite



# References

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**Any Questions?**

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