



**C3 component deposition on  
*Salmonella* O48 cells characterized by  
sialylated lipopolysaccharide  
and different pattern of outer  
membrane proteins**

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# Introduction

Mechanisms employed by pathogens to evade host immunological defenses are not entirely understood

No direct correlation between the C3 protein deposition pattern and bacterial resistance has been observed

This is the first analysis of C3 activation on *Salmonella* isolates belonging to the O48 serogroup and their surface antigens: sialylated lipopolysaccharide (LPS) and outer membrane proteins (OMPs)

Sialylated LPS of other bacteria is described in the context of molecular mimicry connected to the onset of autoimmunity in humans. Sialylation enhances ability to bind human factor H

The mechanisms of OMPs-mediated complement activation are currently intensively investigated, because of principal antigen vaccine potential



# Introduction

## WHY SALMONELLA?

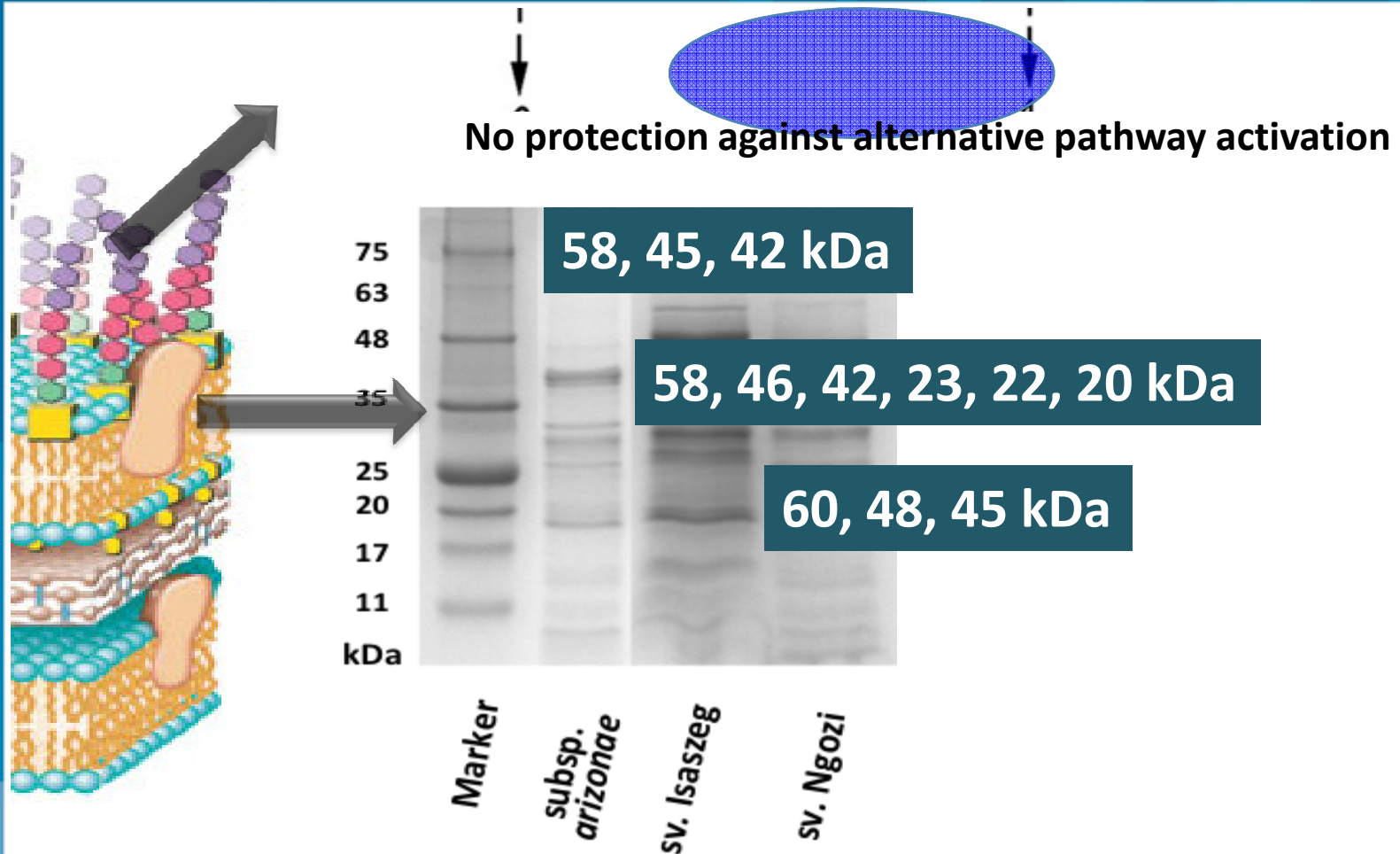
- *Salmonella* infections are among the most common food-borne infections affecting humans in the European Union
- Elderly and otherwise weak patients are more prone to developing severe blood infection
- Post-infectious complications, such as reactive joint inflammation occur in about 10% of the cases  
(*European Centre for Disease Prevention and Control*)

## WHY SALMONELLA OF SEROTYPE O48 ?

***Salmonella* O48 is the only group possessing sialylated LPS among all known *Salmonella* bacteria**



## Introduction: characterization of *Salmonella* O48 LPS and OMPs



- Futoma-Kołodziej B. Bacterial outer membrane proteins - dependent complement activation. *J Mol Immunol* (2016) 1:1
- Futoma-Kołodziej B. Immune response against bacterial lipopolysaccharide. *J Mol Immunol* (2016) 1:1
- Futoma-Kołodziej et al. Presumable role of outer membrane proteins of *Salmonella* containing sialylated lipopolysaccharides serovar Ngozi, sv. Isaszeg and subspecies arizonae in determining susceptibility to human serum. *Gut Pathog* (2015) 7:18.
- Futoma-Kołodziej B. et al. Searching for outer membrane proteins typical of serum-sensitive and serum-resistant phenotypes of *Salmonella*. *InTech, Croatia, Ch 14, 265-290*



# Materials and methods

## Human serum

nonimmune HS was taken from 20 healthy donors in Regional Centre of Transfusion Medicine and Blood Bank the name of Prof. T. Dorobisz in Wrocław, Poland. It was conducted according to the principles expressed in the Declaration of Helsinki.

## C3 concentration

was estimated with radial immunodiffusion method using specific antibodies (The Binding Site, UK).

## Bactericidal assay

was performed according to Doroszkiewicz (1997). The number of colony-forming units (CFU/ml) at time 0 was taken as 100%. Strains with survival rates below 90% after 45 min of incubation were classified as serum sensitive.

## ELISA

Enzyme-linked immunosorbent assay for bacterial cells was carried out by indirect ELISA according to Alberti et al. (1996). Complement C3 activation assay for LPS was performed by indirect sandwich ELISA (Holmskov-Nielsen, 1986).



# Materials and methods

## LPS isolation

Isolation of lipopolysaccharides (LPSs) was performed with an LPS Extraction Kit according to the manufacturer's instruction (Intron Biotechnology, Korea).

## OMP isolation

Isolation of outer membrane proteins (OMPs) was performed with the detergent Zwittergent Z 3-14 (Calbiochem, USA) according to Murphy and Bartos (1989).

OMPs quantifications were done with a bicinchoninic acid (BCA) Protein Assay Kit (Pierce, USA).

## Electrophoresis & immunoblotting

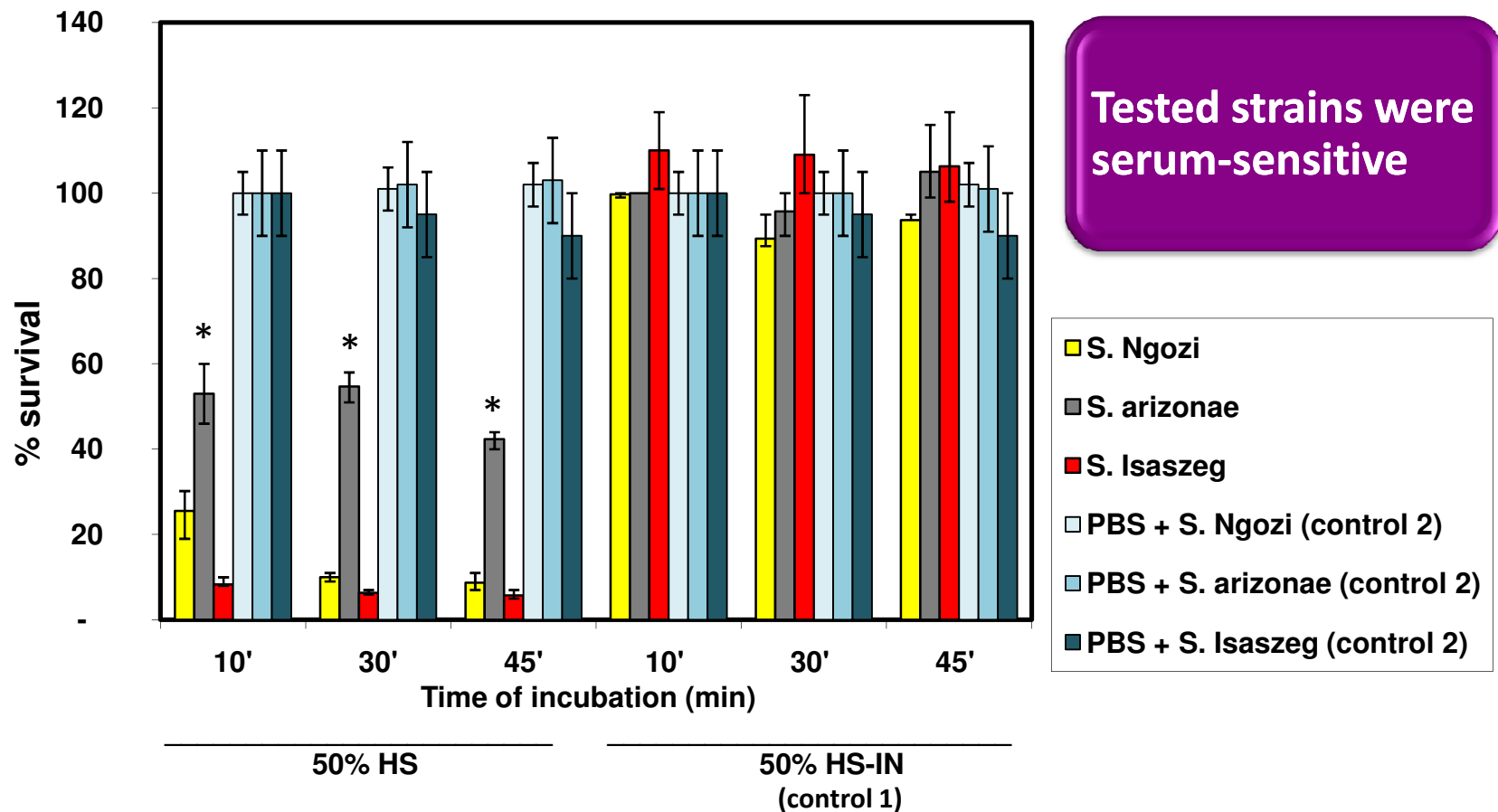
Electrophoresis of OMPs under reducing conditions (SDS-PAGE) by method of Laemmli (1970) and in native (nonreducing) conditions (BN-PAGE) according to Swamy et al. (2006). OMPs were transferred to PVDF membranes and immunoblotted to detect C3 fragments bound to OMPs.

## Statistical analysis

Data for the bactericidal activity of HS were analyzed using the ANOVA Kruskal-Wallis test. In the ELISA tests the ANOVA Friedman and Kendall rank correlation coefficient test was used (Statistica.pl v. 9.0, Statsoft, Kraków, Poland).



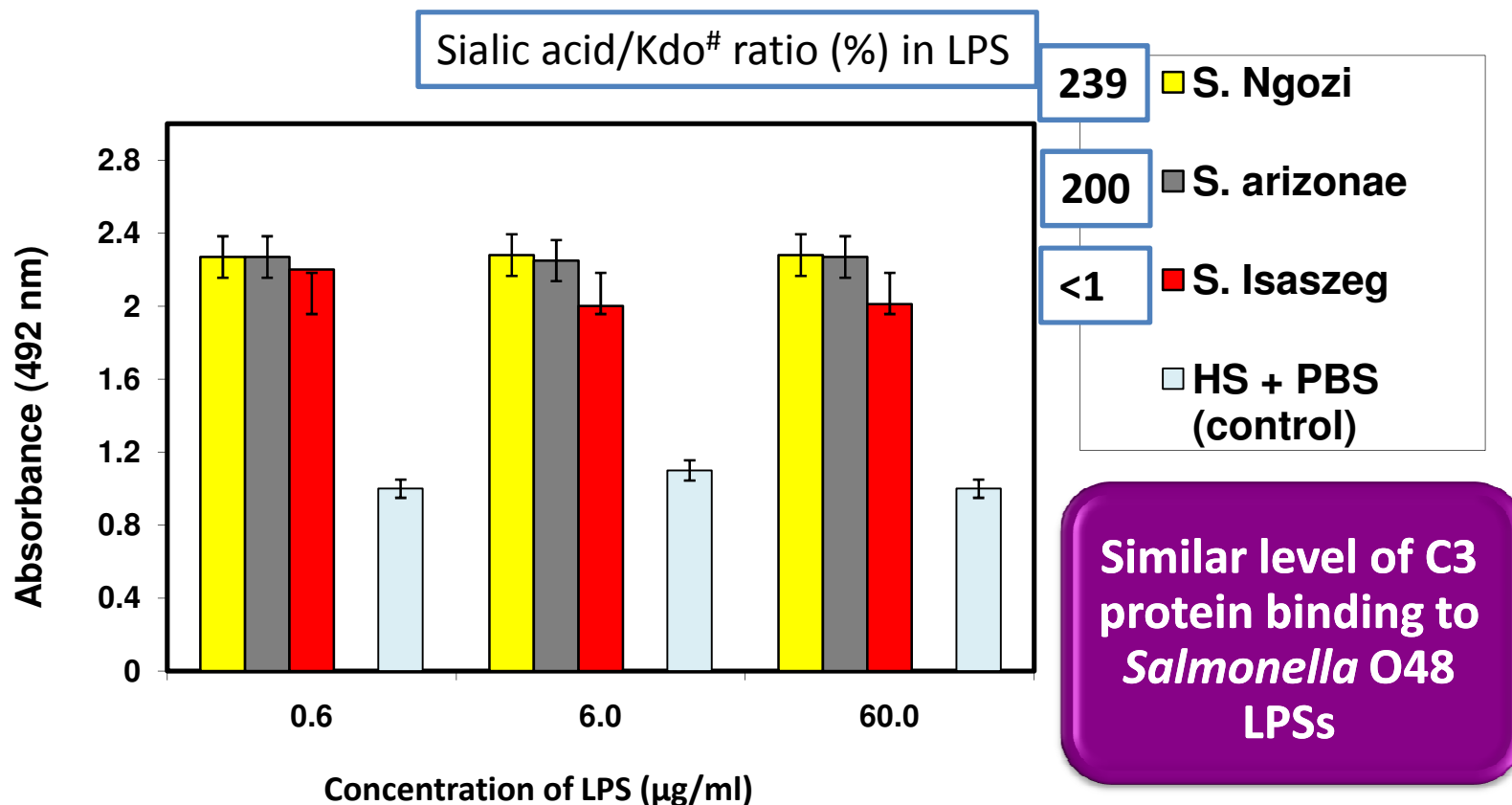
## Results: susceptibility of *Salmonella* strains to the antibacterial activity of human serum (HS)



**Figure 1** Log-phase cultures of the bacteria ( $1 \times 10^5$  CFU/ml) were incubated in 50% human serum (HS), in 50% heat inactivated serum (56°C for 30 min, HS-IN, control 1) or PBS (control 2) for 45 min. Serial dilutions were performed to calculate colony forming units (CFU/ml). The average number of colonies was estimated from three plates. The CFU/ml at time 0 was taken as 100%. Sensitivity to HS differs significantly if  $p$  values are less than 0.05 (\*).



## Results: C3 complement protein depositions on immobilized LPS

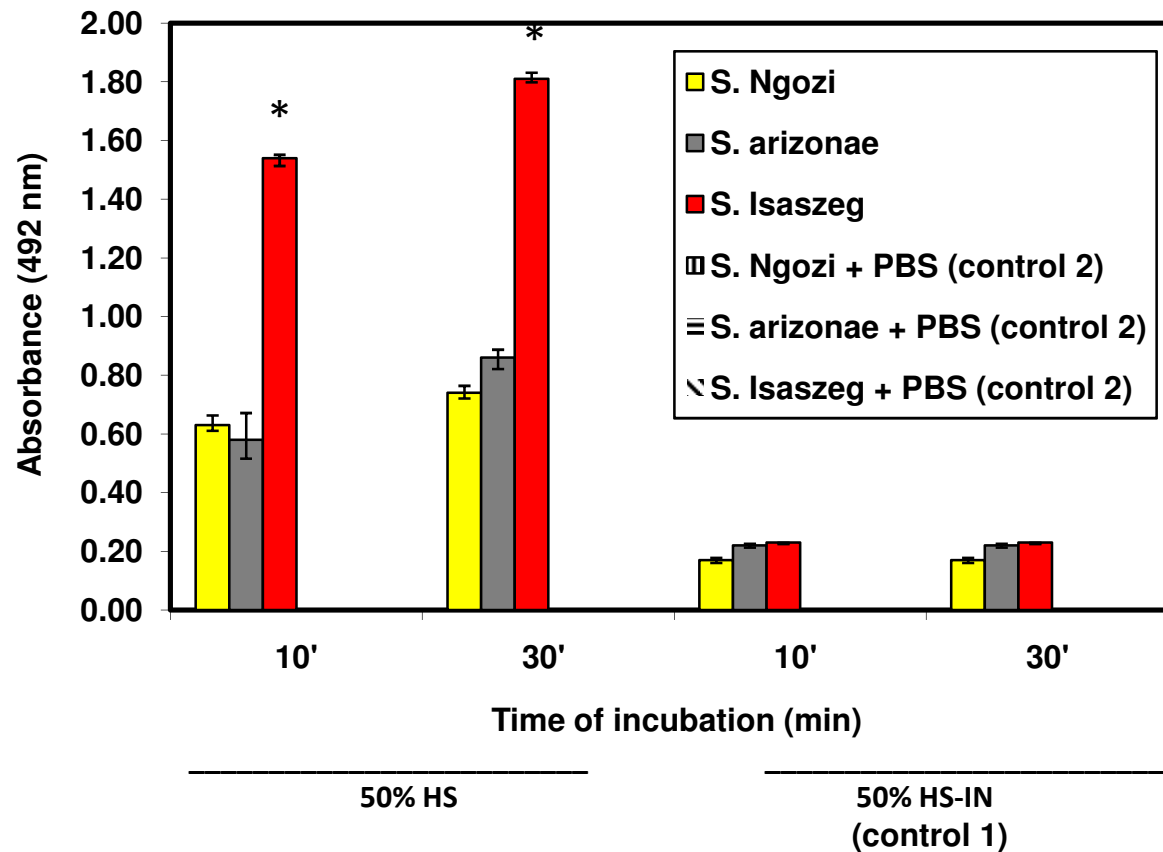


**Figure 2** Sandwich ELISA. Microtiter plate wells coated for 2 h at 37°C with polyclonal rabbit anti-C3c diluted 1/500 in 0.1 M sodium carbonate buffer (pH 9.6). Mixtures of LPSs and 80% HS were incubated for 15 min at 37°C. Mixtures transferred into titration plates and incubated for 45 min at 37°C. C3c detection with polyclonal rabbit anti-C3c (Dako) antibodies diluted 1/2000 in 1% BSA in PBS (pH 7.4). Adding of polyclonal goat anti-rabbit immunoglobulins/HRP diluted 1/2000 in 1% BSA in PBS (incubation 1 h, 37°C). Reading: OPD substrate tablets, at 492 nm.





## Results: C3 complement protein depositions on *Salmonella* cells

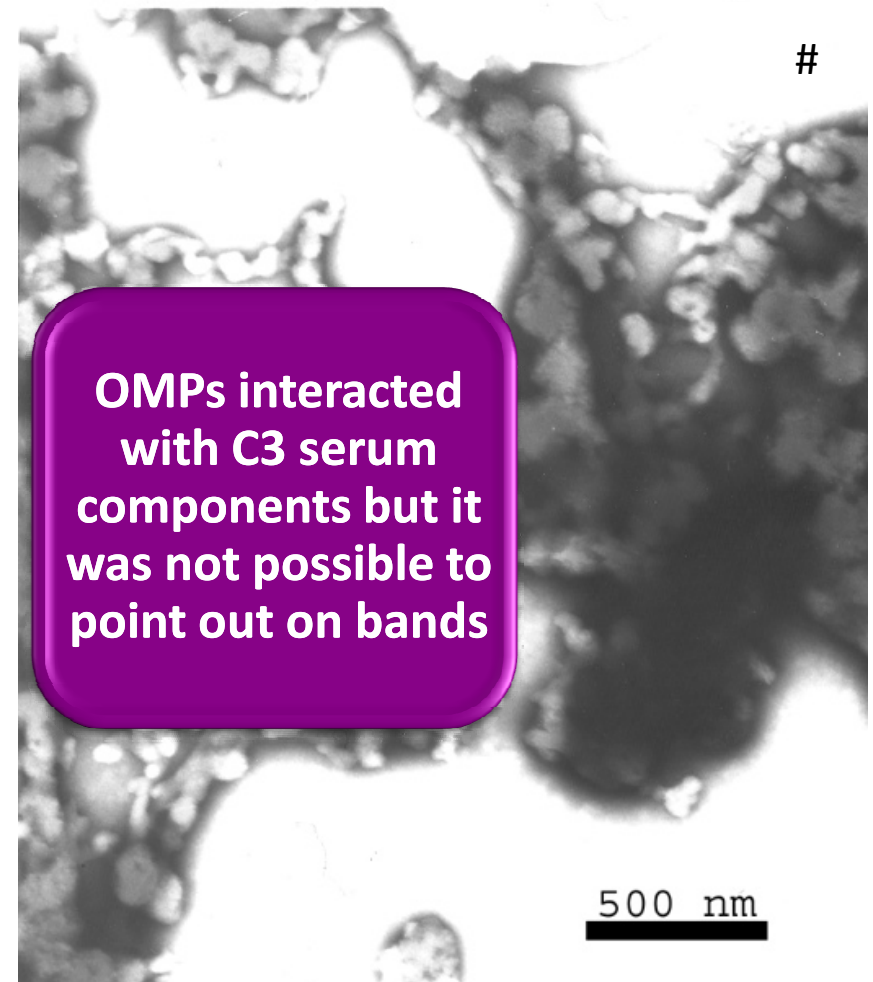
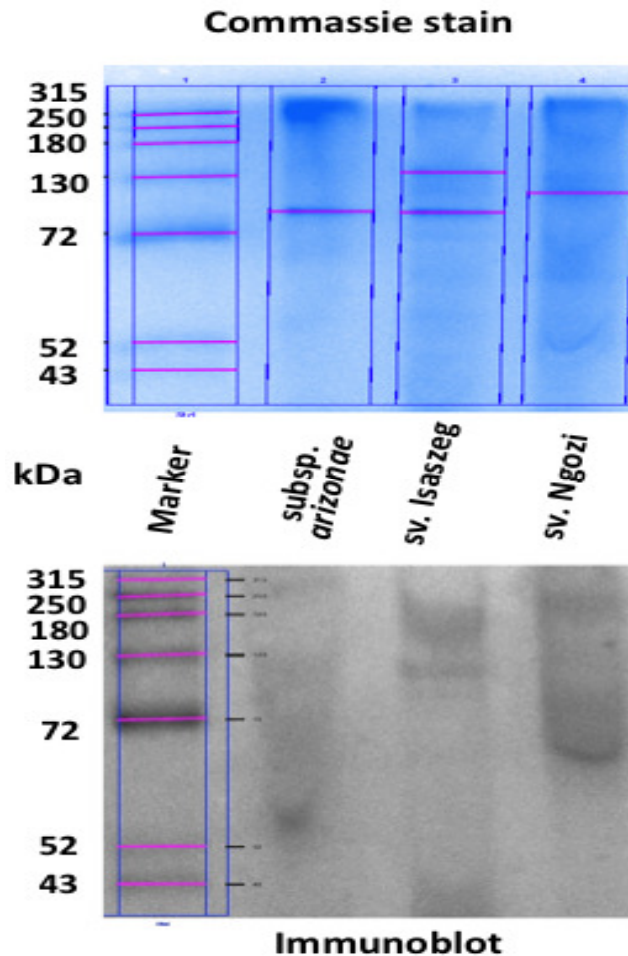


The highest C3 deposition rate was noted for *Salmonella* sv. Isaszeg

**Figure 3** Indirect ELISA. Bacterial cells in log-phase ( $1 \times 10^7$  CFU/ml) were incubated in 50% HS, 50% HS-IN (control 1) or PBS (control 2) for 30 min at 37°C. The same antibodies as were in Sandwich ELISA. Activation of C3 differs significantly if the  $p$  values are less than 0.005 (\*).

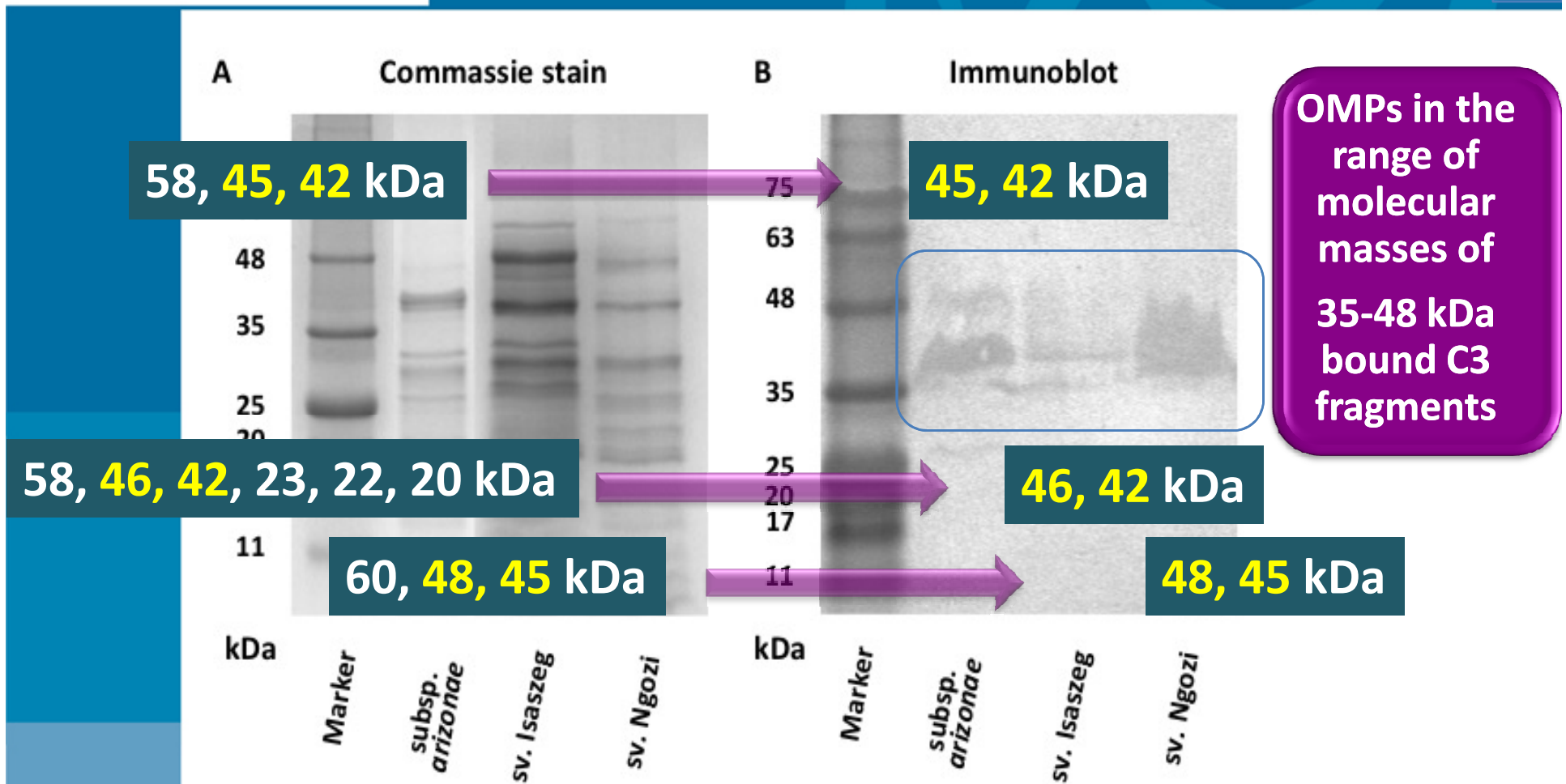


## Results: immunoblot detection of C3c fragments on native (non-denatured) OMPs



**Figure 4** OMPs isolated with Zwittergent Z 3–14 detergent®. OMP patterns were determined by blue native polyacrylamide gel electrophoresis (BN-PAGE) and C3 binding confirmed by Western blotting. Electrotransfer conducted at 100 V for 1 h. Lane 1 molecular-weight marker 26625 (Thermo Scientific). The OMPs concentrations were 10 µg/well.

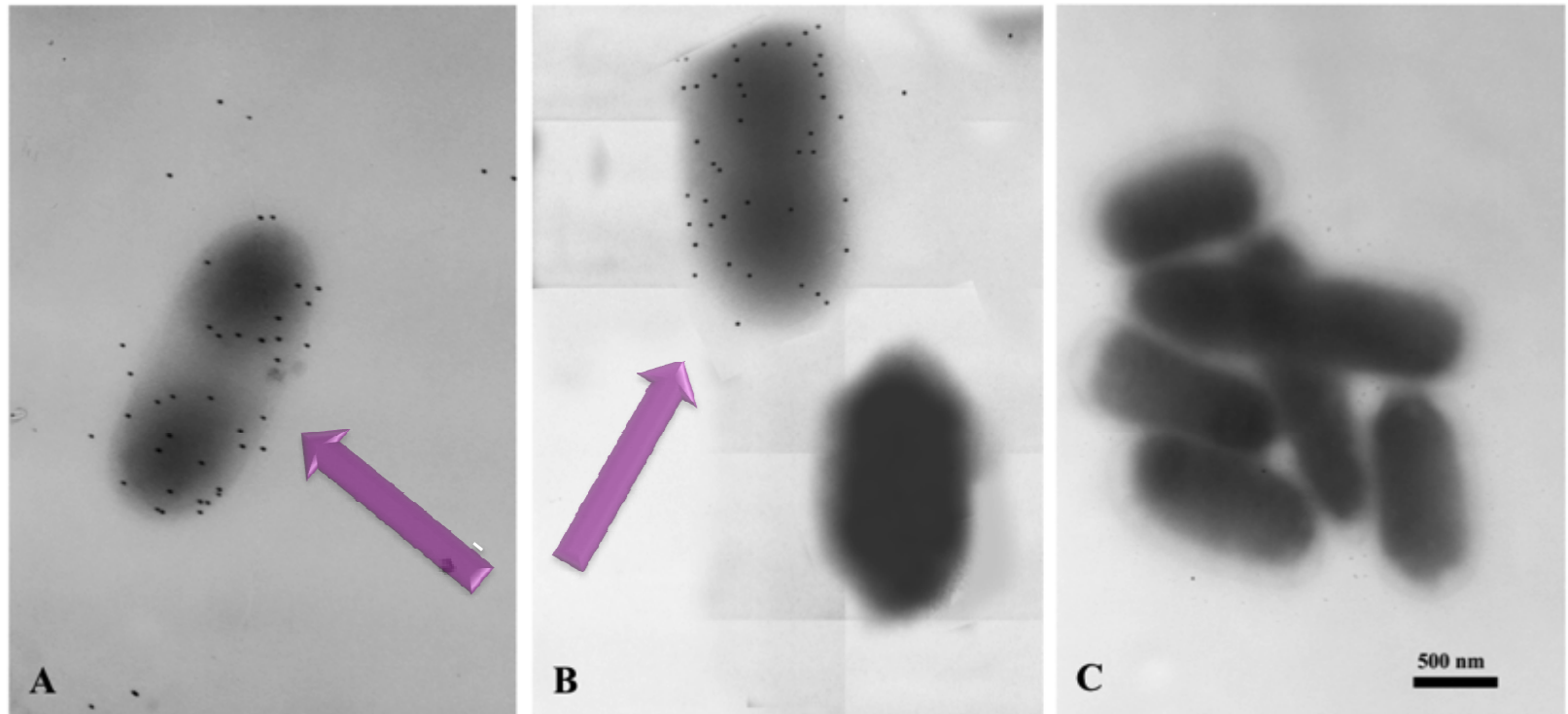
## Results: immunoblot detection of C3c fragments on OMPs under reducing conditions



**Figure 5** Z 3–14 detergent<sup>®</sup> OMPs patterns were determined by sodium dodecyl sulphate-polyacrylamide gel electrophoresis (SDS-PAGE) (10 µg/well) (A) and C3 binding confirmed by Western blotting, (20 µg/well) (B). Electrotransfer conducted at 50 V for 1 h. Lane 1 molecular-weight marker A8889 (AppliChem).



## Results: detection of C3c fragments bound to *Salmonella* O48 cells using TEM microscopy



**Figure 6** TEM micrographs of negatively stained *Salmonella* cells. Bacteria in different stages of growth were incubated in human serum (A, B) were labelled with polyclonal anti-human C3c complement antibodies (Dako, Denmark) and then with protein A-gold. Finally, the specimens stained with 2% aqueous uranyl acetate. The samples were examined under a Tesla BS 540 electron microscope. **A:** labelled cell during the division process, **B:** labelled and not labelled cells not divided, **C:** bacteria not cultured in serum (negative control). Bar indicates 500 nm (data not published).



# Conclusions

I  
No association between C3c deposition on the *Salmonella* O48 cells and their susceptibility to HS was found

II  
C3 activation by LPS was not dependent on the amount of sialic acid in LPS

III  
OMPs of molecular masses: 42, 45, 46, 48 kDa probably determine *Salmonella* O48 sensitivity to HS through C3 activation



# Applications

Gram-negative bacteria with their potential to change the chemical structure of LPS may become distinct strains containing the unique surface antigens patterns. It may generate serious clinical problems of global interest to eliminate bacteria occurring systemic infections with current available drugs

LPS characterization supports the development of a new vaccine against non-typhoidal *Salmonellae* for Africa

Finding factors that control C components deposition on bacterial cells is the way to explain the sophisticated mechanism of bacteraemia leading to sepsis

The phenomenon of mimicry as a cause of autoimmune diseases is unknown. In the future, the premises like those presented may be helpful to construct antibacterial vaccines against autoimmune-connected infections



# Acknowledgements

For co-authors of the main manuscript

*“Presumable role of outer membrane proteins of Salmonella containing sialylated lipopolysaccharides serovar Ngozi, sv. Isaszeg and subspecies arizonae in determining susceptibility to human serum”.*  
*Gut pathogens 7: 18 (2015)*

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