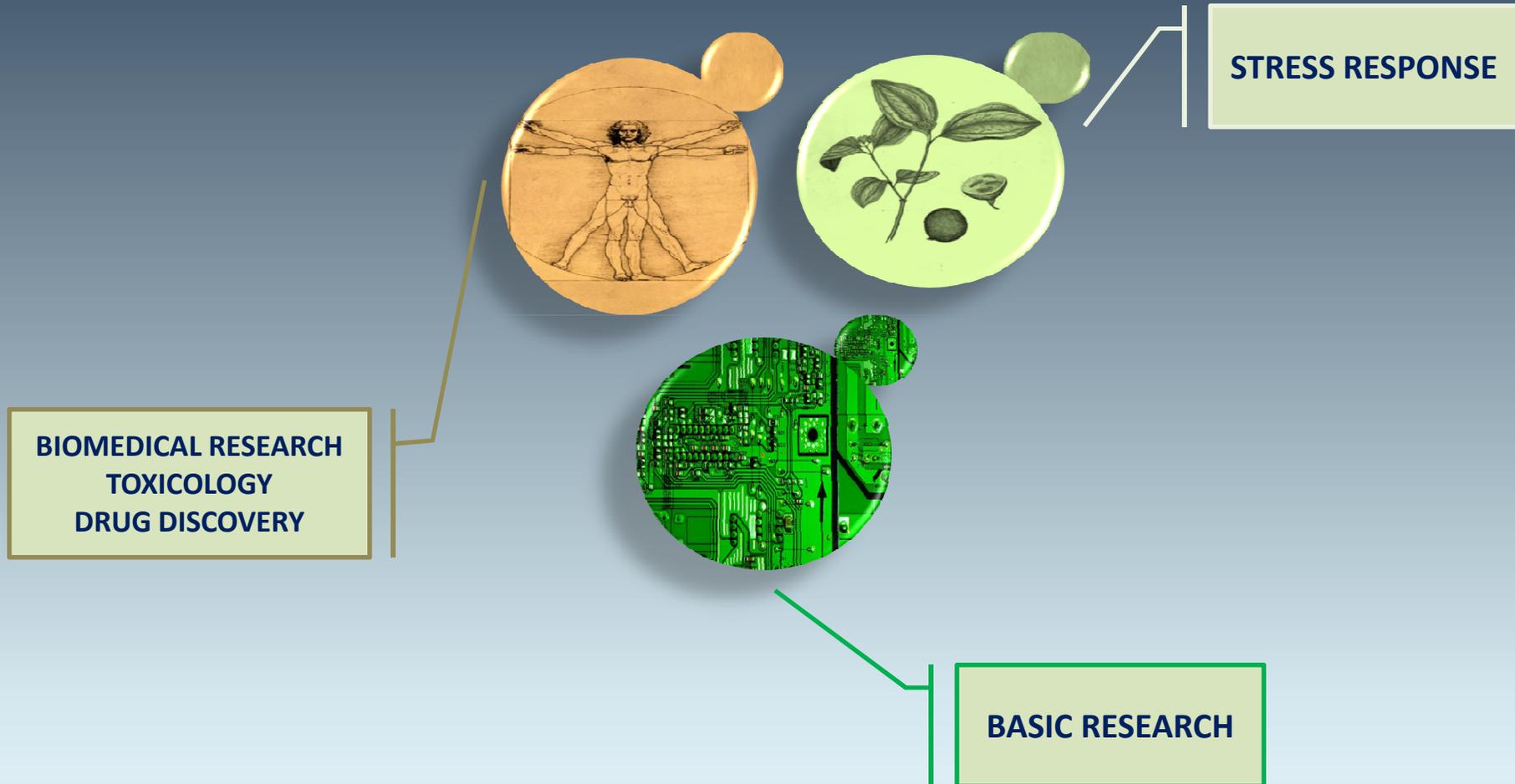


Real time analysis of gene expression in  
living yeast cells: Novel approach for  
accurate cell damage detection

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# *Saccharomyces cerevisiae* as a Model Organism

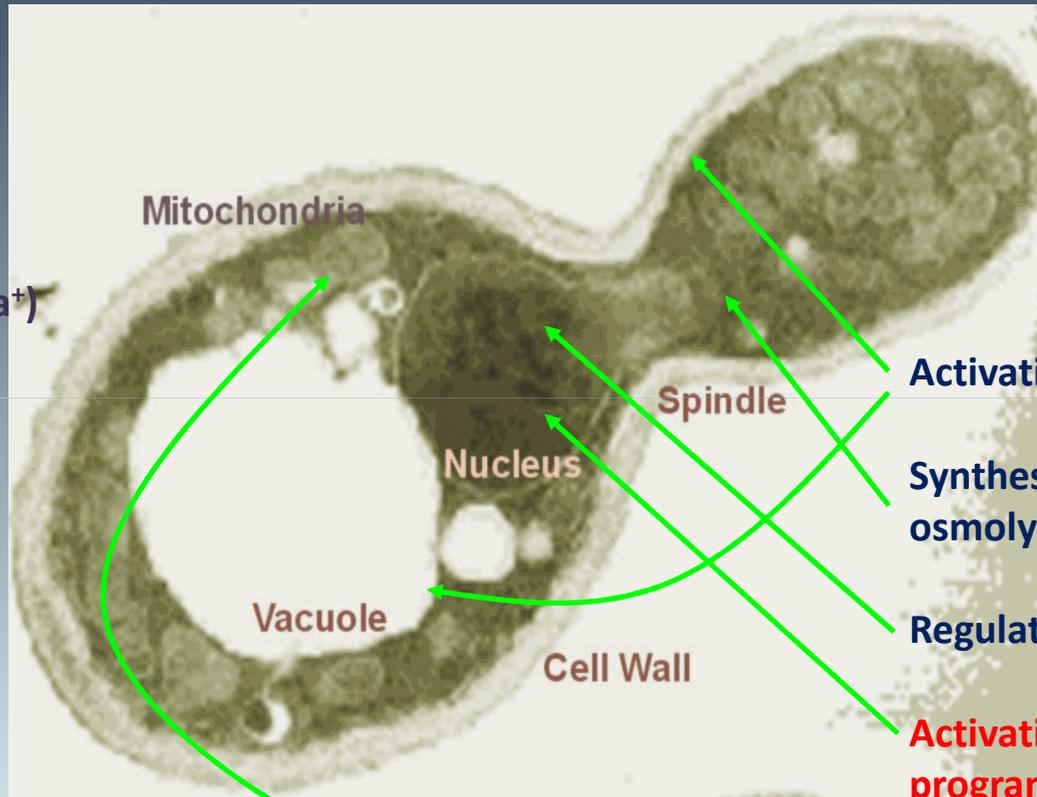


# Yeast: A model for understanding cellular stress responses

## Hyperosmotic Stress



- Water efflux
- Cell shrinking
- Ionic imbalance ( $\text{Na}^+$ )
- Oxidative damage



## Defense Mechanisms



Activation of ion transport

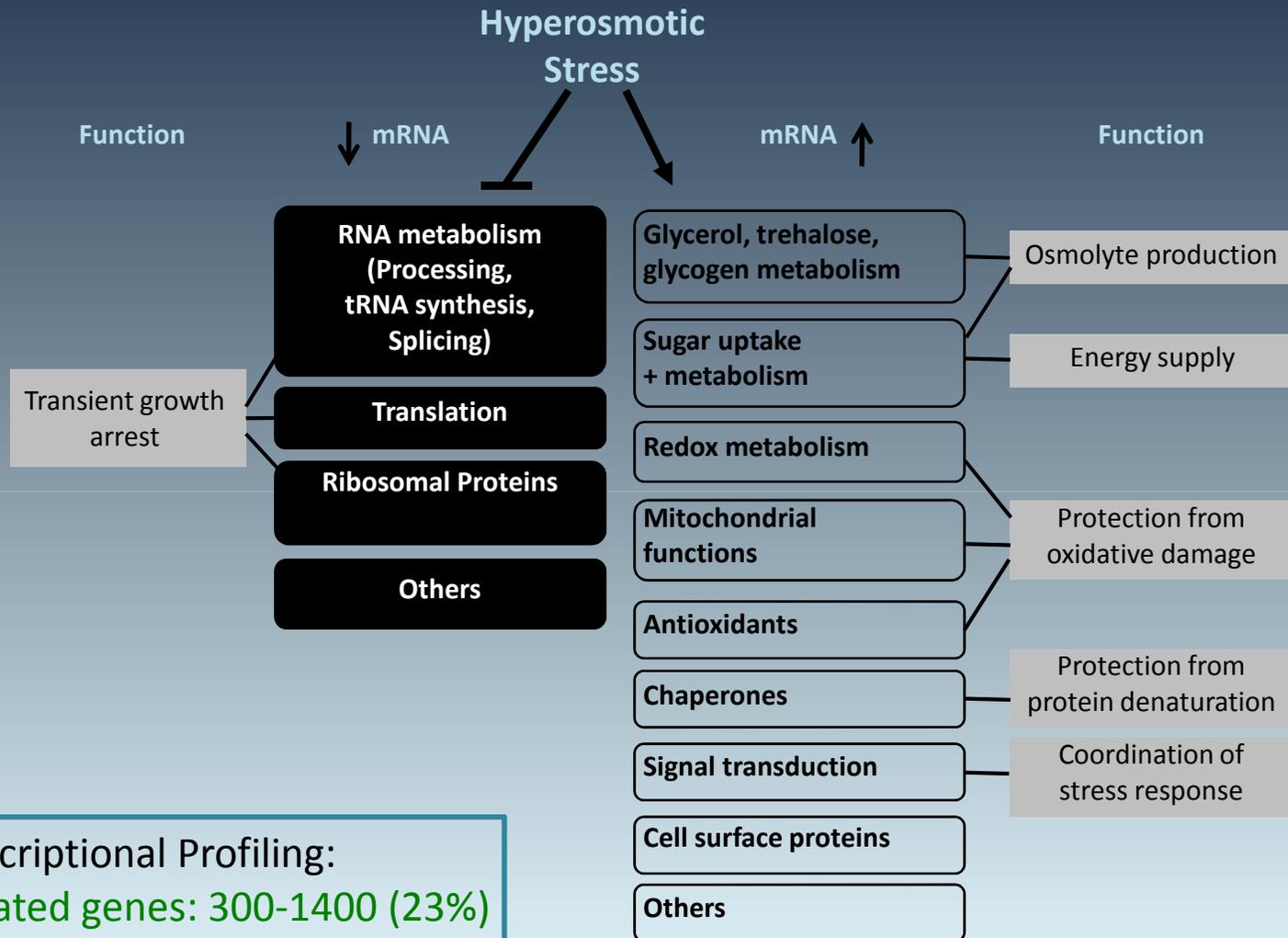
Synthesis/accumulation of osmolytes (glycerol)

Regulated cell cycle arrest

Activation of transcriptional program

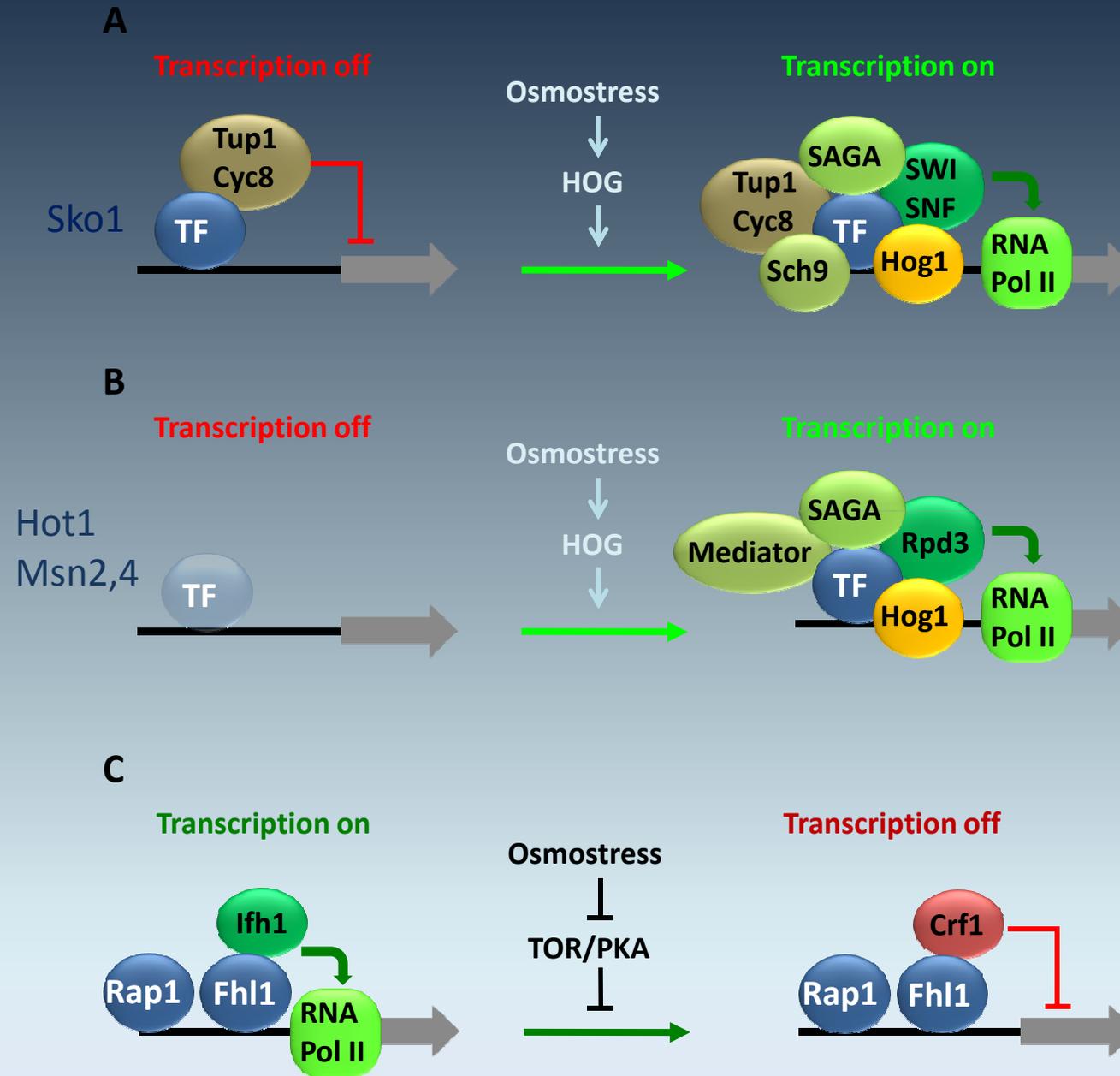
Modulation of mitochondrial activity

# The transcriptional program upon osmostress in yeast



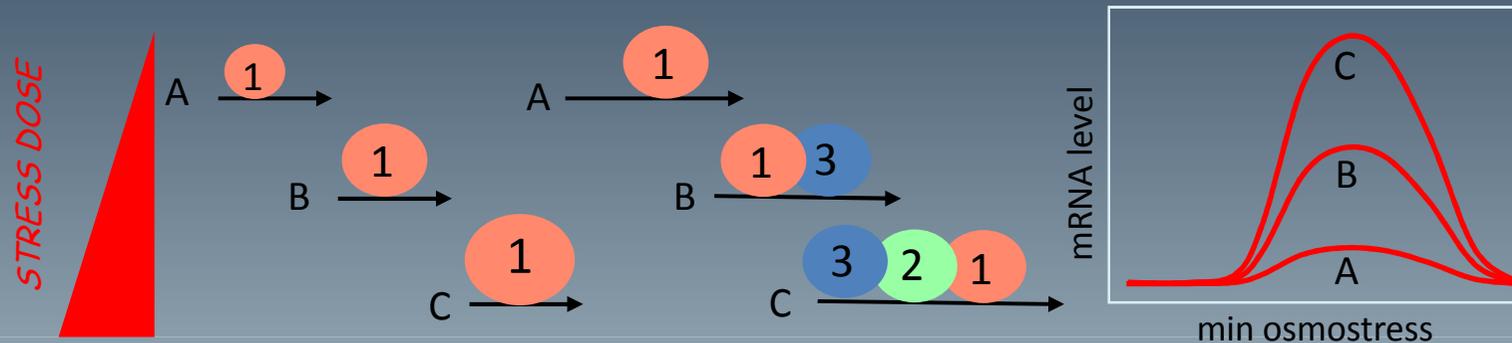
Transcriptional Profiling:  
Activated genes: 300-1400 (23%)  
Repressed genes: up to 1300

## Mechanisms of transcriptional control upon osmostress



## How is gene expression fine tuned during environmental changes?

*Modes of dynamically modulate transcription at different stress doses*



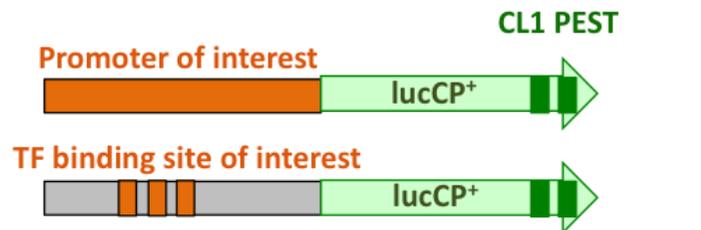
- How is promoter activity gradually regulated over a range of stress doses?*
- What are the molecular mechanisms which confer dose dependent promoter activity?*
- How does cell physiology change the dose sensitive gene expression?*

**We need:** *A continuous, real time transcription assay in the living cell adaptable to multiple stress doses*

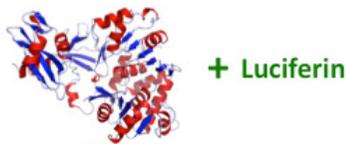
# The use of a real time luciferase assay to monitor promoter dynamics in the living yeast cell

Destabilized firefly luciferase (<15min half life in *S. cerevisiae*)  
Measurements in small culture aliquots in multiwell setup allow parallel analyses upon many environmental conditions - DOSE-RESPONSE for a given promoter  
We can obtain a quantitative value the  $EC_{50}$ .

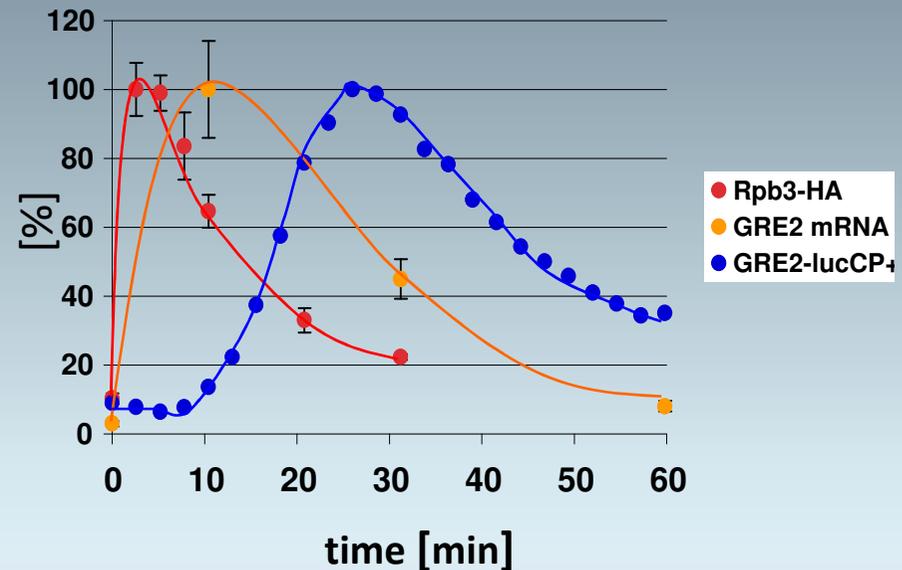
Expression of destabilized luciferase from single copy reporter fusions:



Highly unstable Luciferase  
(Half life =15min)

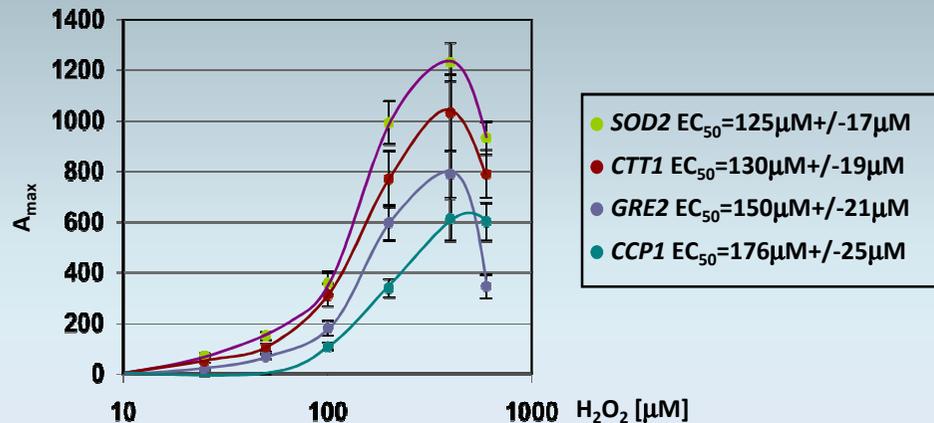
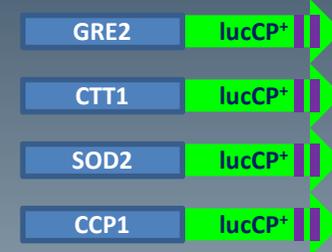
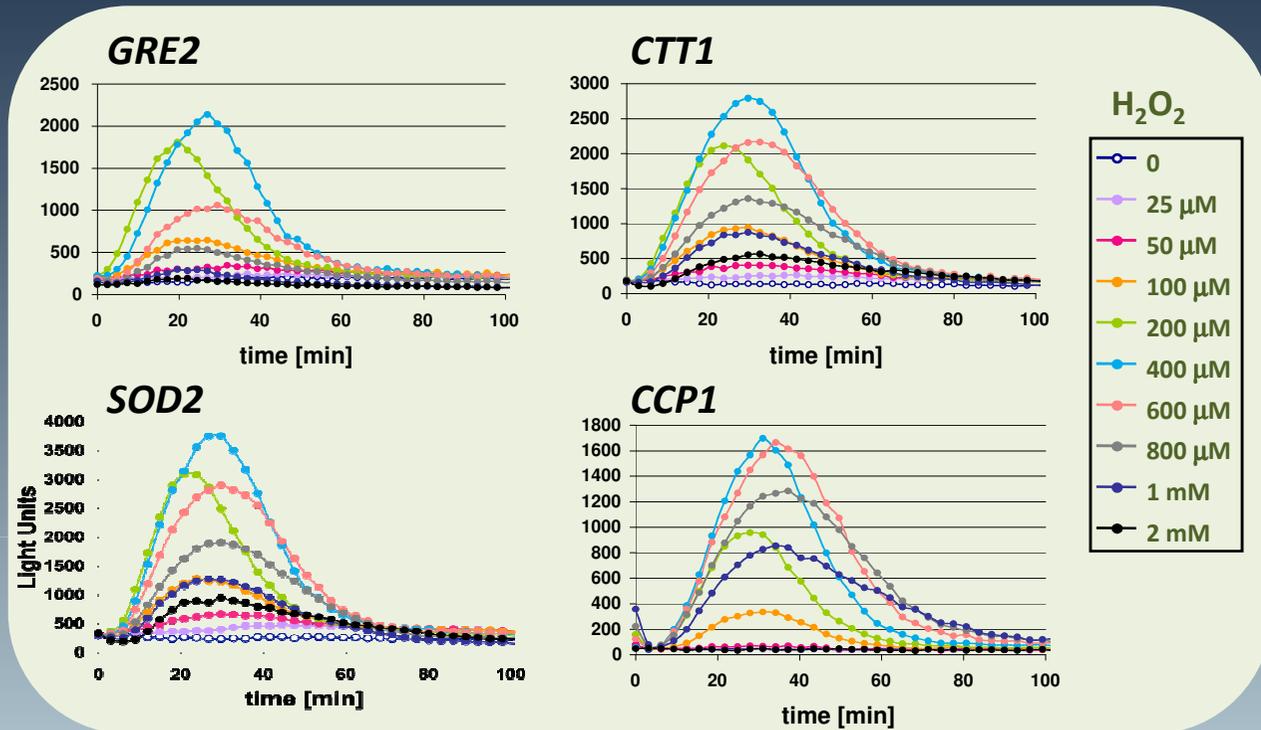


Real time quantification of light production from small aliquots of yeast cultures in a luminometer. Allows to record dynamic gene expression profiles upon many different environmental stimuli simultaneously



*GRE2* induction by 0.3M NaCl

# Quantitative Analysis of Natural Promoters upon Oxidative Stress



*Dose-Response behavior of the GRE2, CTT1, SOD2 and CCP1 promoters*

*SOD2 and CTT1 activities are especially sensitive to hydrogen peroxide stress*

# Quantitative Analysis of cis elements upon Oxidative Stress

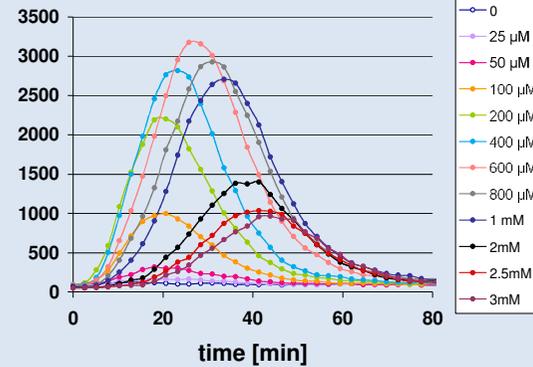
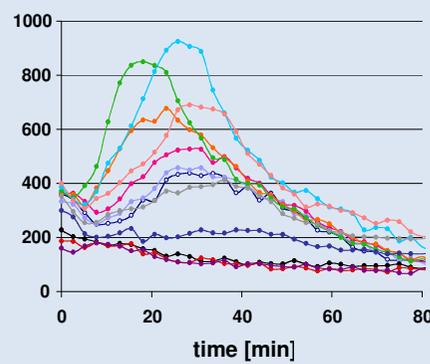
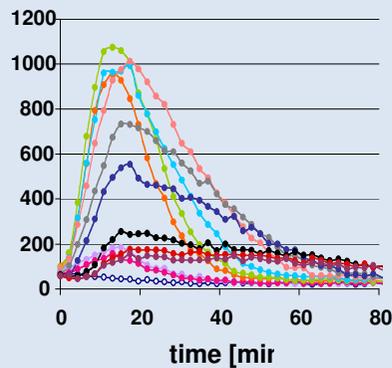
Synthetic Reporter Genes: 3x[TF binding site]-lucCP+



Osmostress → HOG → Sko1  
↓  
CRE

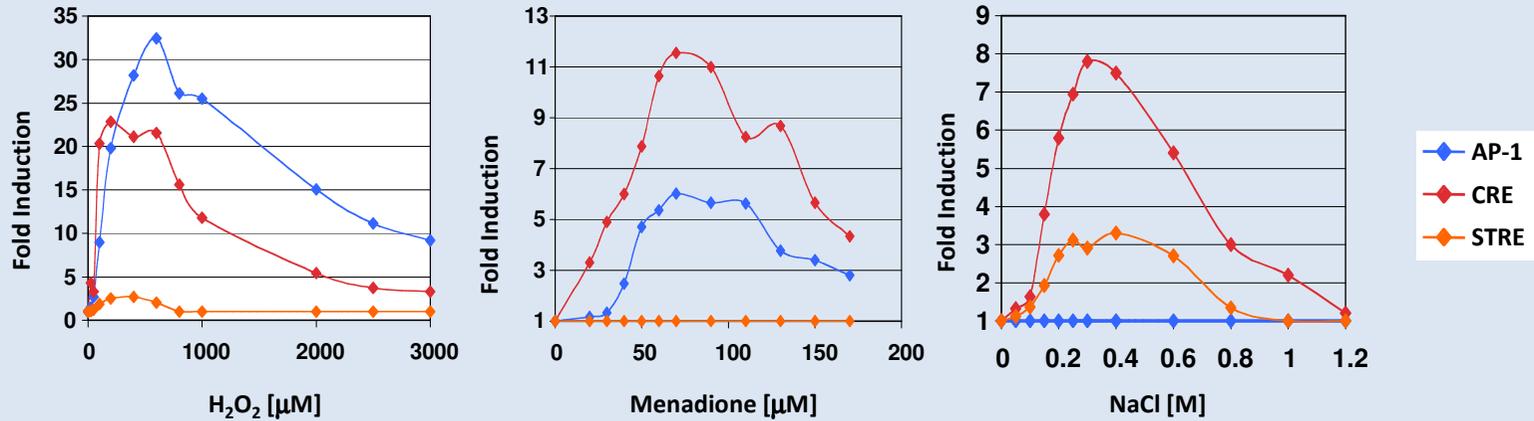
Stress → Msn2, 4  
↓  
STRE

Yap1 ← Oxidative Stress  
↓  
AP-1

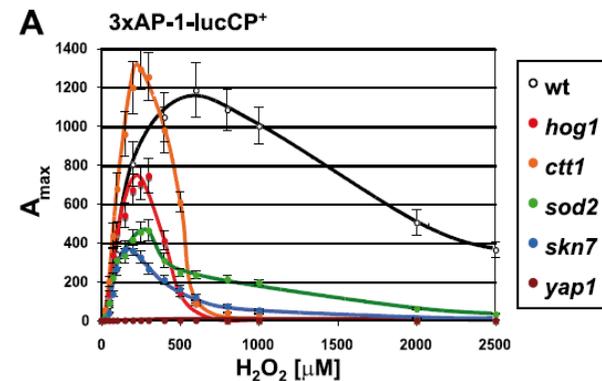


H<sub>2</sub>O<sub>2</sub>

Dose Response for each element under all stress conditions  
(NaCl, H<sub>2</sub>O<sub>2</sub>, Menadione)

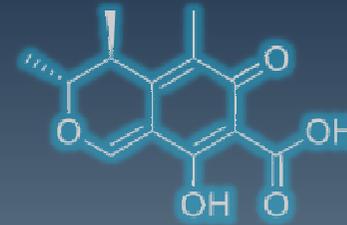


Cell physiology modulates the dose response of stress-activated gene expression

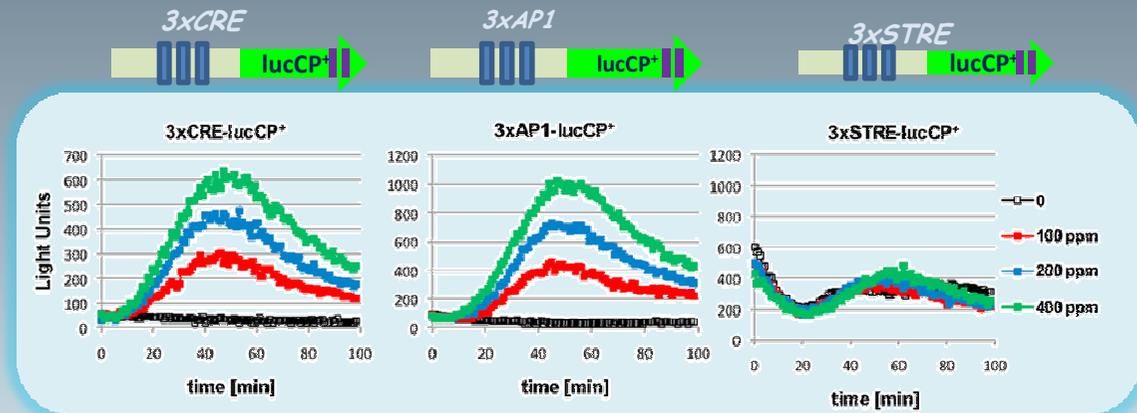
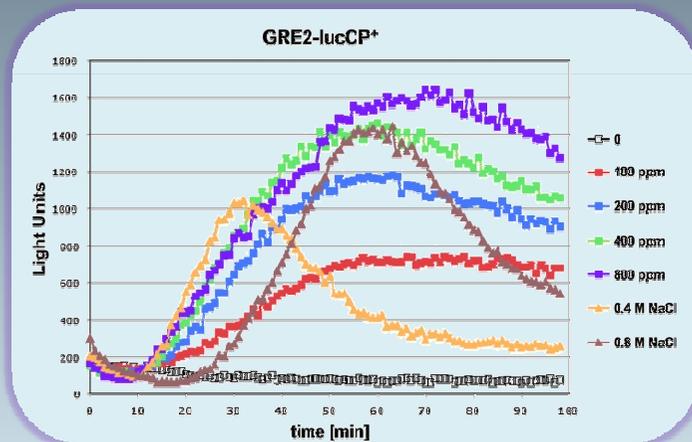


# Application of quantitative yeast model to study mycotoxins

Toxicological study of citrinin.



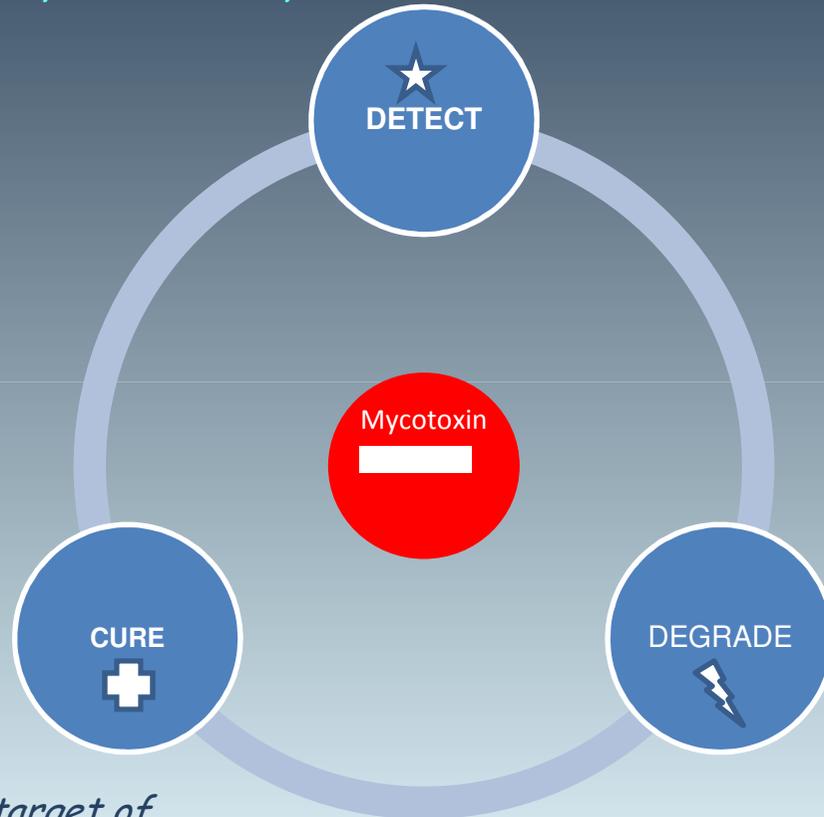
Citrinin triggers an immediate response to oxidative stress characterized by a strong and dose-dependent induction of natural genes.



At present we are using the "dose-response assays" to study the transcriptional behavior of other stress response promoters (Oxidative, DNA Damage, Mitochondrial dysfunction, cell cycle)

## Current projects:

*-Generate artificial promoters, with optimized sensitivity and specificity to one particular mycotoxin .*



*-Identify the molecular target of toxicity of citrinin. Expand the dose-response assays to study the toxicological effect of Ochratoxin A.*

*-Identify new enzymes for biodegradation .*



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*Sonia Squeo*