## **RNA-seq analysis of** *Macrobrachium rosenbergii* **hepatopancreas** MALAYA

# in response to Vibrio parahaemolyticus infection

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### INTRODUCTION

The Malaysian giant freshwater prawn, Macrobrachium rosenbergii, an economically important crustacean worldwide are being affected by Vibriosis, a disease caused by Vibrio strains such as Vibrio parahaemolyticus. M. rosenbergii possesses an innate immune system which provides defence against pathogenic agents. The information regarding the regulation of innate immune system in this species is lacking, thus its necessary in providing solutions to control and minimize the loss of production due to this bacterial disease. In this study, we performed a transcriptome profiling of *M. rosenbergii* hepatopancreas infected with *V. parahaemolyticus* using the 'Next Generation' sequencing method (Illumina HiSeq<sup>TM</sup>2000). A total of 54,295,342 and 54,708,014 high-quality reads obtained from Vibrio-infected and control M. rosenbergii cDNA libraries. The overall de novo assembly and clustering of both reads generated 64,411 unigenes, with an average length of 698 bp. Based on BLASTX search (E-value <10<sup>-5</sup>) against NR, Swissprot, GO, COG and KEGG databases, 22,455 unigenes (34.86% of all unigenes) were annotated with gene descriptions, gene ontology terms, and metabolic pathways. The unigene differential expression analysis revealed 14,569 unigenes were differentially expressed in the infected shrimp compared to the controls. Several differentially expressed genes are involved in various animal immune functions. The large number of transcripts obtained in this study would provide valuable resources for further genomic research into freshwater prawns.

### **OBJECTIVES**

•To perform a transcriptome profiling of the M. infected rosenbergii hepatopancreas with using Illumina HiSeq™ 2000 V.parahaemolyticus, platform.

To discover and determine the role of the immune genes involved in *V.parahaemolyticus* infection.

	Control	V.parahaemolyticus				A: RNA processing and modification
		Infected				B: Chromatin structure and dynamics
			3500 —		_	C: Energy production and conversion
Total number of reads	54,708,014	54,295,342				D: Cell cycle control, cell division, chromosome partitioning
						E: Amino acid transport and metabolism
Total base pairs (bp)	4,923,721,260	4,886,580,780	3000 -			F: Nucleotide transport and metabolism
Q20 value	97.73%	97.77%				G: Carbohydrate transport and metabolism
	+					H: Coenzyme transport and metabolism
Total number of contigs	95,645	123,141	2500 —			1: Lipid transport and metabolism
Mean length of contigs	313	318	s e			J: Translation, ribosomal structure and biogenesis
						K: Transcription
Total number of unigenes	59,050	73,946	5 2000 -			L: Replication, recombination and repair
Mean length of unigenes	479	532	0	-		M: Cell wall/membrane/envelope biogenesis
Mean length of ungenes	475	552	Ê			N: Cell motility
NCBI Nr annotated	19,	799	2 1500 -			O: Posttranslational modification, protein turnover, chaperones
Swiss-Prot annotated	16,	832				P: Inorganic ion transport and metabolism
	-					Q: Secondary metabolites biosynthesis, transport and catabolism
KEGG annotated	14,	706	1000 -			R: General function prediction only
COG annotated	7,8	356				S: Function unknown
GO annotated	6,0	007	500 -			T: Signal transduction mechanisms

RESULTS

#### **MATERIALS & METHODS**

- Challenge test: One *V.parahaemolyticus* challenge group and one negative control group (10 prawns each group) prepared
- At 12 hours post-infection, the hepatopancreas tissues of prawns were dissected and immediately frozen in liquid nitrogen
- **Total RNA extraction**

Wet lab

NGS

- Transcriptome sequencing using the Illumina HiSeq<sup>™</sup> 2000, BGI Shenzhen, China
- *De novo* assembly using the Trinity program
- Clustering of both unigenes done using TIGR Gene Indices clustering tools (TGICL)
- Annotation using BLASTX against NCBI Nr, Swissprot, COG and KEGG
- Gene Ontology (GO) assignment conducted using BLAST2GO software
- · Differential gene expression analysis done using the FPKM approach

#### DISCUSSION/ CONCLUSION

Table 1. Summary of the control and infected transcriptome sequencing



Figure 2. Gene ontology (GO) classification of the 6,007 protein annotated unigenes. Unigenes sequences were systematically classified into GO sub-categories under the Biological Process, Cellular Component and Molecular Function Gene Ontology Catalogue system. Each bar represents the relative abundance of unigenes classified under each sub-category

Category or gene id	Homologues function	Species	FC*
Antimicrobial			
Unigene4120_All	Anti-lipopolysaccharide factor	Macrobrachium rosenbergii	4.13
Unigene37309_All	Crustin	Macrobrachium rosenbergii	5.13
Blood Clotting system			
Unigene13048_All	Clottable protein	Marsupenaeus japonicus	1.42
Unigene34308_All	Transglutaminase	Macrobrachium rosenbergii	11.34
PRPs			
Unigene10978_All	Lectin 1	Macrobrachium rosenbergii	4.4
Unigene23671_All	lipopolysaccharide and beta-1,3-glucan binding protein	Macrobrachium rosenbergii	1.7
Proteinases and Proteinases inhibitors			



Figure 1. Histogram presentation of Cluster of Orthologus Groups (COG) classification of 7,856 known protein annotated unigenes. Each bar represents the number of unigenes classified into each of the 25 COG functional categories



Figure 3. Digital gene expression between control group against V.parahaemolyticus infected group. Each point represents a unigene. The x- and y-axis are the log10 of the normalized expression level (FPKM) of unigene between the two groups. Red and green points indicate significant change at the absolute value of log<sub>2</sub> (FPKM ratio in two groups) ≥1 and fdr =0.001. Red points indicate up-regulated unigenes and green points indicate down-regulated unigenes in the two groups which its expression level is represented by the y-axis. Blue points indicate insignificant differentially expressed unigenes

This study generates significant information on M.rosenbergii immune genes activity during *V.parahaemolyticus* infection.

Several differentially expressed genes identified are involved in various animal immune functions, such as antimicrobial, proteases and protease inhibitors, pattern recognition proteins, heat shock proteins, cell death, oxidative stress, blood clotting system, and prophenoloxidase system.

Results obtained provided a valuable insight into antibacterial mechanisms in *M.rosenbergii* and the role of the differentially expressed immune genes in response to V. parahaemolyticus infection.

#### Alpha-2-macroglobulin Unigene12869\_All Macrobrachium rosenbergii 5.09 CL2365.Contig1 All Caspase 1.04 Marsupenaeus japonicus Unigene26757\_All Hemocyte kazal-type proteinase inhibitor Penaeus monodon 3.87 CL1127.Contig2\_All -6.01 Kazal-type serine proteinase inhibitor 4 Procambarus clarkii Heat shock Proteins Unigene3736\_All Heat shock protein 21 4.61 Macrobrachium rosenbergii Unigene23034\_All Heat shock protein 40 Frankliniella occidentalis 1.8 Oxidative stress 1.01 CL353.Contig2\_All Glutathione S transferase Procambarus clarkii CL2477.Contig1 Catalase Litopenaeus vanname 1.6 ProPO system Unigene12734\_All Prophenoloxidase Macrobrachium rosenbergii 3.15 Unigene7353\_All Prophenoloxidase activating factor 3.79 Fenneropenaeus chinensis Cell death Unigene2555\_All Beclin 1 Megachile rotundata 11.89 Unigene14045\_All 1.61 Program cell death 5-like Penaeus monodon

Table 2. Selected candidate genes involved in M.rosenbergii immune response. \*Fold changes (Log<sub>2</sub> ratio) in gene expression, PRPs- pattern recognition proteins, ProPO- prophenoloxidase



Figure 4. Top 20 KEGG biological pathway classification histograms for annotated unigenes

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